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ISSN: 2451-2591

e-ISSN: 2451-4101

Identification of molecular mechanisms of association amongst comorbidities and COVID-19: An Interactome based Systems Biology Approach

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DOI: 10.5603/mrj.102212

Article type: Original article

Submitted: 2024-08-22

Accepted: 2024-09-09

Published online: 2024-11-14

This article has been peer reviewed and published immediately upon acceptance.

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ORIGINAL ARTICLE

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Identification of molecular mechanisms of association amongst comorbidities and COVID-19: An Interactome based Systems Biology Approach

Short title: Tammanna R. Sahrawat, Association amongst comorbidities and COVID-19: Interactome study

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DOI: 10.5603/mrj.102212

ABSTRACT

Background: Comorbidity has emerged as a major challenge in the last few decades that result from cascades of failures involving complex biochemical and physical interactions of genes, proteins, and metabolites responsible for cellular functions. Various epidemiological and demographic studies conducted since the emergence and global transmission of the SARS-CoV-2 virus have reported that patients with pre-existing medical conditions such as cardiovascular disease, diabetes, hepatitis, lung disease, and kidney disease are more prone to coronavirus infection.

Objective: The present study was undertaken to elucidate the molecular mechanisms that are common amongst COVID-19-associated comorbidities using an interactome-based network biology approach to identify the shared genes/proteins and biological pathways.

Methodology: Genes of COVID-19-associated comorbidity diseases retrieved from disease databases were analyzed using *in silico* bioinformatics and systems network biology tools STRING and Cytoscape plug-ins CytoHubba and CytoCluster.

Results: The shared hub proteins, namely *IL1B*, *ACTB*, *IL6*, *MMP2*, *ALB*, *AKT1*, *MAPK3*, *FN1*, *TNF*, *CCL2*, *VEGFA*, and *TP53*, among various pre-existing comorbidities, revealed their involvement in immunological pathways. All these proteins were also found to have significant associations with ACE2, TMPRSS2, and CD17/BSG, the entry receptors of the COVID-19 virus.

Conclusion: The higher risk factor for COVID-19 in patients with pre-existing comorbidities is due to immune dysfunction that results in their higher susceptibility to infection by SARS-CoV-2 via its entry receptors on the host cells. This study provides novel insights into the association between host genetics and the consequences of viral infection that is responsible for the severity of COVID-19 in patients suffering from pre-existing comorbidities. **Keywords:** Comorbidity, SARS-CoV-2, systems network biology, interactome, COVID-19, *in silico*, hub genes, Cytokine storm (CS), Immune dysfunction

Introduction

A major challenge that has emerged in the last few decades is comorbidity, which refers to the presence of more than one disease in individuals. Disease comorbidity effects result from interactions between molecular components, which in turn may affect other cellular functions as well as mutated gene products.

Several epidemiological and clinical studies have reported that patients with preexisting medical conditions have severe responses to the coronavirus infection. The highest risk factor for infection of COVID-19 resulting in extreme lung injury followed by death is in patients having comorbidities such as cardiovascular disease, diabetes, hepatitis, lung disease, and kidney disease, who may develop [1–3]. The strongest predictors of the prognosis of COVID-19 patients have been shown to be age, gender, and some pre-existing comorbidities. The higher risk group is \geq 60 years, and those with certain hidden conditions, for example, cardiovascular and cerebrovascular sicknesses and diabetes [4].

Epidemiological studies on COVID-19-affected patients revealed that age factors along with comorbidities such as diabetes, lung infection, renal impairment, and liver damage should be taken into consideration, as a substantial number of old patients showed a higher death rate with better survival rates for younger patients infected with COVID-19 [5, 6]. Earlier studies published in 2003 had reported that in patients infected with SARS, acute coronary syndrome and myocardial infarction cause 2 deaths per fifth patient [7, 8]. The principal risk factors for COVID-19 infection are acute respiratory failure and alveolar failure. The target site of SARS is the respiratory tract of an individual, causing mild upper respiratory illness followed by acute respiratory distress with septic shock and a higher infectious rate leading to pulmonary fibrosis by binding to the transmembrane receptor Angiotensin-Converting Enzyme 2 (ACE2) on the respiratory epithelium [9]. The presence of the viral genome of SARS in the hepatic tissue's endothelial cells causes multiple organ failures, including liver failure, which can be attributed to the widely distributed ACE2 receptors that are responsible for the entry of SARS-CoV [10]. The elevated levels of ALT and AST and pro-inflammatory cytokines were reported in liver-damaged patients, and patients with a history of hepatitis were at higher risk of COVID-19 exhibiting exaggerated replication during SARS-CoV infection [11].

