



Targeted nanodelivery systems for personalized cancer therapy

Szymon Roszkowski¹, Zofia Durczyńska², Sylwia Szablewska²

¹Division of Biochemistry and Biogerontology, Collegium Medicum, Nicolaus Copernicus University, Bydgoszcz, Poland

²Department of Oncology, Collegium Medicum, Nicolaus Copernicus University, Bydgoszcz, Poland

ABSTRACT

Conventional cancer therapies such as chemotherapy face challenges such as poor tumor targeting, systemic toxicity, and drug resistance. Nanotechnology offers solutions through advanced drug delivery systems that preferentially accumulate in tumors while avoiding healthy tissues. Recent innovations have enabled the optimization of engineered nanocarriers for extended circulation and tumor localization via both passive and active targeting mechanisms. Passive accumulation exploits the leaky vasculature of tumors, whereas active strategies use ligands to selectively bind cancer cell receptors. Multifunctional nanoparticles also allow the combination of imaging, multiple therapeutic modalities and on-demand drug release within a single platform. Overall, precisely tailored nanotherapeutics that leverage unique pathophysiological traits of malignancies provide opportunities to overcome the limitations of traditional treatment regimens. This emerging field promises more effective and personalized nanomedicine approaches to detect and treat cancer.

The key aspects highlighted in this review include the biological barriers associated with nanoparticles, rational design principles to optimize nanocarrier pharmacokinetics and tumor uptake, passive and active targeting strategies, multifunctionality, and reversal of multidrug resistance.

Key words: nanotherapy; drug delivery; cancer targeting; tumor microenvironment; personalized medicine

Rep Pract Oncol Radiother 2024;29(6):776–788

Introduction

Cancer remains one of the leading causes of death worldwide, prompting intense research into more effective and tailored therapeutic approaches. Conventional cancer treatments such as chemical treatment have limited selectivity, resulting in poor bioavailability at tumor sites and systemic toxicity [1, 2]. The emerging field of nanomedicine offers promising solutions through advanced nanoengineering and nanodelivery systems. Recent innovations in nanotechnology have enabled the development of nanoscale platforms that can preferentially

accumulate in tumors while avoiding healthy tissues. By leveraging the unique pathophysiological traits of malignancies, nanotherapeutics provide opportunities to overcome the limitations of traditional chemotherapy.

Recent advances in nanotechnology have led to the engineering of diverse nanoplatforms that can preferentially accumulate in tumor tissues while reducing exposure to healthy cells. This tumor-targeting ability is achieved by leveraging unique pathophysiological traits of malignancies. For example, tumors typically have leaky vasculature and impaired lymphatic drainage, allowing nanocarriers

Address for correspondence: Szymon Roszkowski, Nicolaus Copernicus University, Collegium Medicum, Division of Biochemistry and Biogerontology, Bydgoszcz, Poland; e-mail: szymonr@cm.umk.pl

This article is available in open access under Creative Common Attribution-Non-Commercial-No Derivatives 4.0 International (CC BY-NC-ND 4.0) license, allowing to download articles and share them with others as long as they credit the authors and the publisher, but without permission to change them in any way or use them commercially

of certain sizes to selectively extravasate from blood vessels and penetrate into the tumor microenvironment. This phenomenon is known as the EPR effect [3]. NPs ranging from 10–100 nm in diameter have been shown to effectively exploit the EPR effect for passive tumor targeting after intravenous administration [4].

Additionally, the tumor microenvironment has abnormal physicochemical properties, such as an acidic pH and high reducing capacity [5]. pH- or redox-responsive nanocarriers can be designed to release their drug payload specifically when exposed to these intracellular tumor conditions [6]. Active targeting strategies further enhance selectivity by functionalizing the nanoparticle surface with targeting ligands [7]. These ligands bind to receptors or antigens that are overexpressed on cancer cells compared to those on normal tissues [8]. Some examples include folate receptors, transferrin receptors and lectins [9]. This molecular recognition guides nanotherapeutics directly to cancer cells while avoiding healthy cells lacking these biomarkers [10].

Overall, nanotechnology provides multiple opportunities to overcome the nonselective biodistribution and severe side effects associated with conventional chemotherapies. Engineering nanocarriers that leverage unique aspects of tumor pathophysiology enables more personalized and precise drug delivery. Both passive and active targeting mechanisms significantly improve nanoparticle accumulation in tumors compared to normal tissues [11]. This approach promises to increase treatment efficacy while reducing systemic toxicity. By developing nanotherapeutics customized to cancer traits at the molecular level, nanomedicine provides promising opportunities for advancing cancer treatment [12].

This review discusses the progress and promise of nanobased targeting strategies for cancer therapy.

Nanobased targeting strategies for cancer therapy

First-generation chemotherapeutics are distributed nonspecifically throughout the body, leading to only a small fraction of the administered dose being localized in the tumor [13]. NPs in the size range of 10–100 nm can exploit leaky tumor vas-

culature to accumulate passively via the EPR effect [14]. Liposomes, polymers, dendrimers, micelles and other nanocarriers have been optimized for extended circulation and tumor localization based on their size, charge and surface chemistry [15]. Ligand-mediated active targeting further improves nanoparticle uptake in cancer cells by facilitating the molecular recognition of overexpressed receptors [16]. Examples of these agents include transferrin, folate and aptamers, which are biomarkers that are upregulated in malignancies [17]. Compared with conventional chemotherapies, combined passive and active strategies enable more precise and personalized delivery.

Another major obstacle addressed by nanocarriers is the poor aqueous solubility of hydrophobic drugs such as paclitaxel [18]. One successful example of protein NPs already used in clinical practice is albumin-bound paclitaxel NPs, which are sold under the name Abraxane [19]. This drug was obtained by high-pressure homogenization of the drug and a bovine albumin solution, resulting in NPs approximately 130 nm in size that can be easily administered intravenously. As shown, Abraxane production can be easily scaled up to industrial levels without loss of stability or therapeutic activity [20]. Therefore, methods such as simple pressure homogenization used in Abraxane represent a promising strategy for the development of other albumin-based formulations. Nevertheless, there is still a need to optimize advanced protein nanostructures for pharmacokinetic properties and drug release profiles.

Encapsulation of drugs with poor solubility in nanoplatforms or conjugation with nanoparticle matrices improves their solubility and enables parenteral administration [21]. Moreover, encapsulation increases the stability of therapeutic drugs, protecting them against enzymatic degradation and the influence of unfavourable pH conditions or ionic forces present in the body [2].

Multifunctionality is another key advantage of NPs. Theranostic nanosystems integrate imaging agents, diagnostic modules, and triggered drug release mechanisms to provide real-time monitoring of therapy [22]. Stimulus-responsive strategies trigger precise drug release within the tumor microenvironment in response to conditions such as acidic pH or elevated oxidative stress levels. Such precise spatiotemporal control over nanobased delivery has

the potential to substantially improve patient outcomes through more targeted therapy while reducing adverse effects. For example, polymeric micelles were engineered to stabilize drug cargo at physiological pH while allowing stimulus-mediated release under acidic tumor microenvironment conditions. *In vitro* and *in vivo* studies have demonstrated that these micelles exhibit desirable effects, including intracellular pH-responsive drug release, infiltration into tumor tissue, and potent antitumour efficacy with minimal toxicity [23].

Biological barriers that NPs can help overcome

The human body contains several defensive barriers that impede the delivery of therapeutics to target sites. NPs have shown promise in overcoming these obstacles [24].

Intracellular delivery

The cell membrane acts as a selectively permeable barrier, limiting the uptake of exogenous materials such as therapeutic nucleic acids, proteins, and drug molecules into cells. Overcoming this delivery challenge is crucial for medical applications relying on bioactive intracellular agents. Cationic nanocarriers can facilitate intracellular access through electrostatic association with negatively charged biomacromolecules such as DNA, coupled with cell entry via endocytosis and membrane destabilization.

For example, cationic lipids and polymers have been extensively utilized to enhance the intracellular delivery of nucleotide therapeutics. A recent study demonstrated that aptamer-functionalized NPs effectively deliver PD-L1 siRNA to triple-negative breast cancer cells, resulting in almost complete suppression of PD-L1 expression within 90 minutes of treatment [25]. The NPs also displayed minimal systemic toxicity *in vivo*. Similarly, lipid-based NPs can effectively bind lapatinib and anti-survivin siRNA for HER2+ breast cancer treatment, potentially enhancing their anticancer activity [26].

Other biomolecular therapeutics have also benefited from cationic nanocarrier-mediated delivery approaches. Recently, Lipid NPs were shown to efficiently deliver gene-editing proteins across the intestinal epithelial layer, proving useful for potential oral drug delivery [27].

Likewise, cationic micelles with shielding polymers can reduce cytotoxicity and maintain cell viability for nonviral gene delivery, offering high potential for *in vivo* applications [28].

Overall, cationic nanosystems have demonstrated significant potential to overcome cell membrane barriers and enable effective intracellular therapeutic delivery through charge-mediated interactions and endocytic internalization. Careful carrier engineering to optimize physicochemical and biological properties can further improve delivery outcomes in diverse biomedical applications [29].

