

Low-grade lipopolysaccharide-related endotoxemia alters coronary thrombus composition and fibrin clot properties in patients with acute ST-segment elevation myocardial infarction

Marcin Sadowski¹, Michał Ząbczyk^{2,3}, Anetta Undas^{2,3}

¹*Collegium Medicum, Jan Kochanowski University, Kielce, Poland*

²*Department of Thromboembolic Disorders, Institute of Cardiology, Jagiellonian University Medical College, Kraków, Poland*

³*Krakow Centre for Medical Research and Technologies, St. John Paul II Hospital, Kraków, Poland*

Correspondence to:

Marcin Sadowski, MD, PhD,
Institute of Medical Sciences,
Jan Kochanowski University,
IX Wieków Kielc 19A, 25–516
Kielce, Poland,
phone: +48 41 349 69 70,
e-mail: msadowski@ujk.edu.pl

Copyright by the Author(s), 2024

DOI: 10.33963/v.phj.102310

Received:

August 21, 2024

Accepted:

August 28, 2024

Early publication date:

August 28, 2024

INTRODUCTION

ST-segment elevation myocardial infarction (STEMI) frequently results from acute coronary flow cessation due to atherosclerotic plaque erosion or disruption followed by intracoronary thrombus (ICT) formation [1]. ICT composition and structure assessed using scanning electron microscopy (SEM) indicates the dominance of fibrin (60%) if thrombi are retrieved by thrombectomy within the first 12 hours from symptom onset [2]. In STEMI patients, the contribution of components within thrombi varies with time, which may affect their stability and resolution [2]. Cardiovascular risk factors, together with low-grade inflammation, promote prothrombotic fibrin clot properties, i.e., faster polymerization and formation of denser fibrin networks composed of thinner fibers, leading to increased resistance to fibrinolysis [3].

Alterations in the intestinal microbiome characterized by the reduction of commensal in favor of pathogenic bacteria, i.e., a gut dysbiosis, have been recently demonstrated as a proinflammatory and prothrombotic factor acting via translocation of bacterial products into the systemic circulation [4, 5]. Lipopolysaccharide (LPS), a glycolipid component of Gram-negative bacteria's outer membrane, is detectable in blood, and increased intestinal permeability, along with impaired LPS degradation, raises circulating LPS concentrations. LPS has been found to act mainly via toll-like receptor 4 expressed, among others, in immune and endothelial cells [6].

Carnevale et al. [4] reported LPS-positive areas within ICT, along with increased plasma P-selectin, a marker of platelet activation, associated with low-grade endotoxemia and gut permeability. They also found positive correlations of serum LPS with white blood cell (WBC) count and C-reactive protein [4]. Previously, we reported positive correlations of fibrinogen, P-selectin, plasminogen activator inhibitor-1 (PAI-1), von Willebrand factor antigen, soluble CD40 ligand, and myeloperoxidase with thrombus fibrin content [7]. After adjustment for fibrinogen and onset-to-thrombectomy time, circulating von Willebrand factor antigen, myeloperoxidase, and P-selectin, together with renal impairment, arterial hypertension, and onset-to-thrombectomy time were the independent predictors of fibrin-rich ICT formation [7]. It has been shown that the formation of denser plasma fibrin clots was independently associated with high fibrin content within the ICT [8]. Little is known about links between coronary thrombus composition and LPS-mediated inflammation [4, 9]. We hypothesized that prothrombotic plasma fibrin clot properties and the high thrombus fibrin content are associated with elevated serum LPS in STEMI patients.

METHODS

The study is a subanalysis of our previous report [7] on ICTs obtained 4.0–16.5 hours from chest-pain onset by manual aspiration during primary coronary intervention in STEMI patients recruited between January

and December 2012. Patients with an estimated glomerular filtration rate below 30 ml/min, acute infection, treatment with systemic corticosteroids or oral anticoagulants, malignancy, and in-stent thrombosis were excluded. SEM details and laboratory tests used are provided in the Supplementary Materials. The study has been approved by the Ethics Committee (6/2011). All participants gave written informed consent.