It is related to other organ harm, for example, cardiovascular damage by means of an increased risk of hypertension through ACE2, gastrointestinal dysfunctionality, diabetes mellitus, liver brokenness, ongoing kidney illness, CNS damages, lung injury, visual dangers like conjunctival hyperemia, chemosis, conjunctivitis, and malignant growth hazard, venous thromboembolism, tuberculosis, maturing, and conceptive danger [12].

A recent study based on an integrated network biology approach reported cancers, pulmonary, hepatic, neurological, hypertensive, and cardiac disorders being linked to COVID-19 along with multiple organ damage [13]. A bioinformatics analysis revealed the genes and pathways were significantly enriched for immune system and cardiovascular-related phenotypes in COVID-19 comorbidities [14]. Another study based on computational methods reported 274 differentially expressed genes common in both COVID-19 patients and patients with comorbidities like cardiovascular disease, diabetes, and obesity, suggesting their role in the increased severity of COVID-19 [15].

Patients with certain pre-existing medical conditions have higher chances of being afflicted with the SARS-CoV-2 virus, followed by severe responses as reported following the emergence and global transmission of COVID-19. Therefore, there arises a need to study disease-disease relationships to better understand the human interactome and molecular causes of disease comorbidities. However, reports on the molecular factors responsible for the association of COVID-19 with pre-existing comorbidities are not well understood and

need to be deciphered for the design of preventive strategies. Therefore, this study was undertaken to identify the underlying common molecular mechanism of association of genes/proteins and biological pathways amongst various disease comorbidities and COVID-19 using a systems biology-based interactome approach. This study would give meaningful insights into understanding the etiopathology and the molecular association of disease comorbidity and COVID-19.

Material and methods

Data Mining

Genes reported to be associated with diabetes mellitus, cardiovascular diseases, pulmonary fibrosis, kidney diseases, hepatitis C, and hepatitis B were retrieved from the DisGeNet database (http://www.disgenet.org) [16], and genes common amongst these six diseases were obtained using PHP programming.

Construction and analysis of interactome

STRING database

A protein-protein interaction (PPI) network was constructed using the STRING database (Search Tool for the Retrieval of Interacting Genes/Proteins) database (https://string-db.org/) that performs analysis based on known and predicted PPIs, including physical and functional associations [17] for the genes found common amongst the six diseases.

Cytoscape

Network analysis was performed using Cytoscape, an open-source software project that integrates expression data with biomolecular interaction networks and molecular states to form a unified conceptual framework [18].

ClusterONE (Clustering with Overlapping Neighborhood Expansion) plugin of Cytoscape (http://apps.cytoscape.org/apps/clusterone) was used for the identification of highly connected regions in the form of clusters [19].

CytoHubba plugin of Cytoscape (http://apps.cytoscape.org/apps/cytohubba) was used for ranking the hub genes using the MCC (maximal clique centrality) scoring method [20], was used for the analysis of the interactome.

Reactome knowledgebase

The hub genes were mapped into their respective biomolecular pathways using Reactome (https://reactome.org), which is a peer-reviewed knowledge base that captures information about genes and molecules involved in pathways [21].

Results

Genes involved in diabetes mellitus (3134), cardiovascular diseases (1756), pulmonary fibrosis (924), kidney diseases (1180), hepatitis C (1768), and hepatitis B (1449) were retrieved from the DisGeNET database. One hundred and ten common genes among all six diseases were shortlisted.