Delivery across epithelial barriers

Epithelial tissues found in the gastrointestinal tract, lungs, kidneys and other organs form highly selective permeation barriers essential for proper physiological functioning. Tight junctions between adjacent epithelial cells strongly limit the passive transport of exogenous substances due to their extreme impermeability [30]. Orally administered drugs face additional obstacles, including enzymatic degradation in the stomach and poor intestinal solubility. Nanoparticle carriers can overcome several of these delivery challenges to enhance therapeutic uptake across mucosae.

NPs can encapsulate labile drugs, protecting them from harsh conditions in the gastrointestinal environment [31]. The nanoparticle surface can also be functionalized with tight junction modulators to transiently breach paracellular pathways [32]. Alternatively, nanoparticle size (~100 nm) and surface properties may be tailored to promote cell-mediated active transport via transcytosis [33]. Through such mechanisms, NPs increase therapeutic absorption by the intestinal epithelium following oral delivery.

Similarly, for pulmonary delivery, NPs preserve sensitive biomolecular components from airway clearance mechanisms and affiliated enzymes. Cationic NPs strongly interact with negatively charged lung epithelia, triggering caveolae/clathrin-mediated endocytosis and transcellular migration [34]. As such, NPs significantly intensify the transport efficacy of respiratory therapeutics compared with free drugs. Appropriately engineered NPs thus promote delivery across diverse epithelial barriers, advancing oral, nasal and inhalational pharmacotherapy.

Delivery within the tumor microenvironment

Tumor biology limits therapeutic efficacy through multiple mechanisms. Angiogenesis spurs chaotic blood vessel development, impairing drug perfusion. The resulting high interstitial fluid pressure further impedes nanoparticle penetration into the tumor core [35]. Hypoxic and acidic tumor zones resist both chemo- and radiotherapy due to diminished mechanisms of cell death [36]. Overcoming these complex transport and physiological barriers constitutes a major goal in drug delivery science.

The enhanced permeability and retention (EPR) effect provides one strategy to improve tumor-tropic nanoparticle delivery. Aberrant capillary fenestrations permit tumor nanoparticle accumulation, a phenomenon further enhanced by the absence of functional lymphatic vessels [37]. However, deeper tumor penetration necessitates additional targeting and stimulus responsiveness [38]. Multistage NPs undergo stepwise size alteration to migrate through narrow tumor labyrinths. For example, protonation in acidic hypoxic regions triggers polymer expansion, facilitating the release of inner drug payloads [39]. Overall, nanovehicles enable modular solutions to meet the challenges presented by the hostile tumor microenvironment.

Delivery to target immune cells

However, targeting NPs to specific subsets of immune cells is difficult but necessary for modulating immunity. NPs with functional groups activating immune cells in lymph nodes have been designed [40]. Additionally, antibody-coated NPs recognize antigens on the surface of target cells. For example, polymer NPs coated with antibodies directed against CD40, DEC-205 and CD11c receptors on dendritic cells showed increased antigen uptake and the ability to stimulate T cells compared to NPs without such targeting [41]. Similarly, lipid NPs with fragments of antibodies against T-cell antigens allowed for selective labelling and stimulation of these cells after administration to the body [42].

Recently, approaches have focused on delivering NPs to dendritic cells, which are key antigen-presenting cells. For example, manose-modified NPs show increased uptake by dendritic cells via receptor-dependent endocytosis [43]. Lipid-calcium-phosphate NPs coated with single-chain antibodies also achieve selective targeting of dendritic cells in lymph nodes [44]. This leads to localized delivery of antigens and adjuvants, stimulating strong cytotoxic T-cell responses.

Thus, NPs can be used to deliver drugs or vaccines through surface groups that recognize antigens on immune cells to enable more precise and effective interactions with specific types of

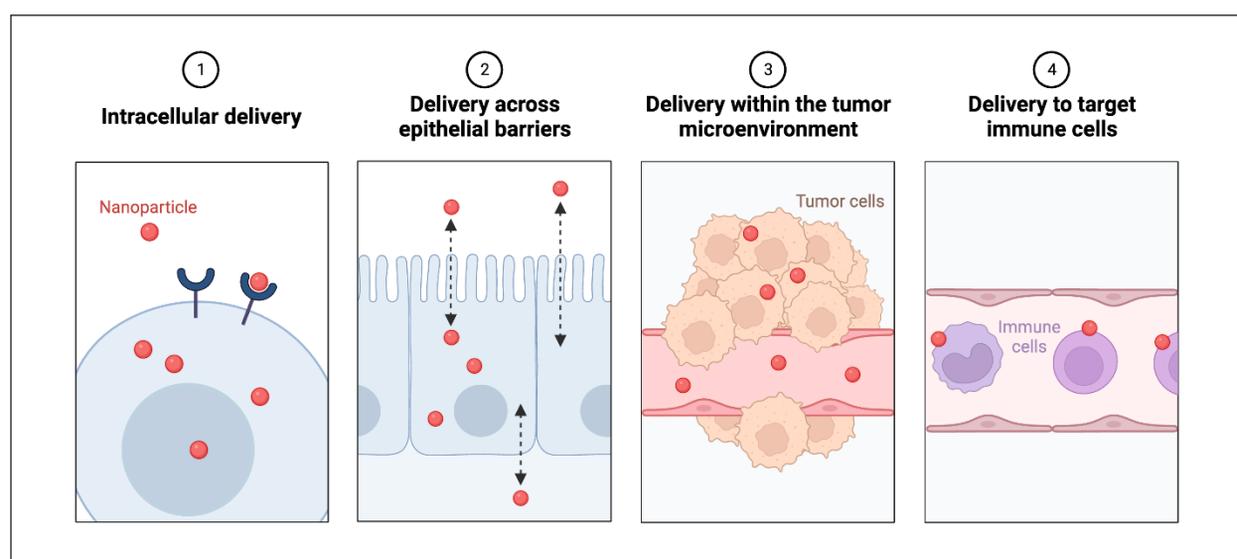


Figure 1. Biological barriers that nanoparticles can help overcome

leukocytes. The use of such targeted nanocarriers improves the delivery of therapy to specific effector cells and increases immune responses against target antigens.

In summary, the modular design and tunable properties of NPs make them extremely adaptable delivery vehicles capable of penetrating various biological barriers and accessing hard-to-reach targets.

Precisely engineered nanocarriers for optimized tumor targeting

The selective delivery of chemotherapeutic agents to tumor tissues while avoiding healthy organs is a major challenge in cancer therapy. Nanocarriers have emerged as a promising solution for optimizing drug accumulation in the tumor area through the enhanced permeation and retention (EPR) effect. This effect allows NPs to selectively extravasate through the abnormal leaky vasculature of the tumor and accumulate there. However, the EPR effect is heterogeneous among patients and tumor types, limiting its reliability for precise drug targeting [45]. Therefore, rationally designing nanocarriers with optimized sizes, shapes, and surface properties is essential for enhancing tumor selectivity.

Size and shape modulation

The size and shape of nanocarriers directly impact their circulation time and penetration through tumor tissues. It has been shown that the nanoparticle size should be approximately 100 nm to benefit from both extended blood circulation times through reduced renal clearance and good tumor penetration [46]. Furthermore, compared with spheres, nonspherical nanocarriers, such as nanorods, have demonstrated enhanced tumor targeting owing to their superior margination toward vessel walls and improved diffusion inside tumor tissues [47].

Surface functionalization

Actively targeted nanocarriers can more precisely deliver their drug payload to cancer cells by exploiting ligand-receptor interactions. Specific receptors tend to be overexpressed on the membranes of cancer cells compared to healthy cells. Thus, decorating nanocarriers with ligands that bind these

receptors allows preferential uptake into tumor tissues via receptor-mediated endocytosis [48].

A variety of ligands, including peptides, antibodies, aptamers and small molecules, have been investigated for the active targeting of nanocarriers [49]. These ligands bind to receptors such as transferrin, folate, epidermal growth factor and interleukin receptors, which are commonly upregulated in cancer cells. Receptor-ligand binding triggers endocytosis of the nanocarrier, directing it into endosomes and lysosomes, where the encapsulated drugs are released. This leads to enhanced intracellular drug accumulation and cytotoxicity in cancer cells [50].

Moreover, since ligand-receptor binding is saturable, drug uptake can be optimized by tuning the density of ligands on the nanocarrier surface. Multivalent nanocarriers with multiple copies of targeting ligands have been shown to augment the targeting specificity and further improve drug delivery [51].

A combination of passive and active targeting strategies is designed to maximize selectivity through enhanced permeability, optimized particle geometry, and specific cancer cell recognition. Tailoring NPs by leveraging cancer pathophysiology and molecular profiles significantly improves specificity compared to untargeted vehicles and conventional chemotherapies [52]. These advances have brought the field closer to realizing truly targeted nanotherapeutics.

Multifunctional nanosystems for cancer therapy

Multifunctional nanosystems are promising platforms for cancer diagnosis and therapy. They combine multiple functions, such as the detection of cancer cells, drug delivery, photodynamic therapy and gene therapy [53]. NPs are modified with appropriate ligands to target their action on cancer cells and elements of the tumor microenvironment [54].