Statistical analysis

Continuous variables were expressed as means (standard deviations) or medians (interquartile ranges [IQR]). To reflect the wide ranges of the thrombus components, the exact ranges were provided. Differences were tested by Student's t-test or by the Mann-Whitney U test. Categorical variables were compared by chi-square or Fisher's exact test. The Pearson or Spearman rank correlation coefficients were calculated. A *P*-value <0.05 was considered statistically significant. Statistical analyses were performed with IBM SPSS Statistics 21.0.

RESULTS AND DISCUSSION

The study patients have been presented elsewhere [7]. Forty patients (24 [60%] women) aged 36 to 90 years were studied (16 patients in the pre-percutaneous coronary intervention [PCI] group and 24 patients in the post-PCI group). Cardiovascular risk factors distribution and past medical history were similar in both groups (Supplementary material, *Table S1*). The median symptom onset-to-thrombectomy time was 390 min (IQR 240–990 min) in the pre-PCI group and 420 min (IQR 285–885 min) in the post-PCI group. Median LPS in the pre-PCI group was 36.1 (34.9–38.9) pg/ml (range 32.8–44.9 pg/ml) and 35.9 (33.9–39.0) pg/ml (range 32.4–41.2 pg/ml) in the post-PCI group. There was no association between LPS and age, sex, comorbidities, and medications used in either group. In the post-PCI group, LPS was positively correlated with the onset-to-thrombectomy time ($r = 0.54$; $P = 0.002$). The pre-PCI group with LPS above the median had greater cardiac troponin T on admission (Supplementary material, *Table S1*), and LPS was correlated with WBC count ($r = 0.5$; $P = 0.001$). Platelet count, fibrinogen, or markers of platelet, neutrophil, and endothelial activation showed no associations with LPS (Supplementary material, *Table S1*).

Analysis of 323 SEM images showed that mean fibrin, red blood cell (RBC), platelet, and white blood cell content were estimated at 49.1% (range 2.7%–88.5%), 24.2% (range 0%–58.7%), 11.6% (range 0.1%–65.8%), and 3.7% (range 0%–31.3%), respectively. In the post-PCI group with LPS levels >35.9 pg/ml, thrombus RBC content was lower, while fibrin content was greater, compared to the remainder (25.8% [19.7–46.6] vs. 45.5% [27.9–63.0]; $P = 0.04$), and this association was even stronger in patients with LPS levels in the top quartile compared to the remainder (28.3% [19.3–48.8] vs. 59.6% [32.7–77.1]; $P = 0.024$). In the post-PCI group, serum LPS was positively correlated with

fibrin content ($r = 0.52$; $P = 0.007$) and negatively with RBC content ($r = -0.47$; $P = 0.018$). In the pre-PCI groups, these correlations were absent (*Figure 1*).

In the pre-PCI group, patients with LPS levels >36.1 pg/ml had longer clot lysis time (CLT) (Supplementary material, *Table S1*). LPS was negatively associated with K_s -TF ($r = -0.47$; $P = 0.017$) in the pre-PCI group, while positive correlations between LPS and CLT were observed in the pre- and post-PCI groups ($r = 0.74$; $P = 0.012$, and $r = 0.36$; $P = 0.045$, respectively, *Figure 1*). No associations with fibrinolysis-related proteins, including PAI-1, a major regulator of CLT, were detected.

Our study is the first to show that greater ICT fibrin content, along with lower plasma clot permeability and reduced lysability, are associated with increased serum LPS levels in STEMI patients. This suggests that endotoxemia in STEMI may affect fibrin clot phenotype and thrombus composition. In the current study, thrombus fibrin and RBC content were found to have a new modulator, i.e., LPS, which correlated with WBC and cardiac troponin T on admission but not with other markers of inflammation and myocardial or endothelial injury.