Network construction of the 110 genes common amongst the diseases (Table 1) was done using the STRING database, and only 107 genes/proteins were present in *Homo sapiens*, which were used for the construction of an interaction network having 107 nodes and 1989 edges that represent genes and their interactions, respectively (Fig. 1).

On analysis of the network obtained from STRING using Cytoscape plug-in ClusterONE, four clusters were obtained, out of which only one cluster had a significant p-value (< 0.05) (Fig. 2 A & B). This network was analyzed with CytoHubba to identify hub nodes that represent highly connected nodes having a higher likelihood of being involved in an essential interaction [22, 23]. The top 50 ranked genes were obtained, where red, yellow, and gray nodes represent highly, least significant genes, and outliers, respectively (Fig. 3).

Twenty common genes were found to be significant, namely *IL6*, *TNF*, *IL1B*, *AKT1*, *ACTB*, *CXCL8*, *CCL2*, *VEGFA*, *ALB*, *TP53*, *IL10*, *MAPK3*, *FN1*, *PTGS2*, *STAT3*, *PPARG*, *IL4*, *TLR4*, *IFNG*, and *MMP2*, amongst the six diseases, *i.e.*, cardiovascular disease, diabetes mellitus, hepatitis, pulmonary fibrosis, and kidney disease. A network of these 20 hub genes was constructed in STRING, and the resulting network had 190 edges and an average node degree of 19, which indicates the number of interactions (at the score threshold) that a protein has on average in the network (Fig. 4).

The hub genes/proteins common among the comorbidities were found to be enriched in 500 biological pathways (FDR \leq 0.05) using the Reactome knowledgebase, with most of them being associated with the immune system, signal transduction, disease, and transcription-associated pathways (Table 2).

Infection of host cells by SARS-CoV-2 has been reported to require the presence of entry receptors, namely transmembrane receptors ACE2 (Angiotensin-Converting Enzyme 2), TMPRSS2 (Type II Transmembrane Serine Protease), and CD147 (cluster of differentiation 147)/EMMPRIN (Extracellular matrix metalloproteinase inducer)/BSG (Basigin) [24–26]. A study proposed that CD147 and ACE2 allow entry of the virus into the cell cytoplasm by sequential activation of NLRP3 inflammasome, resulting in cleavage of interleukins IL-1β and IL-18 [24] followed by binding of SARS-CoV-2 to ACE2. Subsequently, TMPRSS2 primes the SARS-CoV-2 spike protein that allows membrane fusion and its entry into the host cell [24, 27].

To identify the association of pre-existing disease comorbidities with SARS-CoV-2, an interaction network of the 20 hub proteins identified with the SARS-CoV-2 entry receptors, namely ACE2, TMPRSS2, and CD147/BSG, was constructed in STRING (Fig. 5).

In the present study, SARS-CoV-2 entry receptors TMPRSS2, ACE2, and CD147/BSG showed interactions with 2 (excluding ACE2), 8 (excluding TMPRSS2), and 7 hub proteins, respectively, that were common amongst the comorbidities (Table 3).

From the analysis of the network, it is observed that out of the 20 hub proteins only 12 proteins, *i.e.*, IL1B, ACTB, IL6, MMP2, ALB, AKT1, MAPK3, FN1, TNF, CCL2, VEGFA, and TP53, have significant interactions with the SARS-CoV-2 entry receptors. Each of these 12 proteins was studied using literature mining to understand their association with SARS-CoV-2 infection, given their significant number of interactions with SARS-CoV-2 entry receptors TMPRSS2, ACE2, and CD147/BSG [24-26].

Discussion

Protein kinase B alpha (PKBalpha/Akt-1) was found to interact with all the 11 proteins previously shortlisted and has significant interactions with all the SARS-CoV-2 entry receptors. AKT1 is reported to be involved in the regulation of cell proliferation and transformation thereby playing an essential role in immune cell modulation, resulting in diabetes and cardiovascular diseases, among others [28–30]. Through its downstream targets GSK3 and GLUT4, AKT increases cellular metabolism, and alterations in AKT signaling affect various cardiovascular pathological processes such as atherosclerosis [31]. During the progression of acute kidney injury to chronic kidney disease, the renal fibrosis and tubular dedifferentiation observed have been attributed to AKT, along with its role in increasing the oncogenic potential of the Hepatitis B virus X protein by phosphorylation [32, 33]. AKT has been reported to affect the progression and severity of COVID-19 along with IL10, TNF, and other proteins in non-alcoholic fatty liver disease patients [34]. In COVID-19/asthma comorbidity, AKT1 along with TP53, ALB, IL-6, TNF, and VEGFA have been reported to be hub proteins [35].