One example is gold NPs coated with folic acid and monoclonal antibodies directed against the HER2 receptor [55]. This allows these nanosystems to selectively bind to breast cancer cells and subsequently release the drugs they contain [56]. Additionally, gold NPs generate heat under infrared radiation, causing hyperthermia and leading

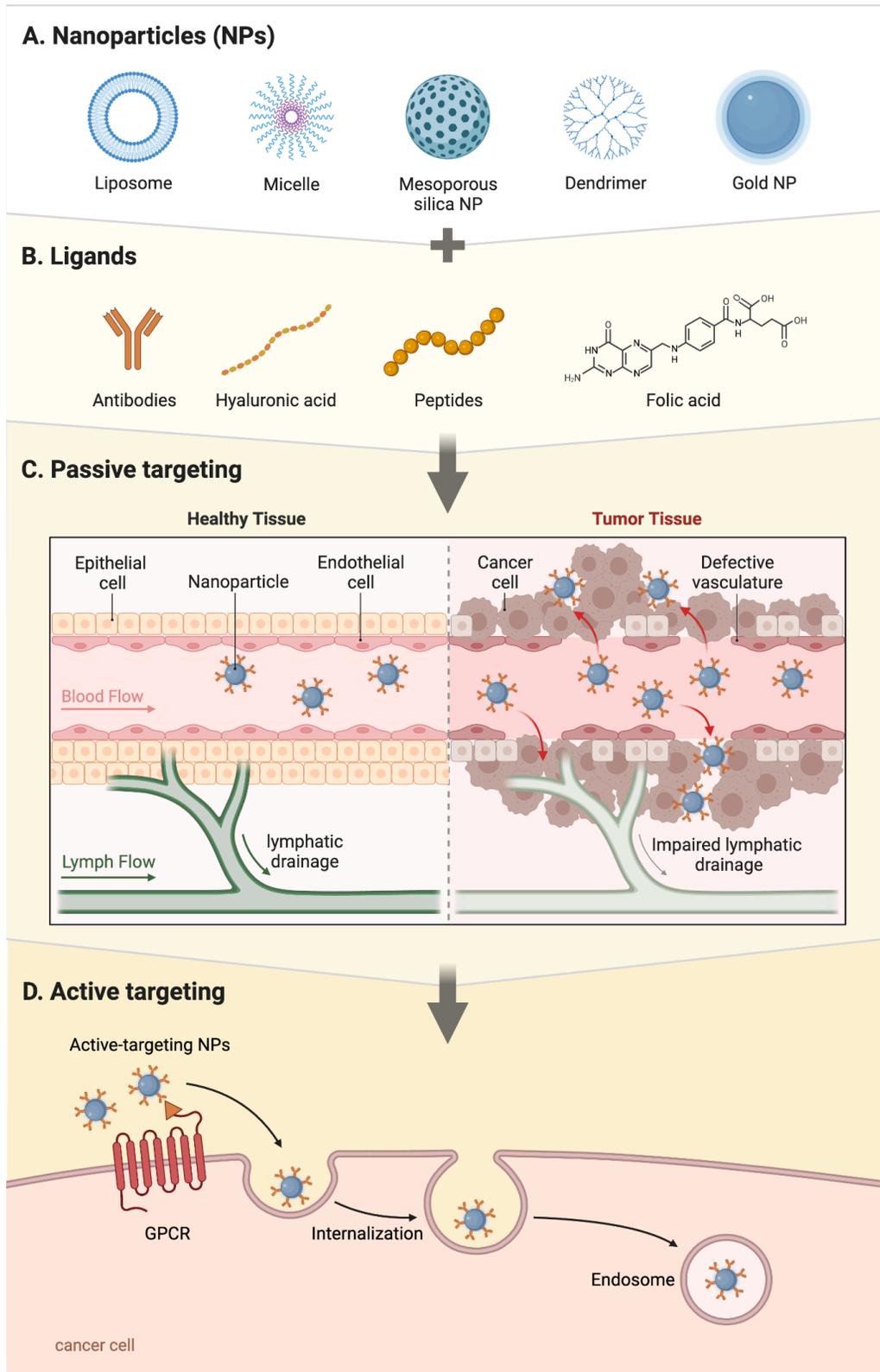


Figure 2. active and passive targeting of nanoparticles to cancer cells. **A.** Nanoparticles (NPs) — NPs are shown as small, rounded objects that can transport drugs or other therapeutic agents; **B.** Ligands — ligands are shown as small molecules attached to the nanoparticle surface that can recognize and bind to receptors on cancer cell surfaces; **C.** Passive targeting — NPs accumulate in the tumor via the enhanced permeability and retention effect. They do not have targeting ligands; **D.** Active targeting — NPs have ligands on their surface that selectively bind to receptors on cancer cells. This leads to enhanced nanoparticle accumulation in the tumor

to the destruction of cancer cells [57]. Therefore, these NPs have two functions — they deliver drugs and induce hyperthermia.

Another type of promising nanocarrier is mesoporous silica NPs, which, in addition to drug delivery, can be surface modified to obtain additional diagnostic functionalities. For example, by introducing iron oxide nuclei into the structure of these carriers, these NPs gain superparamagnetic properties that enable their use as contrast agents in magnetic resonance imaging [58].

Moreover, the surface of silica NPs can be modified with fluorescent probes whose signal depends on the local pH. Owing to this approach, these nanosystems can be used not only for monitoring drug release but also for imaging the tumor micro-environment and assessing therapeutic response [59].

A separate class of promising nanocarriers for theranostic applications in oncology is made up of superparamagnetic iron oxide NPs (SPIONs). The magnetic core allows precise monitoring of biodistribution via magnetic resonance imaging methods. Moreover, through appropriate

surface modification, SPIONs can selectively deliver drugs to cancer cells and tumors [60]. These properties make superparamagnetic iron nanostructures attractive, multifunctional therapeutic and diagnostic platforms [61].

Here are some additional examples of multifunctional nanosystems used in cancer theranostics:

Cancer theranostics are currently using increasingly advanced nanoparticle systems that combine the possibilities of cancer diagnostics and therapy [62]. The main advantage of these materials is the integration of many functions in one nanostructure, which allows for the achievement of a synergistic effect and increased effectiveness of treatment [63].

One example of such systems is graphene NPs coated with platinum compounds and the fluorescent label nigrosin [64]. They can simultaneously detect cancer cells via fluorescence and destroy them by local tissue heating with graphene and platinum [65, 66].

Another type of multifunctional nanocarrier consists of mesoporous silica NPs with anticancer substances trapped in the pores [67]. They release

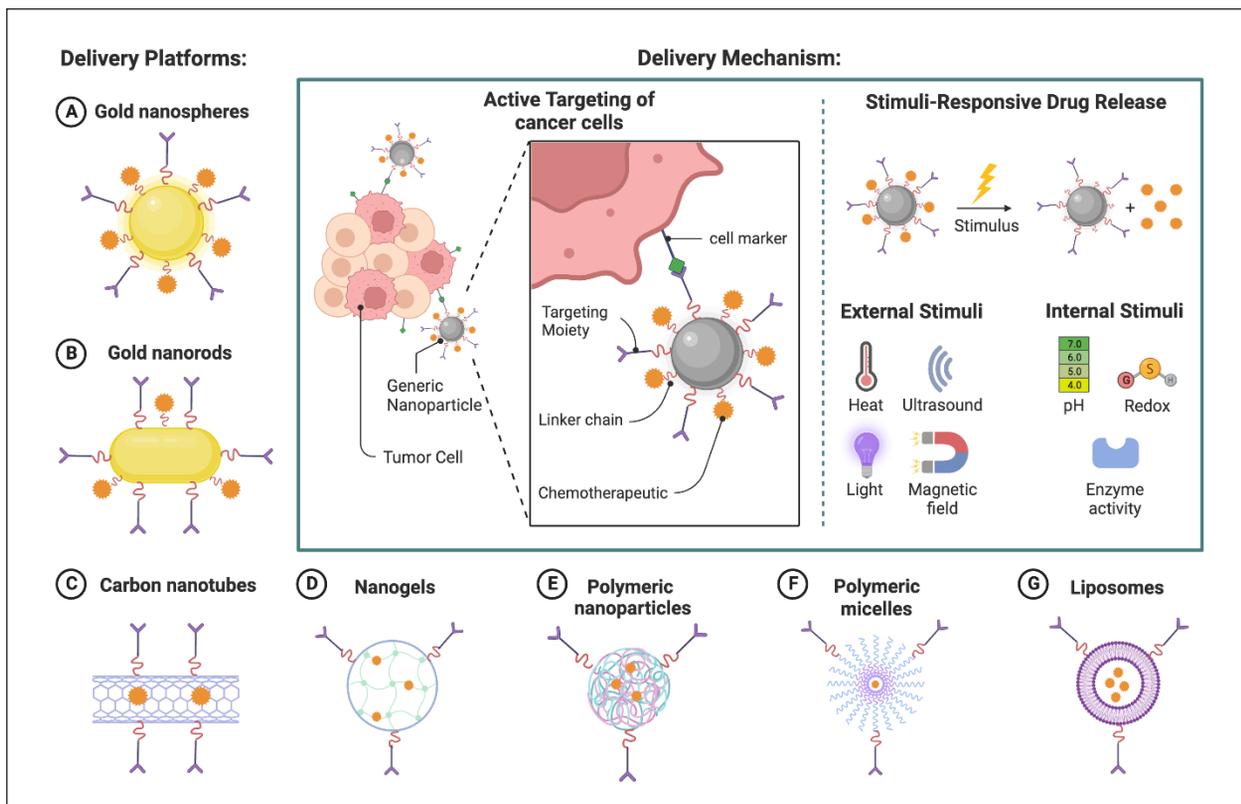


Figure 3. Nanoparticle-mediated targeted drug delivery to cancer cells

drugs gradually and directly into cancer cells [68]. Additionally, by adapting the surface of these NPs, they can be actively targeted to cancer tumors [69].