Nunes et al. [10] investigated LPS-dependent clot formation and architecture, but they did not study clot properties. Ząbczyk et al. [11] reported that endotoxemia may contribute to increased thrombin generation and PAI-1-mediated hypofibrinolysis in patients with pulmonary embolism [11]. This is consistent with our findings on the LPS-mediated antifibrinolytic effect of LPS in patients with atrial fibrillation [12]. Together with CLT, we also measured K_s , a key measure reflecting a pore size within fibrin networks. Lower K_s and prolonged CLT indicate a prothrombotic clot phenotype. In our study, K_s was negatively correlated with CLT in the pre- and post-PCI groups ($r = -0.7$; $P = 0.006$ and $r = -0.47$; $P = 0.012$, respectively), which reflects consistency of the data. An inverse association of K_s with LPS in the pre-PCI group suggests that low-grade endotoxemia in STEMI contributes to dense fibrin clot formation, likely via inflammatory mechanisms.

Study limitations should be acknowledged. The sample size is small, and the patients' pre- and post-PCI laboratory tests were assessed. Therefore, our results are preliminary and do not refer to the current antiplatelet strategies [13] with infrequent use of thrombectomy in STEMI. We did not assess zonulin, a marker of gut permeability. However, it is well established that the main source of circulating LPS in MI patients is the gut [4, 14], with a contribution from other sources being unlikely. However, we believe that the impact of elevated LPS on ICT composition may be evaluated regardless of its origin and that the hypothesis-generating results may stimulate further investigation in the field of endotoxemia-related hemostatic alterations.

We conclude that STEMI patients with low-grade LPS-mediated endotoxemia tend to form fibrin-rich intracoronary thrombi in association with less permeable and resistant-to-lysis plasma clots.