Levels of proinflammatory cytokines and chemokines such as TNF- α , IL-6, and IL-1 β have been found to be elevated in cytokine storm (CS), which is a hallmark of COVID-19 pathogenesis [36, 37]. TNF can induce cells to release cytokines and evoke various intracellular signaling pathways, such as inflammation, apoptosis, programmed cell necrosis, and immunity [38, 39]. IL-6 has been reported to be associated with the host's response to infection, which affects the viral load and severity of the disease in a patient [40]. Therefore, CS is responsible for the escalation of disease severity resulting in septic shock and multiorgan failure in COVID-19 patients with comorbidities [41].

Mitogen-activated protein kinase (MAPK3) plays an essential role in viral infections and other important cellular physiological and pathological processes as it regulates cell growth, differentiation, and inflammatory response. Studies have also shown reprogramming caused by tumor therapy is due to a link between MAPK signals and control of the inflammatory network [42, 43].

Matrix metalloproteinases (MMPs) are involved in the release of substrates such as growth factors that are anchored at the extracellular matrix or cell membrane and also contribute to injurious processes in lung pathologies. Following SARS-CoV-2 infection, clinical studies report an increase in plasma MMPs, suggesting their release by the host's immune system and target cells [44]. Fibronectin (FN) is a glycoprotein that is present in high concentrations in blood and is a component of the extracellular matrix. In critically ill COVID-19 patients, it has been reported as a disease marker of severity and also found to be associated with different kinds of inflammatory diseases [45]. Actin cytoplasmic 1 (ACTB) is a highly conserved protein found in the cytoplasm and a component of the cytoskeleton. It is involved in the proper functioning of the immune system, cell motility, and division, and in COVID-19 patients it had a low expression, resulting in immune cell infiltration and altered immune function [46].

In the present study, association of shared hub genes/proteins, namely *IL1B*, *ACTB*, *IL6*, *MMP2*, *ALB*, *AKT1*, *MAPK3*, *FN1*, *TNF*, *CCL2*, *VEGFA*, and *TP53*, among various preexisting comorbidities (cardiovascular disease, diabetes mellitus, hepatitis, pulmonary fibrosis, and kidney disease) with COVID-19 have been identified, which were found to be directly involved in immunological responses. Moreover, each of these hub proteins was found to have significant associations with ACE2, TMPRSS2, and CD17/BSG, which are the entry receptors of the COVID-19 virus. People suffering from pre-existing comorbidities have impaired immune functions, making them more likely to contract viral infections, including SARS-CoV-2. The genes identified can be further explored to develop preventive therapeutic interventions for patients with pre-existing comorbidities to contain the severity of SARS-CoV-2 and decrease morbidity and mortality rates.

Conclusions

This study provides novel insights into the associated molecular mechanisms in preexisting comorbidities with COVID-19 using a robust interactome-based systems biology approach. Controlling the inflammatory response proteins is a common critical factor amongst the six disease co-morbidities studied and the cytokine storm, which is the hallmark of COVID-19. The higher risk factors for COVID-19 in patients with pre-existing comorbidities are associated with immune dysfunction, and two proteins, AKT1 and TNF, identified in the present study having multiple interactions with the entry receptors of SARS-CoV-2, can be used as biomarkers and explored further for the design of potential therapeutic strategies. A synergistic approach is needed to block pathologies at multiple levels using therapeutic interventions to inhibit viral infection and regulate the dysfunctional immune responses to alleviate the severity of COVID-19 in patients having pre-existing comorbidities.