Another solution is to modify liposomes with hyaluronic acid, which facilitates their accumulation in tumors [70]. In addition to chemical drugs, these liposomes may contain contrast agents that enable magnetic resonance imaging [71]. This allows real-time monitoring of the distribution of liposomes containing the drug into the tumor.

The possibilities of cancer theranostics are also expanded by polymer NPs with specific surface ligands that direct them to cancer cells [72]. They can deliver interfering RNA molecules directly to these cells, enabling the silencing of selected genes involved in the progression of the disease [73].

In turn, after entering the cell, lipid NPs with recombinant fusion proteins release the embedded genetic material [74]. This process facilitates cancer gene therapy by providing factors that regulate gene expression or DNA editing complexes [75].

The integration of various diagnostic and treatment methods within single NPs significantly increases the effectiveness of oncological therapies [76]. Importantly, the properties of these nanosystems can be precisely tailored individually to the patient's profile, enabling a personalized approach to therapy [77]. Multifunctional nanotheranostics create new perspectives in cancer therapy.

Reversing multidrug resistance

Multidrug resistance (MDR) is the main cause of chemotherapeutic failure in cancer patients. The process involves the ability of cancer cells to actively remove various anticancer drugs from inside the cell, which significantly reduces their concentration and effectiveness [78]. However, there are several promising strategies for reversing MDR resistance in cancer cells.

Due to their unique physicochemical properties, NPs are a promising platform for drug delivery and overcoming MDR resistance in cancer cells [79]. They can be functionalized by adding appropriate ligands recognized by receptors on cancer cells to their surface [80]. This leads to active uptake of NPs from the circulation and targeted transport to the tumor [81].

Moreover, the electrical charge and hydrophobicity of NPs can be masked by the addition of

biopolymer or PEG coatings. This prevents their detection and removal by ABC transport pumps [82]. ABC transporters are proteins located in the cell membrane that use energy from ATP to actively transport various substrates across the membrane to the outside of the cell [83]. The family of ABC transporters includes P-glycoprotein (P-gp), breast cancer resistance protein (BCRP), and multidrug resistance protein (MRP) [84]. They are produced in excess in the membranes of cancer cells. They capture anticancer drugs from inside the cell and actively remove them from the cell [85]. This leads to a decrease in the intracellular concentration of these drugs, preventing them from achieving a therapeutic effect [86]. ABC transporters are therefore responsible for the development of MDR through the pumping out of chemotherapeutic drugs from cancer cells [87]. Their inhibition or bypass via NPs is a promising strategy for overcoming cancer resistance during treatment [88].

Another strategy is gene therapy, which involves introducing specific genes into cancer cells to combat MDR resistance [89]. The main challenge is the efficient and selective transfer of genetic material to cancer cells [90]. NPs provide an ideal carrier platform in this case [91]. They provide genes encoding enzymes that metabolize drugs, increasing their intracellular concentration, or genes that inhibit apoptosis [92]. This restores the sensitivity of cancer cells to treatment and reverses multidrug resistance by modulating key signalling pathways [93].

Another method is photodynamic cancer therapy (PDT), which involves the activation of a photosensitizer using light of a specific wavelength, which leads to the production of reactive oxygen species that destroy cancer cells. The main obstacle is the low solubility and nonselective distribution of photosensitizers in the body [94,95]. NPs can increase the effectiveness of PDT in several ways.

First, photosensitizers are immobilized, and their solubility is increased. Photosensitizing molecules often have low solubility in water, which makes them difficult to use. The use of polymers (PLGA) or lipid nanocarriers allows for increased solubility, improved release kinetics and modified distribution routes of these compounds from the bloodstream to tissues [96].

Another mechanism is to target PDT by functionalizing the surface of NPs with ligands rec-

ognized by receptors overexpressed on cancer cells, such as transferrin or folic acid. This leads to the selective uptake of nanocarriers from the circulation, mainly to the target tissue, reducing systemic toxicity [97].

NPs can also help photosensitizers overcome MDR mechanisms and accumulate in cancer cells by masking their charge or hydrophobicity, which prevents them from being pumped out by transport systems [98].

Conclusions

The presented literature review indicates that precisely targeted drug delivery systems using nanotechnology are a promising therapeutic strategy for cancer treatment, allowing us to overcome the limitations of conventional cytotoxic chemotherapy.

The developed nanotherapeutics can selectively accumulate in cancer tumors through enhanced permeability and retention (EPR) and functionalization of the surface of nanocarriers with ligands recognizing receptors overexpressed on cancer cells. This results in improved bioavailability of drugs at the target site while minimizing systemic toxicity.

Additionally, remarkable progress has been made in designing nanoplatfoms that exploit unique features of cancer pathophysiology for targeted transport of therapeutics. Both passive and active targeting strategies significantly improve nanoparticle accumulation in tumors compared to normal tissues.

The combined approaches further enhance selectivity through optimized particle geometry, surface functionalization, and cancer cell recognition. These advances have brought us closer to developing truly personalized nanomedicine.

Additionally, reversible modulation of multi-drug resistance in tumors using precisely designed therapeutic nanocarriers allows us to overcome the key limitations of conventional chemotherapy. This enables high concentrations of intracellular therapeutics to be achieved. Thus, owing to the precise adaptation of the structure to the specificity of the tumor microenvironment, nanotherapeutics can bypass the barriers that prevent the effectiveness of typical cytostatics.

Perspectives

In the future, continued innovations in nanocarrier designs and targeting mechanisms promise more precise spatiotemporal control over drug release in the tumor microenvironment. Stimulus-responsive and theranostic strategies also enable real-time monitoring of nanobased therapies. Such integrated diagnostic and therapeutic functions within multifunctional NPs will be crucial to improving patient outcomes. Importantly, the modular and tunable properties of these nanosystems enable continuous improvement to maximize treatment personalization on the basis of cancer molecular profiles.

Given the rapid progression of anticancer nanomedicine, even more sophisticated and patient-tailored therapeutic methods based on precise drug delivery nanosystems are expected in the near future.

Conflict of interest

The authors have no relevant financial or nonfinancial interests to disclose.

The authors have no conflicts of interest to declare that are relevant to the content of this article.

All the authors certify that they have no affiliations with or involvement in any organization or entity with any financial interest or nonfinancial interest in the subject matter or materials discussed in this manuscript.

The authors have no financial or proprietary interests in any material discussed in this article.

Funding

Authors declare no funding.

Ethical approval

Ethical approval was not necessary for the preparation of this article.

Consent to participate not applicable.

References

1. Minelli C, Lowe SB, Stevens MM. Engineering nanocomposite materials for cancer therapy. *Small*. 2010; 6(21): 2336–2357, doi: [10.1002/sml.201000523](https://doi.org/10.1002/sml.201000523), indexed in Pubmed: [20878632](https://pubmed.ncbi.nlm.nih.gov/20878632/).
2. Blanco E, Shen H, Ferrari M. Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nat Biotechnol*. 2015; 33(9): 941–951, doi: [10.1038/nbt.3330](https://doi.org/10.1038/nbt.3330), indexed in Pubmed: [26348965](https://pubmed.ncbi.nlm.nih.gov/26348965/).