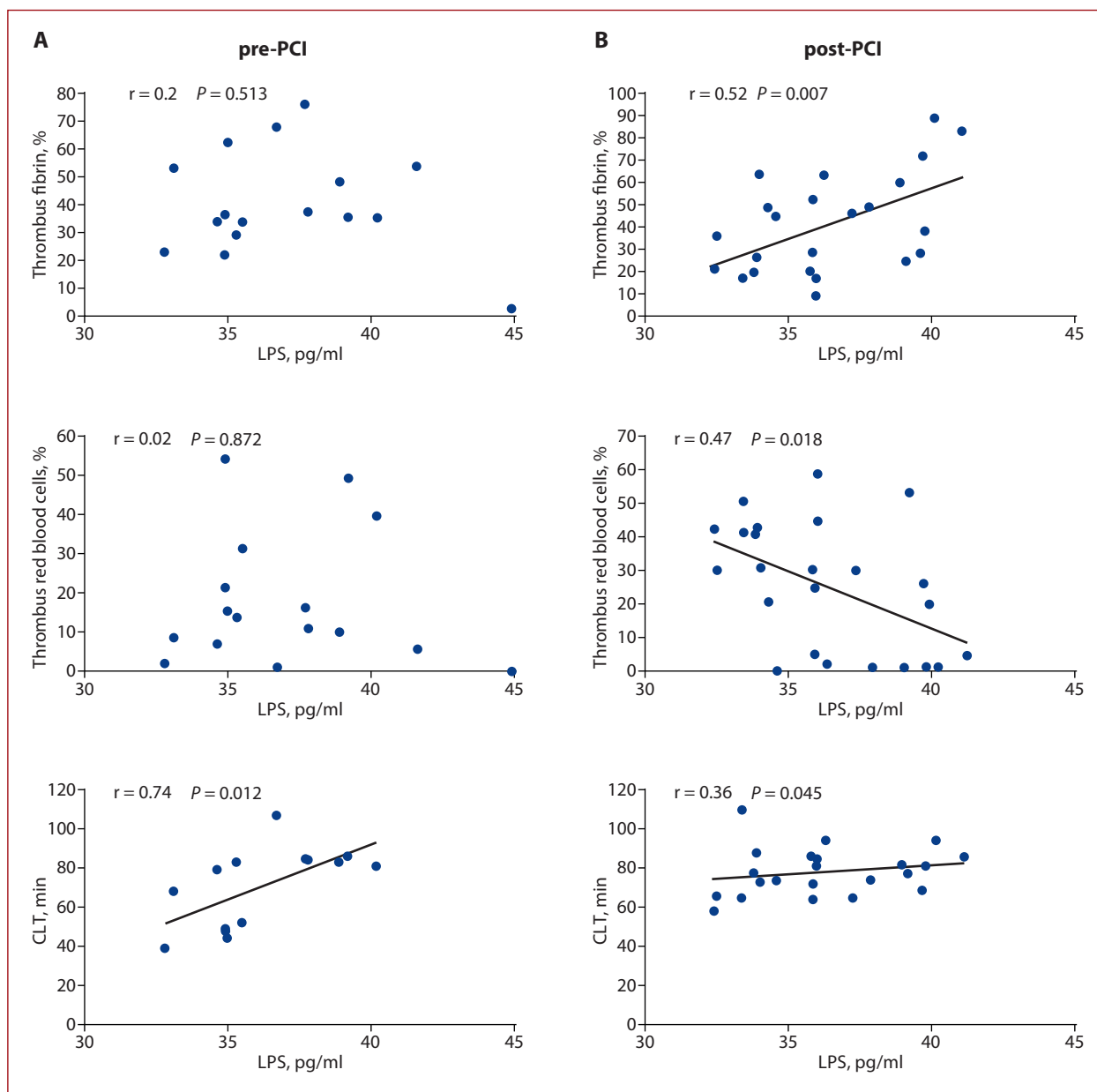


Figure 1. Correlations between serum lipopolysaccharide (LPS) levels and intracoronary thrombus contents, and serum LPS levels and clot lysis time (CLT) in the pre-percutaneous coronary intervention (PCI) (A) and post-PCI (B) groups

Supplementary material

Supplementary material is available at https://journals.viamedica.pl/polish_heart_journal.

Article information

Conflict of interest: None declared.

Funding: None.

Open access: This article is available in open access under Creative Commons Attribution-Non-Commercial-No Derivatives 4.0 International (CC BY-NC-ND 4.0) license, which allows downloading and sharing articles with others as long as they credit the authors and the publisher, but without permission to change them in any way or use them commercially. For commercial use, please contact the journal office at polishheartjournal@ptkardio.pl

REFERENCES

1. Thygesen K, Alpert JS, Jaffe AS, et al. Fourth universal definition of myocardial infarction. *Eur Heart J.* 2019;40(3):237–269, doi: [10.1093/eurheartj/ehy462](https://doi.org/10.1093/eurheartj/ehy462), indexed in Pubmed: [30165617](https://pubmed.ncbi.nlm.nih.gov/30165617/).
2. Alkarithi G, Duval C, Shi Y, et al. Thrombus structural composition in cardiovascular disease. *Arterioscler Thromb Vasc Biol.* 2021;41(9):2370–2383, doi: [10.1161/ATVBAHA.120.315754](https://doi.org/10.1161/ATVBAHA.120.315754), indexed in Pubmed: [34261330](https://pubmed.ncbi.nlm.nih.gov/34261330/).
3. Ząbczyk M, Ariëns RAS, Undas A. Fibrin clot properties in cardiovascular disease: from basic mechanisms to clinical practice. *Cardiovasc Res.* 2023;119(1):94–111, doi: [10.1093/cvr/cvad017](https://doi.org/10.1093/cvr/cvad017), indexed in Pubmed: [36662542](https://pubmed.ncbi.nlm.nih.gov/36662542/).
4. Carnevale R, Sciarretta S, Valenti V, et al. Low-grade endotoxaemia enhances artery thrombus growth via Toll-like receptor 4: Implication for myocardial infarction. *Eur Heart J.* 2020;41(33):3156–3165, doi: [10.1093/eurheartj/ehz893](https://doi.org/10.1093/eurheartj/ehz893), indexed in Pubmed: [31898723](https://pubmed.ncbi.nlm.nih.gov/31898723/).
5. Violi F, Nocella C, Bartimoccia S, et al. Gut dysbiosis-derived low-grade endotoxemia: A common basis for liver and cardiovascular disease.