Article information

Funding: The author would like to acknowledge the funding received under grant no.
S&T&RE/Rp/147/FY(21-22) AA/10/ 2021/ 1013-1020 from Department of Science and Technology and Renewable Energy, UT, Chandigarh, India, under the short-term research project scheme.
Acknowledgments: The author would like to thank their parent institute Panjab University Chandigarh, India for providing the infrastructure for carrying out the research.
Conflict of interest: The author declares there is no conflict of interest.
Ethics approval and consent to participate: The study did not involve human subjects or animal models and therefore no ethical clearance was required.

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CDKN2AAHSA1MTCO2P12*PPARGC1ACHI3L1NLRP3CNR1RBM45SLCO6A1COX8ACRKMAPK14CCN2CTNNB1ACEAGTEGFRAKT1ESR1ALBF2FN1SIRT1BRD4FOSMTORGABPARNF19APOLDIP2PTPN22IL37CXCR3ANGPT2GPTANXA1HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2NEE2L2NEKP1NOTCH1SEPDINE1APCP1	5				
CRKMAPK14CCN2CTNNB1ACEAGTEGFRAKT1ESR1ALBF2FN1SIRT1BRD4FOSMTORGABPARNF19APOLDIP2PTPN22IL37CXCR3ANGPT2GPTANXA1HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	CDKN2A	AHSA1	MTCO2P12*	PPARGC1A	CHI3L1
AGTEGFRAKT1ESR1ALBF2FN1SIRT1BRD4FOSMTORGABPARNF19APOLDIP2PTPN22IL37CXCR3ANGPT2GPTANXA1HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	NLRP3	CNR1	RBM45	SLCO6A1	COX8A
F2FN1SIRT1BRD4FOSMTORGABPARNF19APOLDIP2PTPN22IL37CXCR3ANGPT2GPTANXA1HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	CRK	MAPK14	CCN2	CTNNB1	ACE
MTORGABPARNF19APOLDIP2PTPN22IL37CXCR3ANGPT2GPTANXA1HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	AGT	EGFR	AKT1	ESR1	ALB
IL37CXCR3ANGPT2GPTANXA1HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	F2	FN1	SIRT1	BRD4	FOS
HLA-DRB1HMGB1HMOX1HSPA1BHSPA4APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	MTOR	GABPA	RNF19A	POLDIP2	PTPN22
APOA1ICAM1IFNGIGF1APRTIL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	IL37	CXCR3	ANGPT2	GPT	ANXA1
IL1AIL1BIL1RNIL4IL6CXCL8IL10IL17AIL18GSTK1LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	HLA-DRB1	HMGB1	HMOX1	HSPA1B	HSPA4
CXCL8 IL10 IL17A IL18 GSTK1 LEP LGALS3 MIR145 MIR146A* MIR155* MIR21 MFAP1 MMP2 MMP14 COX2	APOA1	ICAM1	IFNG	IGF1	APRT
LEPLGALS3MIR145MIR146A*MIR155*MIR21MFAP1MMP2MMP14COX2	IL1A	IL1B	IL1RN	IL4	IL6
MIR21 MFAP1 MMP2 MMP14 COX2	CXCL8	IL10	IL17A	IL18	GSTK1
	LEP	LGALS3	MIR145	MIR146A*	MIR155*
NEEDID NEKRI NOTCHI SEDDINEI ABCRI	MIR21	MFAP1	MMP2	MMP14	COX2
NFE2EZ NFRDI NOTCHI SERFINEI ADCDI	NFE2L2	NFKB1	NOTCH1	SERPINE1	ABCB1
PIK3CG PPARG PRKAA1 MAPK1 MAPK3	PIK3CG	PPARG	PRKAA1	MAPK1	МАРКЗ
MAPK8 PTGS2 RELA ACTB CCL2	MAPK8	PTGS2	RELA	ACTB	CCL2
CCL5 GORASP1 WNK1 SOAT1 SPP1	CCL5	GORASP1	WNK1	SOAT1	SPP1
STAT3 STAT4 SYT1 ADAM17 TERT	STAT3	STAT4	SYT1	ADAM17	TERT
TGFB1 THBS1 TLR2 TLR4 TNF	TGFB1	THBS1	TLR2	TLR4	TNF
TNFRSF1A TP53 C3 VDR VEGFA	TNFRSF1A	TP53	C3	VDR	VEGFA
VTN VWF CXCR4 AIMP2 CAT	VTN	VWF	CXCR4	AIMP2	CAT
CAV1 SOCS1 TNFSF10 GRAP2 GDF15	CAV1	SOCS1	TNFSF10	GRAP2	GDF15
*Genes in grey boxes are not present in <i>Homo sapiens</i>					