3. Maeda H, Wu J, Sawa T, et al. Tumor vascular permeability and the EPR effect in macromolecular therapeutics: a review. *J Control Release*. 2000; 65(1-2): 271–284, doi: [10.1016/s0168-3659\(99\)00248-5](https://doi.org/10.1016/s0168-3659(99)00248-5), indexed in Pubmed: [10699287](https://pubmed.ncbi.nlm.nih.gov/10699287/).
4. Alexis F, Pridgen E, Molnar LK, et al. Factors affecting the clearance and biodistribution of polymeric nanoparticles. *Mol Pharm*. 2008; 5(4): 505–515, doi: [10.1021/mp800051m](https://doi.org/10.1021/mp800051m), indexed in Pubmed: [18672949](https://pubmed.ncbi.nlm.nih.gov/18672949/).
5. Estrella V, Chen T, Lloyd M, et al. Acidity generated by the tumor microenvironment drives local invasion. *Cancer Res*. 2013; 73(5): 1524–1535, doi: [10.1158/0008-5472.CAN-12-2796](https://doi.org/10.1158/0008-5472.CAN-12-2796), indexed in Pubmed: [23288510](https://pubmed.ncbi.nlm.nih.gov/23288510/).
6. Peer D, Karp JM, Hong S, et al. Nanocarriers as an emerging platform for cancer therapy. *Nat Nanotechnol*. 2007; 2(12): 751–760, doi: [10.1038/nnano.2007.387](https://doi.org/10.1038/nnano.2007.387), indexed in Pubmed: [18654426](https://pubmed.ncbi.nlm.nih.gov/18654426/).
7. Danhier F. To exploit the tumor microenvironment: Since the EPR effect fails in the clinic, what is the future of nanomedicine? *J Control Release*. 2016; 244(Pt A): 108–121, doi: [10.1016/j.jconrel.2016.11.015](https://doi.org/10.1016/j.jconrel.2016.11.015), indexed in Pubmed: [27871992](https://pubmed.ncbi.nlm.nih.gov/27871992/).
8. Jones DS, Silverman AP, Cochran JR. Developing therapeutic proteins by engineering ligand-receptor interactions. *Trends Biotechnol*. 2008; 26(9): 498–505, doi: [10.1016/j.tibtech.2008.05.009](https://doi.org/10.1016/j.tibtech.2008.05.009), indexed in Pubmed: [18675482](https://pubmed.ncbi.nlm.nih.gov/18675482/).
9. Qian ZM, Li H, Sun H, et al. Targeted drug delivery via the transferrin receptor-mediated endocytosis pathway. *Pharmacol Rev*. 2002; 54(4): 561–587, doi: [10.1124/pr.54.4.561](https://doi.org/10.1124/pr.54.4.561), indexed in Pubmed: [12429868](https://pubmed.ncbi.nlm.nih.gov/12429868/).
10. Farokhzad OC, Cheng J, Teply BA, et al. Targeted nanoparticle-aptamer bioconjugates for cancer chemotherapy in vivo. *Proc Natl Acad Sci U S A*. 2006; 103(16): 6315–6320, doi: [10.1073/pnas.0601755103](https://doi.org/10.1073/pnas.0601755103), indexed in Pubmed: [16606824](https://pubmed.ncbi.nlm.nih.gov/16606824/).
11. Bertrand N, Wu J, Xu X, et al. Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv Drug Deliv Rev*. 2014; 66: 2–25, doi: [10.1016/j.addr.2013.11.009](https://doi.org/10.1016/j.addr.2013.11.009), indexed in Pubmed: [24270007](https://pubmed.ncbi.nlm.nih.gov/24270007/).
12. Shi J, Kantoff PW, Wooster R, et al. Cancer nanomedicine: progress, challenges and opportunities. *Nat Rev Cancer*. 2017; 17(1): 20–37, doi: [10.1038/nrc.2016.108](https://doi.org/10.1038/nrc.2016.108), indexed in Pubmed: [27834398](https://pubmed.ncbi.nlm.nih.gov/27834398/).
13. Maeda H. The enhanced permeability and retention (EPR) effect in tumor vasculature: the key role of tumor-selective macromolecular drug targeting. *Adv Enzyme Regul*. 2001; 41: 189–207, doi: [10.1016/s0065-2571\(00\)00013-3](https://doi.org/10.1016/s0065-2571(00)00013-3), indexed in Pubmed: [11384745](https://pubmed.ncbi.nlm.nih.gov/11384745/).
14. Cabral H, Matsumoto Y, Mizuno K, et al. Accumulation of sub-100 nm polymeric micelles in poorly permeable tumours depends on size. *Nat Nanotechnol*. 2011; 6(12): 815–823, doi: [10.1038/nnano.2011.166](https://doi.org/10.1038/nnano.2011.166), indexed in Pubmed: [22020122](https://pubmed.ncbi.nlm.nih.gov/22020122/).
15. Gaumet M, Vargas A, Gurny R, et al. Nanoparticles for drug delivery: the need for precision in reporting particle size parameters. *Eur J Pharm Biopharm*. 2008; 69(1): 1–9, doi: [10.1016/j.ejpb.2007.08.001](https://doi.org/10.1016/j.ejpb.2007.08.001), indexed in Pubmed: [17826969](https://pubmed.ncbi.nlm.nih.gov/17826969/).
16. Nguyen PV, Hervé-Aubert K, Chourpa I, et al. Active targeting strategy in nanomedicines using anti-EGFR ligands - A promising approach for cancer therapy and diagnosis. *Int J Pharm*. 2021; 609: 121134, doi: [10.1016/j.ijpharm.2021.121134](https://doi.org/10.1016/j.ijpharm.2021.121134), indexed in Pubmed: [34571073](https://pubmed.ncbi.nlm.nih.gov/34571073/).
17. Talekar M, Kendall J, Denny W, et al. Targeting of nanoparticles in cancer: drug delivery and diagnostics. *Anticancer Drugs*. 2011; 22(10): 949–962, doi: [10.1097/CAD.0b013e32834a4554](https://doi.org/10.1097/CAD.0b013e32834a4554), indexed in Pubmed: [21970851](https://pubmed.ncbi.nlm.nih.gov/21970851/).
18. Singla AK, Garg A, Aggarwal D. Paclitaxel and its formulations. *Int J Pharm*. 2002; 235(1-2): 179–192, doi: [10.1016/s0378-5173\(01\)00986-3](https://doi.org/10.1016/s0378-5173(01)00986-3), indexed in Pubmed: [11879753](https://pubmed.ncbi.nlm.nih.gov/11879753/).
19. Yuan H, Guo H, Luan X, et al. Albumin Nanoparticle of Paclitaxel (Abraxane) Decreases while Taxol Increases Breast Cancer Stem Cells in Treatment of Triple Negative Breast Cancer. *Mol Pharm*. 2020; 17(7): 2275–2286, doi: [10.1021/acs.molpharmaceut.9b01221](https://doi.org/10.1021/acs.molpharmaceut.9b01221), indexed in Pubmed: [32485107](https://pubmed.ncbi.nlm.nih.gov/32485107/).
20. Moreno-Aspitia A, Perez EA. Nanoparticle albumin-bound paclitaxel (ABI-007): a newer taxane alternative in breast cancer. *Future Oncol*. 2005; 1(6): 755–762, doi: [10.2217/14796694.1.6.755](https://doi.org/10.2217/14796694.1.6.755), indexed in Pubmed: [16556053](https://pubmed.ncbi.nlm.nih.gov/16556053/).
21. Montané X, Bajek A, Roszkowski K, et al. Encapsulation for Cancer Therapy. *Molecules*. 2020; 25(7), doi: [10.3390/molecules25071605](https://doi.org/10.3390/molecules25071605), indexed in Pubmed: [32244513](https://pubmed.ncbi.nlm.nih.gov/32244513/).
22. Janib SM, Moses AS, MacKay JA. Imaging and drug delivery using theranostic nanoparticles. *Adv Drug Deliv Rev*. 2010; 62(11): 1052–1063, doi: [10.1016/j.addr.2010.08.004](https://doi.org/10.1016/j.addr.2010.08.004), indexed in Pubmed: [20709124](https://pubmed.ncbi.nlm.nih.gov/20709124/).
23. Bae Y, Nishiyama N, Fukushima S, et al. Preparation and biological characterization of polymeric micelle drug carriers with intracellular pH-triggered drug release property: tumor permeability, controlled subcellular drug distribution, and enhanced in vivo antitumor efficacy. *Bioconjug Chem*. 2005; 16(1): 122–130, doi: [10.1021/bc0498166](https://doi.org/10.1021/bc0498166), indexed in Pubmed: [15656583](https://pubmed.ncbi.nlm.nih.gov/15656583/).
24. Allen TM, Cullis PR. Drug delivery systems: entering the mainstream. *Science*. 2004; 303(5665): 1818–1822, doi: [10.1126/science.1095833](https://doi.org/10.1126/science.1095833), indexed in Pubmed: [15031496](https://pubmed.ncbi.nlm.nih.gov/15031496/).
25. Camorani S, Tortorella S, Agnello L, et al. Aptamer-Functionalized Nanoparticles Mediate PD-L1 siRNA Delivery for Effective Gene Silencing in Triple-Negative Breast Cancer Cells. *Pharmaceutics*. 2022; 14(10), doi: [10.3390/pharmaceutics14102225](https://doi.org/10.3390/pharmaceutics14102225), indexed in Pubmed: [36297659](https://pubmed.ncbi.nlm.nih.gov/36297659/).
26. Eljack S, David S, Chourpa I, et al. Formulation of Lipid-Based Nanoparticles for Simultaneous Delivery of Lapatinib and Anti-Survivin siRNA for HER2+ Breast Cancer Treatment. *Pharmaceuticals (Basel)*. 2022; 15(12), doi: [10.3390/ph15121452](https://doi.org/10.3390/ph15121452), indexed in Pubmed: [36558904](https://pubmed.ncbi.nlm.nih.gov/36558904/).
27. Yang T, Han H, Chen Y, et al. Study the lipidoid nanoparticle mediated genome editing protein delivery using 3D intestinal tissue model. *Bioact Mater*. 2021; 6(11): 3671–3677, doi: [10.1016/j.bioactmat.2021.03.027](https://doi.org/10.1016/j.bioactmat.2021.03.027), indexed in Pubmed: [33898871](https://pubmed.ncbi.nlm.nih.gov/33898871/).
28. Richter F, Leer K, Martin L, et al. The impact of anionic polymers on gene delivery: how composition and assembly help evading the toxicity-efficiency dilemma. *J Nanobiotechnology*. 2021; 19(1): 292, doi: [10.1186/s12951-021-00994-2](https://doi.org/10.1186/s12951-021-00994-2), indexed in Pubmed: [34579715](https://pubmed.ncbi.nlm.nih.gov/34579715/).
29. Terada T, Kulkarni JA, Huynh A, et al. Characterization of Lipid Nanoparticles Containing Ionizable Cationic Lipids Using Design-of-Experiments Approach. *Langmuir*. 2021; 37(3): 1120–1128, doi: [10.1021/acs.langmuir.0c03039](https://doi.org/10.1021/acs.langmuir.0c03039), indexed in Pubmed: [33439022](https://pubmed.ncbi.nlm.nih.gov/33439022/).