- Kardiol Pol. 2023; 81(6): 563–571, doi: [10.33963/KP.a2023.0115](https://doi.org/10.33963/KP.a2023.0115), indexed in Pubmed: [37191190](https://pubmed.ncbi.nlm.nih.gov/37191190/).
6. Vaure C, Liu Y. A comparative review of toll-like receptor 4 expression and functionality in different animal species. *Front Immunol.* 2014; 5: 316, doi: [10.3389/fimmu.2014.00316](https://doi.org/10.3389/fimmu.2014.00316), indexed in Pubmed: [25071777](https://pubmed.ncbi.nlm.nih.gov/25071777/).
 7. Sadowski M, Ząbczyk M, Undas A. Coronary thrombus composition: links with inflammation, platelet and endothelial markers. *Atherosclerosis.* 2014; 237(2): 555–561, doi: [10.1016/j.atherosclerosis.2014.10.020](https://doi.org/10.1016/j.atherosclerosis.2014.10.020), indexed in Pubmed: [25463088](https://pubmed.ncbi.nlm.nih.gov/25463088/).
 8. Zalewski J, Bogaert J, Sadowski M, et al. Plasma fibrin clot phenotype independently affects intracoronary thrombus ultrastructure in patients with acute myocardial infarction. *Thromb Haemost.* 2015; 113(6): 1258–1269, doi: [10.1160/TH14-09-0801](https://doi.org/10.1160/TH14-09-0801), indexed in Pubmed: [25739375](https://pubmed.ncbi.nlm.nih.gov/25739375/).
 9. Violi F, Cammisotto V, Bartimoccia S, et al. Gut-derived low-grade endotoxaemia, atherothrombosis and cardiovascular disease. *Nat Rev Cardiol.* 2023; 20(1): 24–37, doi: [10.1038/s41569-022-00737-2](https://doi.org/10.1038/s41569-022-00737-2), indexed in Pubmed: [35840742](https://pubmed.ncbi.nlm.nih.gov/35840742/).
 10. Nunes JM, Fillis T, Page MJ, et al. Gingipain R1 and lipopolysaccharide from have major effects on blood clot morphology and mechanics. *Front Immunol.* 2020; 11: 1551, doi: [10.3389/fimmu.2020.01551](https://doi.org/10.3389/fimmu.2020.01551), indexed in Pubmed: [32793214](https://pubmed.ncbi.nlm.nih.gov/32793214/).
 11. Ząbczyk M, Kruk A, Natorka J, et al. Low-grade endotoxemia in acute pulmonary embolism: Links with prothrombotic plasma fibrin clot phenotype. *Thromb Res.* 2023; 232: 70–76, doi: [10.1016/j.thromres.2023.10.020](https://doi.org/10.1016/j.thromres.2023.10.020), indexed in Pubmed: [37949000](https://pubmed.ncbi.nlm.nih.gov/37949000/).
 12. Sadowski M, Ząbczyk M, Undas A. Impaired fibrinolysis in patients with atrial fibrillation and elevated circulating lipopolysaccharide. *J Thromb Thrombolysis.* 2024; 57(5): 842–851, doi: [10.1007/s11239-024-02980-5](https://doi.org/10.1007/s11239-024-02980-5), indexed in Pubmed: [38643439](https://pubmed.ncbi.nlm.nih.gov/38643439/).
 13. Carlin S, de Vries TAC, Budaj A, et al. Dual pathway inhibition for atherosclerotic cardiovascular disease: Recent advances. *Kardiol Pol.* 2022; 80(12): 1200–1210, doi: [10.33963/KP.a2022.0283](https://doi.org/10.33963/KP.a2022.0283), indexed in Pubmed: [36601884](https://pubmed.ncbi.nlm.nih.gov/36601884/).
 14. Loffredo L, Ivanov V, Ciobanu N, et al. Low-grade endotoxemia and NOX2 in patients with coronary microvascular angina. *Kardiol Pol.* 2022; 80(9): 911–918, doi: [10.33963/KP.a2022.0130](https://doi.org/10.33963/KP.a2022.0130), indexed in Pubmed: [35579023](https://pubmed.ncbi.nlm.nih.gov/35579023/).