 Table 1. Genes common amongst the diseases Diabetes mellitus, Cardiovascular diseases,
 Pulmonary fibrosis, Kidney diseases, Hepatitis C and B

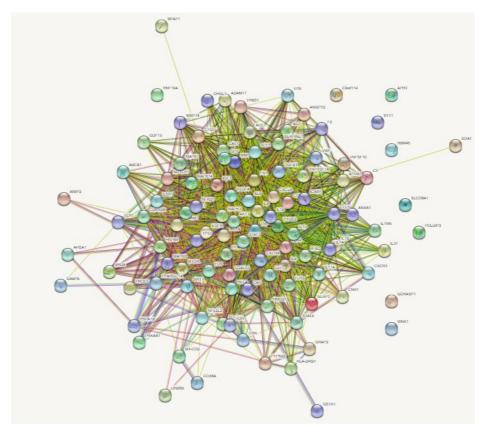


Figure 1. Protein-protein interaction network of common genes amongst six diseases obtained from STRING containing 107 nodes and 1989 edges

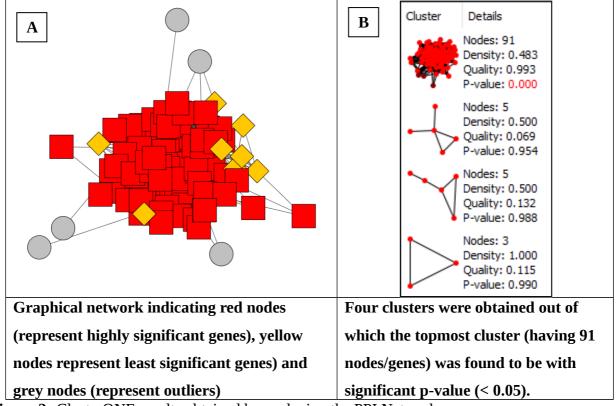


Figure 2. ClusterONE results obtained by analyzing the PPI Network

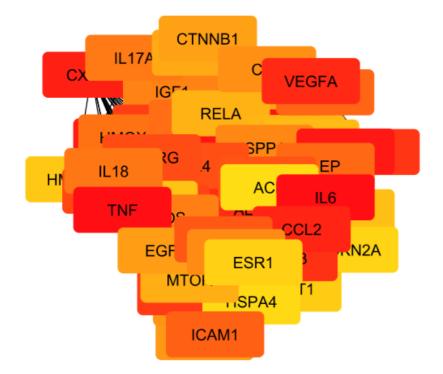


Figure 3. Graphical view of ranked hub nodes obtained from Cytoscape plug-in Cytohubba with color coding

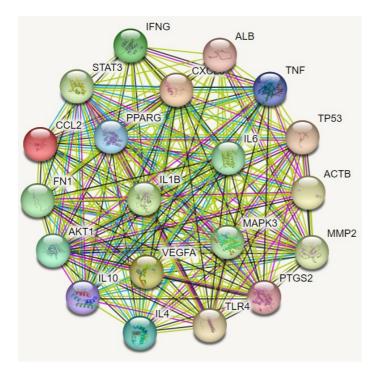


Figure 4. Protein-Protein interaction network of 20 hub genes (nodes) from STRING having 190 edges responsible for co-morbidity of the six diseases