30. Chelakkot C, Ghim J, Ryu SHo. Mechanisms regulating intestinal barrier integrity and its pathological implications. *Exp Mol Med*. 2018; 50(8): 1–9, doi: [10.1038/s12276-018-0126-x](https://doi.org/10.1038/s12276-018-0126-x), indexed in Pubmed: [30115904](https://pubmed.ncbi.nlm.nih.gov/30115904/).
31. Lin CH, Chen CH, Lin ZC, et al. Recent advances in oral delivery of drugs and bioactive natural products using solid lipid nanoparticles as the carriers. *J Food Drug Anal*. 2017; 25(2): 219–234, doi: [10.1016/j.jfda.2017.02.001](https://doi.org/10.1016/j.jfda.2017.02.001), indexed in Pubmed: [28911663](https://pubmed.ncbi.nlm.nih.gov/28911663/).
32. Ensign LM, Cone R, Hanes J. Oral drug delivery with polymeric nanoparticles: the gastrointestinal mucus barriers. *Adv Drug Deliv Rev*. 2012; 64(6): 557–570, doi: [10.1016/j.addr.2011.12.009](https://doi.org/10.1016/j.addr.2011.12.009), indexed in Pubmed: [22212900](https://pubmed.ncbi.nlm.nih.gov/22212900/).
33. Lai SK, Wang YY, Hanes J. Mucus-penetrating nanoparticles for drug and gene delivery to mucosal tissues. *Adv Drug Deliv Rev*. 2009; 61(2): 158–171, doi: [10.1016/j.addr.2008.11.002](https://doi.org/10.1016/j.addr.2008.11.002), indexed in Pubmed: [19133304](https://pubmed.ncbi.nlm.nih.gov/19133304/).
34. Bailey MM, Berkland CJ. Nanoparticle formulations in pulmonary drug delivery. *Med Res Rev*. 2009; 29(1): 196–212, doi: [10.1002/med.20140](https://doi.org/10.1002/med.20140), indexed in Pubmed: [18958847](https://pubmed.ncbi.nlm.nih.gov/18958847/).
35. Hashemi Goradel N, Ghiyami-Hour F, Jahangiri S, et al. Nanoparticles as new tools for inhibition of cancer angiogenesis. *J Cell Physiol*. 2018; 233(4): 2902–2910, doi: [10.1002/jcp.26029](https://doi.org/10.1002/jcp.26029), indexed in Pubmed: [28543172](https://pubmed.ncbi.nlm.nih.gov/28543172/).
36. Harrison L, Blackwell K. Hypoxia and anemia: factors in decreased sensitivity to radiation therapy and chemotherapy? *Oncologist*. 2004; 9 Suppl 5: 31–40, doi: [10.1634/theoncologist.9-90005-31](https://doi.org/10.1634/theoncologist.9-90005-31), indexed in Pubmed: [15591420](https://pubmed.ncbi.nlm.nih.gov/15591420/).
37. Keereweer S, Van Driel PB, Snoeks TJA, et al. Optical image-guided cancer surgery: challenges and limitations. *Clin Cancer Res*. 2013; 19(14): 3745–3754, doi: [10.1158/1078-0432.CCR-12-3598](https://doi.org/10.1158/1078-0432.CCR-12-3598), indexed in Pubmed: [23674494](https://pubmed.ncbi.nlm.nih.gov/23674494/).
38. Wong C, Stylianopoulos T, Cui J, et al. Multistage nanoparticle delivery system for deep penetration into tumor tissue. *Proc Natl Acad Sci U S A*. 2011; 108(6): 2426–2431, doi: [10.1073/pnas.1018382108](https://doi.org/10.1073/pnas.1018382108), indexed in Pubmed: [21245339](https://pubmed.ncbi.nlm.nih.gov/21245339/).
39. Torchilin V. Tumor delivery of macromolecular drugs based on the EPR effect. *Adv Drug Deliv Rev*. 2011; 63(3): 131–135, doi: [10.1016/j.addr.2010.03.011](https://doi.org/10.1016/j.addr.2010.03.011), indexed in Pubmed: [20304019](https://pubmed.ncbi.nlm.nih.gov/20304019/).
40. Irvine DJ, Hanson MC, Rakhra K, et al. Synthetic Nanoparticles for Vaccines and Immunotherapy. *Chem Rev*. 2015; 115(19): 11109–11146, doi: [10.1021/acs.chemrev.5b00109](https://doi.org/10.1021/acs.chemrev.5b00109), indexed in Pubmed: [26154342](https://pubmed.ncbi.nlm.nih.gov/26154342/).
41. Cruz LJ, Rosalia RA, Kleinovink JW, et al. Targeting nanoparticles to CD40, DEC-205 or CD11c molecules on dendritic cells for efficient CD8(+) T cell response: a comparative study. *J Control Release*. 2014; 192: 209–218, doi: [10.1016/j.jconrel.2014.07.040](https://doi.org/10.1016/j.jconrel.2014.07.040), indexed in Pubmed: [25068703](https://pubmed.ncbi.nlm.nih.gov/25068703/).
42. Stephan MT, Moon JJ, Um SHo, et al. Therapeutic cell engineering with surface-conjugated synthetic nanoparticles. *Nat Med*. 2010; 16(9): 1035–1041, doi: [10.1038/nm.2198](https://doi.org/10.1038/nm.2198), indexed in Pubmed: [20711198](https://pubmed.ncbi.nlm.nih.gov/20711198/).
43. Buschow SI, Lasonder E, van Deutekom HWM, et al. Dominant processes during human dendritic cell maturation revealed by integration of proteome and transcriptome at the pathway level. *J Proteome Res*. 2010; 9(4): 1727–1737, doi: [10.1021/pr9008546](https://doi.org/10.1021/pr9008546), indexed in Pubmed: [20131907](https://pubmed.ncbi.nlm.nih.gov/20131907/).
44. Rosalia RA, Cruz LJ, van Duikeren S, et al. CD40-targeted dendritic cell delivery of PLGA-nanoparticle vaccines induce potent anti-tumor responses. *Biomaterials*. 2015; 40: 88–97, doi: [10.1016/j.biomaterials.2014.10.053](https://doi.org/10.1016/j.biomaterials.2014.10.053), indexed in Pubmed: [25465442](https://pubmed.ncbi.nlm.nih.gov/25465442/).
45. Fang J, Islam W, Maeda H. Exploiting the dynamics of the EPR effect and strategies to improve the therapeutic effects of nanomedicines by using EPR effect enhancers. *Adv Drug Deliv Rev*. 2020; 157: 142–160, doi: [10.1016/j.addr.2020.06.005](https://doi.org/10.1016/j.addr.2020.06.005), indexed in Pubmed: [32553783](https://pubmed.ncbi.nlm.nih.gov/32553783/).
46. Nel A, Ruoslahti E, Meng H. New Insights into “Permeability” as in the Enhanced Permeability and Retention Effect of Cancer Nanotherapeutics. *ACS Nano*. 2017; 11(10): 9567–9569, doi: [10.1021/acsnano.7b07214](https://doi.org/10.1021/acsnano.7b07214), indexed in Pubmed: [29065443](https://pubmed.ncbi.nlm.nih.gov/29065443/).
47. Toy R, Peiris PM, Ghaghada KB, et al. Shaping cancer nanomedicine: the effect of particle shape on the in vivo journey of nanoparticles. *Nanomedicine (Lond)*. 2014; 9(1): 121–134, doi: [10.2217/nnm.13.191](https://doi.org/10.2217/nnm.13.191), indexed in Pubmed: [24354814](https://pubmed.ncbi.nlm.nih.gov/24354814/).
48. Kibria G, Hatakeyama H, Harashima H. Cancer multidrug resistance: mechanisms involved and strategies for circumvention using a drug delivery system. *Arch Pharm Res*. 2014; 37(1): 4–15, doi: [10.1007/s12272-013-0276-2](https://doi.org/10.1007/s12272-013-0276-2), indexed in Pubmed: [24272889](https://pubmed.ncbi.nlm.nih.gov/24272889/).
49. Byrne JD, Betancourt T, Brannon-Peppas L. Active targeting schemes for nanoparticle systems in cancer therapeutics. *Adv Drug Deliv Rev*. 2008; 60(15): 1615–1626, doi: [10.1016/j.addr.2008.08.005](https://doi.org/10.1016/j.addr.2008.08.005), indexed in Pubmed: [18840489](https://pubmed.ncbi.nlm.nih.gov/18840489/).
50. Jaracz S, Chen J, Kuznetsova LV, et al. Recent advances in tumor-targeting anticancer drug conjugates. *Bioorg Med Chem*. 2005; 13(17): 5043–5054, doi: [10.1016/j.bmc.2005.04.084](https://doi.org/10.1016/j.bmc.2005.04.084), indexed in Pubmed: [15955702](https://pubmed.ncbi.nlm.nih.gov/15955702/).
51. Elias DR, Poloukhina A, Popik V, et al. Effect of ligand density, receptor density, and nanoparticle size on cell targeting. *Nanomedicine*. 2013; 9(2): 194–201, doi: [10.1016/j.nano.2012.05.015](https://doi.org/10.1016/j.nano.2012.05.015), indexed in Pubmed: [22687896](https://pubmed.ncbi.nlm.nih.gov/22687896/).
52. Parveen S, Sahoo SK. Polymeric nanoparticles for cancer therapy. *J Drug Target*. 2008; 16(2): 108–123, doi: [10.1080/10611860701794353](https://doi.org/10.1080/10611860701794353), indexed in Pubmed: [18274932](https://pubmed.ncbi.nlm.nih.gov/18274932/).
53. Saptarshi SR, Duschl A, Lopata AL. Interaction of nanoparticles with proteins: relation to bio-reactivity of the nanoparticle. *J Nanobiotechnology*. 2013; 11: 26, doi: [10.1186/1477-3155-11-26](https://doi.org/10.1186/1477-3155-11-26), indexed in Pubmed: [23870291](https://pubmed.ncbi.nlm.nih.gov/23870291/).
54. Shi J, Votruba AR, Farokhzad OC, et al. Nanotechnology in drug delivery and tissue engineering: from discovery to applications. *Nano Lett*. 2010; 10(9): 3223–3230, doi: [10.1021/nl102184c](https://doi.org/10.1021/nl102184c), indexed in Pubmed: [20726522](https://pubmed.ncbi.nlm.nih.gov/20726522/).
55. Dreaden EC, Austin LA, Mackey MA, et al. Size matters: gold nanoparticles in targeted cancer drug delivery. *Ther Deliv*. 2012; 3(4): 457–478, doi: [10.4155/tde.12.21](https://doi.org/10.4155/tde.12.21), indexed in Pubmed: [22834077](https://pubmed.ncbi.nlm.nih.gov/22834077/).
56. Cruz E, Kayser V. Synthesis and Enhanced Cellular Uptake In Vitro of Anti-HER2 Multifunctional Gold Nanoparticles. *Cancers (Basel)*. 2019; 11(6), doi: [10.3390/cancers11060870](https://doi.org/10.3390/cancers11060870), indexed in Pubmed: [31234432](https://pubmed.ncbi.nlm.nih.gov/31234432/).
57. Huang X, El-Sayed IH, Qian W, et al. Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods. *J Am Chem Soc*. 2006; 128(6): 2115–2120, doi: [10.1021/ja057254a](https://doi.org/10.1021/ja057254a), indexed in Pubmed: [16464114](https://pubmed.ncbi.nlm.nih.gov/16464114/).
58. Liong M, Lu J, Kovochich M, et al. Multifunctional inorganic nanoparticles for imaging, targeting, and drug delivery.