Gene **Pathway name** p-value FDR Genes in pathway count Interleukin-10 1.11E-IL10; IL6; CXCL8; IL1B; STAT3; 1.28E-14 8 signaling 16 CCL2; PTGS2; TNF Interleukin-4 IL10; CXCL8; MMP2; STAT3; 1.11Eand interleukin-FN1; PTGS2; TNF; VEGFA; IL4; 1.28E-14 14 16 IL6; IL1B; CCL2; AKT1; TP53 13 signaling IL10; CXCL8; MMP2; STAT3; Signaling by FN1; PTGS2; TNF; VEGFA; IL4; 1.11E-16 1.28E-14 interleukins 16 IL6; IFNG; IL1B; CCL2; AKT1; *TP53; MAPK3* IL10; CXCL8; MMP2; STAT3; Cytokine 1.11E-FN1; PTGS2; TNF; VEGFA; IL4; signaling in the 1.28E-14 16 16 IL6; IFNG; IL1B; CCL2; AKT1; immune system *TP53; MAPK3* IL10; CXCL8; MMP2; STAT3; FN1; PTGS2; TNF; ACTB; 1.11E-Immune system 1.28E-14 18 VEGFA; IL4; IL6; IFNG; IL1B; 16 CCL2; AKT1; TLR4; TP53; MAPK3 Senescenceassociated 8.17E-IL6; CXCL8; IL1B; STAT3; 7.84E-07 5 secretory 09 MAPK3 phenotype (SASP) Cellular IL6; CXCL8; IL1B; ALB; STAT3; 7.68E-6.29E-06 CCL2; AKT1; TLR4; TP53; 11 responses to 08 stress MAPK3; VEGFA Cellular IL6; CXCL8; IL1B; ALB; STAT3; 9.68Eresponses to 6.52E-06 11 CCL2; AKT1; TLR4; TP53; 08 stimuli MAPK3; VEGFA Cellular IL6; CXCL8; IL1B; STAT3; TP53; 1.02E-6.52E-06 6 senescence 07 МАРКЗ Signaling by 6.35E-STAT3; FN1; AKT1; ACTB; 0.0012 6 receptor 05 MAPK3; VEGFA tyrosine kinases

Table 2. The top 15 most significantly enriched pathways obtained from Reactome

Signal transduction	0.0012	0.0107	12	CXCL8; MMP2; STAT3; FN1; TNF; ACTB; VEGFA; IL6; AKT1; CCL2; PPARG; TP53; MAPK3
Disease	0.0013	0.0118	11	IL10; IL6; IL1B; STAT3; FN1; AKT1; TLR4; TP53; ACTB; MAPK3; VEGFA
Infectious disease	0.0023	0.0172	7	IL10; IL6; IL1B; AKT1; ACTB; MAPK3; VEGFA
Gene expression (transcription)	0.0024	0.0172	8	IL6; IFNG; AKT1; PPARG; TP53; ACTB; MAPK3; VEGFA

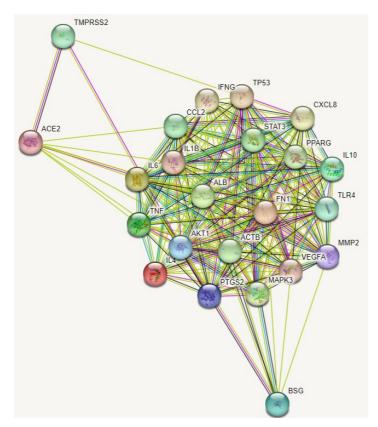


Figure 5. Interaction network of hub proteins common amongst comorbidities with SARS-CoV-2 entry receptors constructed in STRING

SADS CoV 2 on two recontors	TMPRSS		CD147/BS
SARS-CoV-2 entry receptors	2	ACE2	G
JIS	AKT-1	AKT-1	AKT-1
btc		ACTB	ACTB
l ece		FN1	FN1
Å r		ALB	ALB
	TP53	TNF	VEGFA
5 6		IL6	MAPK3
		ILIB	MMP2
C			
-S.			
AF			
e N			
- Ē			
l tit			
Cti,			
l l l l l l l l l l l l l l l l l l l			
Cal			
1			
20 0			
l vii			
ha			
Hub proteins having significant interactions with the SARS-CoV-2 entry receptors			
X			
ā			
Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.Ξ.		CCL2	

Table 3. SARS-CoV-2 entry receptors having direct interactions with hub proteins of comorbidities