- ACS Nano. 2008; 2(5): 889–896, doi: [10.1021/nn800072t](https://doi.org/10.1021/nn800072t), indexed in Pubmed: [19206485](https://pubmed.ncbi.nlm.nih.gov/19206485/).
59. Chen L, Zhou X, Nie W, et al. Multifunctional Redox-Responsive Mesoporous Silica Nanoparticles for Efficient Targeting Drug Delivery and Magnetic Resonance Imaging. *ACS Appl Mater Interfaces*. 2016; 8(49): 33829–33841, doi: [10.1021/acsami.6b11802](https://doi.org/10.1021/acsami.6b11802), indexed in Pubmed: [27960384](https://pubmed.ncbi.nlm.nih.gov/27960384/).
60. Huang G, Chen H, Dong Y, et al. Superparamagnetic iron oxide nanoparticles: amplifying ROS stress to improve anticancer drug efficacy. *Theranostics*. 2013; 3(2): 116–126, doi: [10.7150/thno.5411](https://doi.org/10.7150/thno.5411), indexed in Pubmed: [23423156](https://pubmed.ncbi.nlm.nih.gov/23423156/).
61. Nance E, Timbie K, Miller GW, et al. Non-invasive delivery of stealth, brain-penetrating nanoparticles across the blood-brain barrier using MRI-guided focused ultrasound. *J Control Release*. 2014; 189: 123–132, doi: [10.1016/j.jconrel.2014.06.031](https://doi.org/10.1016/j.jconrel.2014.06.031), indexed in Pubmed: [24979210](https://pubmed.ncbi.nlm.nih.gov/24979210/).
62. Xie J, Lee S, Chen X. Nanoparticle-based therapeutic agents. *Adv Drug Deliv Rev*. 2010; 62(11): 1064–1079, doi: [10.1016/j.addr.2010.07.009](https://doi.org/10.1016/j.addr.2010.07.009), indexed in Pubmed: [20691229](https://pubmed.ncbi.nlm.nih.gov/20691229/).
63. Lammers T, Aime S, Hennink WE, et al. Theranostic nanomedicine. *Acc Chem Res*. 2011; 44(10): 1029–1038, doi: [10.1021/ar200019c](https://doi.org/10.1021/ar200019c), indexed in Pubmed: [21545096](https://pubmed.ncbi.nlm.nih.gov/21545096/).
64. Khatun Z, Nurunnabi Md, Nafiujjaman Md, et al. Photoluminescent graphene nanoparticles for cancer phototherapy and imaging. *ACS Appl Mater Interfaces*. 2014; 6(15): 12413–12421, doi: [10.1021/am504071z](https://doi.org/10.1021/am504071z), indexed in Pubmed: [25054687](https://pubmed.ncbi.nlm.nih.gov/25054687/).
65. Robinson JT, Tabakman SM, Liang Y, et al. Ultrasmall reduced graphene oxide with high near-infrared absorbance for photothermal therapy. *J Am Chem Soc*. 2011; 133(17): 6825–6831, doi: [10.1021/ja2010175](https://doi.org/10.1021/ja2010175), indexed in Pubmed: [21476500](https://pubmed.ncbi.nlm.nih.gov/21476500/).
66. Yang K, Zhang S, Zhang G, et al. Graphene in mice: ultrahigh in vivo tumor uptake and efficient photothermal therapy. *Nano Lett*. 2010; 10(9): 3318–3323, doi: [10.1021/nl100996u](https://doi.org/10.1021/nl100996u), indexed in Pubmed: [20684528](https://pubmed.ncbi.nlm.nih.gov/20684528/).
67. Mamaeva V, Sahlgrén C, Lindén M. Mesoporous silica nanoparticles in medicine—recent advances. *Adv Drug Deliv Rev*. 2013; 65(5): 689–702, doi: [10.1016/j.addr.2012.07.018](https://doi.org/10.1016/j.addr.2012.07.018), indexed in Pubmed: [22921598](https://pubmed.ncbi.nlm.nih.gov/22921598/).
68. Slowing II, Vivero-Escoto JL, Wu CW, et al. Mesoporous silica nanoparticles as controlled release drug delivery and gene transfection carriers. *Adv Drug Deliv Rev*. 2008; 60(11): 1278–1288, doi: [10.1016/j.addr.2008.03.012](https://doi.org/10.1016/j.addr.2008.03.012), indexed in Pubmed: [18514969](https://pubmed.ncbi.nlm.nih.gov/18514969/).
69. Hom C, Lu J, Liang M, et al. Mesoporous silica nanoparticles facilitate delivery of siRNA to shutdown signaling pathways in mammalian cells. *Small*. 2010; 6(11): 1185–1190, doi: [10.1002/smll.200901966](https://doi.org/10.1002/smll.200901966), indexed in Pubmed: [20461725](https://pubmed.ncbi.nlm.nih.gov/20461725/).
70. Choi KiY, Min KH, Yoon HY, et al. PEGylation of hyaluronic acid nanoparticles improves tumor targetability in vivo. *Biomaterials*. 2011; 32(7): 1880–1889, doi: [10.1016/j.biomaterials.2010.11.010](https://doi.org/10.1016/j.biomaterials.2010.11.010), indexed in Pubmed: [21159377](https://pubmed.ncbi.nlm.nih.gov/21159377/).
71. Hossann M, Wang T, Wiggenshorn M, et al. Size of thermosensitive liposomes influences content release. *J Control Release*. 2010; 147(3): 436–443, doi: [10.1016/j.jconrel.2010.08.013](https://doi.org/10.1016/j.jconrel.2010.08.013), indexed in Pubmed: [20727921](https://pubmed.ncbi.nlm.nih.gov/20727921/).
72. Farokhzad OC, Jon S, Khademhosseini A, et al. Nanoparticle-aptamer bioconjugates: a new approach for targeting prostate cancer cells. *Cancer Res*. 2004; 64(21): 7668–7672, doi: [10.1158/0008-5472.CAN-04-2550](https://doi.org/10.1158/0008-5472.CAN-04-2550), indexed in Pubmed: [15520166](https://pubmed.ncbi.nlm.nih.gov/15520166/).
73. Wang AZ, Bagalkot V, Vasilliou CC, et al. Superparamagnetic iron oxide nanoparticle-aptamer bioconjugates for combined prostate cancer imaging and therapy. *ChemMedChem*. 2008; 3(9): 1311–1315, doi: [10.1002/cmdc.200800091](https://doi.org/10.1002/cmdc.200800091), indexed in Pubmed: [18613203](https://pubmed.ncbi.nlm.nih.gov/18613203/).
74. Sahay G, Querbes W, Alabi C, et al. Efficiency of siRNA delivery by lipid nanoparticles is limited by endocytic recycling. *Nat Biotechnol*. 2013; 31(7): 653–658, doi: [10.1038/nbt.2614](https://doi.org/10.1038/nbt.2614), indexed in Pubmed: [23792629](https://pubmed.ncbi.nlm.nih.gov/23792629/).
75. Yin H, Kanasty RL, Eltoukhy AA, et al. Non-viral vectors for gene-based therapy. *Nat Rev Genet*. 2014; 15(8): 541–555, doi: [10.1038/nrg3763](https://doi.org/10.1038/nrg3763), indexed in Pubmed: [25022906](https://pubmed.ncbi.nlm.nih.gov/25022906/).
76. Niculescu AG, Grumezescu AM. Novel Tumor-Targeting Nanoparticles for Cancer Treatment—A Review. *Int J Mol Sci*. 2022; 23(9), doi: [10.3390/ijms23095253](https://doi.org/10.3390/ijms23095253), indexed in Pubmed: [35563645](https://pubmed.ncbi.nlm.nih.gov/35563645/).
77. Khan MdI, Hossain MI, Hossain MK, et al. Recent Progress in Nanostructured Smart Drug Delivery Systems for Cancer Therapy: A Review. *ACS Appl Bio Mater*. 2022; 5(3): 971–1012, doi: [10.1021/acsabm.2c00002](https://doi.org/10.1021/acsabm.2c00002), indexed in Pubmed: [35226465](https://pubmed.ncbi.nlm.nih.gov/35226465/).
78. Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer*. 2002; 2(1): 48–58, doi: [10.1038/nrc706](https://doi.org/10.1038/nrc706), indexed in Pubmed: [11902585](https://pubmed.ncbi.nlm.nih.gov/11902585/).
79. Brigger I, Dubernet C, Couvreur P, et al. Tamoxifen encapsulation within polyethylene glycol-coated nanospheres. A new antiestrogen formulation. *Int J Pharm*. 2001; 214(1–2): 37–42, doi: [10.1016/s0378-5173\(00\)00628-1](https://doi.org/10.1016/s0378-5173(00)00628-1), indexed in Pubmed: [11282234](https://pubmed.ncbi.nlm.nih.gov/11282234/).
80. Dai Q, Wilhelm S, Ding D, et al. Quantifying the Ligand-Coated Nanoparticle Delivery to Cancer Cells in Solid Tumors. *ACS Nano*. 2018; 12(8): 8423–8435, doi: [10.1021/acsnano.8b03900](https://doi.org/10.1021/acsnano.8b03900), indexed in Pubmed: [30016073](https://pubmed.ncbi.nlm.nih.gov/30016073/).
81. Maeda H. Vascular permeability in cancer and infection as related to macromolecular drug delivery, with emphasis on the EPR effect for tumor-selective drug targeting. *Proc Jpn Acad Ser B Phys Biol Sci*. 2012; 88(3): 53–71, doi: [10.2183/pjab.88.53](https://doi.org/10.2183/pjab.88.53), indexed in Pubmed: [22450535](https://pubmed.ncbi.nlm.nih.gov/22450535/).
82. Zhang D, Liu L, Wang J, et al. Drug-loaded PEG-PLGA nanoparticles for cancer treatment. *Front Pharmacol*. 2022; 13: 990505, doi: [10.3389/fphar.2022.990505](https://doi.org/10.3389/fphar.2022.990505), indexed in Pubmed: [36059964](https://pubmed.ncbi.nlm.nih.gov/36059964/).
83. Dean M, Hamon Y, Chimini G. The human ATP-binding cassette (ABC) transporter superfamily. *J Lipid Res*. 2001; 42(7): 1007–1017, indexed in Pubmed: [11441126](https://pubmed.ncbi.nlm.nih.gov/11441126/).
84. Scotto KW. Transcriptional regulation of ABC drug transporters. *Oncogene*. 2003; 22(47): 7496–7511, doi: [10.1038/sj.onc.1206950](https://doi.org/10.1038/sj.onc.1206950), indexed in Pubmed: [14576854](https://pubmed.ncbi.nlm.nih.gov/14576854/).
85. Ambudkar SV, Kimchi-Sarfaty C, Sauna ZE, et al. P-glycoprotein: from genomics to mechanism. *Oncogene*. 2003; 22(47): 7468–7485, doi: [10.1038/sj.onc.1206948](https://doi.org/10.1038/sj.onc.1206948), indexed in Pubmed: [14576852](https://pubmed.ncbi.nlm.nih.gov/14576852/).
86. Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer*. 2002; 2(1): 48–58, doi: [10.1038/nrc706](https://doi.org/10.1038/nrc706), indexed in Pubmed: [11902585](https://pubmed.ncbi.nlm.nih.gov/11902585/).
87. Fletcher JI, Haber M, Henderson MJ, et al. ABC transporters in cancer: more than just drug efflux pumps. *Nat Rev Cancer*. 2010; 10(2): 147–156, doi: [10.1038/nrc2789](https://doi.org/10.1038/nrc2789), indexed in Pubmed: [20075923](https://pubmed.ncbi.nlm.nih.gov/20075923/).

88. Sangtani A, Petryayeva E, Susumu K, et al. Nanoparticle-Peptide-Drug Bioconjugates for Unassisted Defeat of Multidrug Resistance in a Model Cancer Cell Line. *Bioconj Chem.* 2019; 30(3): 525–530, doi: [10.1021/acs.bioconjchem.8b00755](https://doi.org/10.1021/acs.bioconjchem.8b00755), indexed in Pubmed: [30735042](https://pubmed.ncbi.nlm.nih.gov/30735042/).
89. Huszthy PC, Girolou T, Tsinkalovsky O, et al. Remission of invasive, cancer stem-like glioblastoma xenografts using lentiviral vector-mediated suicide gene therapy. *PLoS One.* 2009; 4(7): e6314, doi: [10.1371/journal.pone.0006314](https://doi.org/10.1371/journal.pone.0006314), indexed in Pubmed: [19617915](https://pubmed.ncbi.nlm.nih.gov/19617915/).
90. Wang K, Kievit FM, Zhang M. Nanoparticles for cancer gene therapy: Recent advances, challenges, and strategies. *Pharmacol Res.* 2016; 114: 56–66, doi: [10.1016/j.phrs.2016.10.016](https://doi.org/10.1016/j.phrs.2016.10.016), indexed in Pubmed: [27771464](https://pubmed.ncbi.nlm.nih.gov/27771464/).
91. Tzeng SY, Patel KK, Wilson DR, et al. In situ genetic engineering of tumors for long-lasting and systemic immunotherapy. *Proc Natl Acad Sci U S A.* 2020; 117(8): 4043–4052, doi: [10.1073/pnas.1916039117](https://doi.org/10.1073/pnas.1916039117), indexed in Pubmed: [32034097](https://pubmed.ncbi.nlm.nih.gov/32034097/).
92. Varlamova EG, Goltyaev MV, Mal'tseva VN, et al. Mechanisms of the Cytotoxic Effect of Selenium Nanoparticles in Different Human Cancer Cell Lines. *Int J Mol Sci.* 2021; 22(15), doi: [10.3390/ijms22157798](https://doi.org/10.3390/ijms22157798), indexed in Pubmed: [34360564](https://pubmed.ncbi.nlm.nih.gov/34360564/).
93. Mazumdar S, Chitkara D, Mittal A. Exploration and insights into the cellular internalization and intracellular fate of amphiphilic polymeric nanocarriers. *Acta Pharm Sin B.* 2021; 11(4): 903–924, doi: [10.1016/j.apsb.2021.02.019](https://doi.org/10.1016/j.apsb.2021.02.019), indexed in Pubmed: [33996406](https://pubmed.ncbi.nlm.nih.gov/33996406/).
94. Bechet D, Couleaud P, Frochot C, et al. Nanoparticles as vehicles for delivery of photodynamic therapy agents. *Trends Biotechnol.* 2008; 26(11): 612–621, doi: [10.1016/j.tibtech.2008.07.007](https://doi.org/10.1016/j.tibtech.2008.07.007), indexed in Pubmed: [18804298](https://pubmed.ncbi.nlm.nih.gov/18804298/).
95. Allison RR, Sibata CH. Oncologic photodynamic therapy photosensitizers: a clinical review. *Photodiagnosis Photodyn Ther.* 2010; 7(2): 61–75, doi: [10.1016/j.pdpdt.2010.02.001](https://doi.org/10.1016/j.pdpdt.2010.02.001), indexed in Pubmed: [20510301](https://pubmed.ncbi.nlm.nih.gov/20510301/).
96. Ricci-Júnior E, Marchetti JM, Ricci-Júnior E, et al. Zinc(II) phthalocyanine loaded PLGA nanoparticles for photodynamic therapy use. *Int J Pharm.* 2006; 310(1-2): 187–195, doi: [10.1016/j.ijpharm.2005.10.048](https://doi.org/10.1016/j.ijpharm.2005.10.048), indexed in Pubmed: [16442755](https://pubmed.ncbi.nlm.nih.gov/16442755/).
97. Master AM, Sen Gupta A. EGF receptor-targeted nanocarriers for enhanced cancer treatment. *Nanomedicine (Lond).* 2012; 7(12): 1895–1906, doi: [10.2217/nnm.12.160](https://doi.org/10.2217/nnm.12.160), indexed in Pubmed: [23249333](https://pubmed.ncbi.nlm.nih.gov/23249333/).
98. Wong HoL, Rauth AM, Bendayan R, et al. A new polymer-lipid hybrid nanoparticle system increases cytotoxicity of doxorubicin against multidrug-resistant human breast cancer cells. *Pharm Res.* 2006; 23(7): 1574–1585, doi: [10.1007/s11095-006-0282-x](https://doi.org/10.1007/s11095-006-0282-x), indexed in Pubmed: [16786442](https://pubmed.ncbi.nlm.nih.gov/16786442/).