Does endocan level increase in women with polycystic ovary syndrome? A case — control study

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ABSTRACT

Objectives: To evaluate endocan levels of patients with polycystic ovary syndrome (PCOS) in comparison to healthy women.

Material and methods: A cross-sectional case-control study on 88 patients with PCOS (mean age, 22.06 ± 4.24 years; body mass index [BMI], 23.9 ± 4.74 kg/m²) and 87 age- and BMI-matched healthy women (mean age, 23.71 ± 4.42 years; BMI, 22.15 ± 3.03 kg/m²).

Results: Serum endocan level was significantly higher in PCOS group than control group (540.9 ± 280.3 pg/mL vs. 355.5 ± 233.5 pg/mL, respectively; p < 0.001). The presence of polycystic ovary finding on ultrasonography or oligomenorrhea did not produce significant effect on serum endocan levels (p > 0.05). In PCOS group, endocan level was negatively correlated with BMI and C-reactive protein level, and positively correlated with high density lipoprotein level (p < 0.05).

Conclusions: Blood endocan level is increased in PCOS. Further studies are needed to evaluate the clinical value of blood endocan level as a marker for the risk of cardiovascular and metabolic diseases in patients with PCOS.

Key words: endocan, polycystic ovary syndrome, inflammation, case-control

INTRODUCTION

Polycystic ovary syndrome (PCOS) affects around 7% of women and is characterized by hyperandrogenism with or without oligo-anovulation [1]. The clinical signs and symptoms of PCOS change from puberty to perimenopausal period of patients. In addition to cosmetic complaints such as hirsutism and acne, patients with PCOS may have obesity, menstrual irregularity, and infertility. Increased levels of androgens in PCOS are associated with metabolic diseases including excess fat accumulation, obesity, and insulin resistance [1].

In patients with PCOS, the risk of cardiovascular and metabolic diseases, primarily diabetes, has increased [2–6]. For this reason, it is important not only to manage PCOS in terms of patient’s specific complaints, but also to define potential risk of cardiovascular and metabolic diseases associated with this syndrome, and to inform patients about this risk. Previous studies have reported an inflammatory process underlying PCOS, which is associated with metabolic diseases, hyperandrogenism, insulin resistance, abdominal obesity, and endothelial damage [2–8]. Endothelium dysfunction, a chronic inflammatory process which is an early sign of atherosclerosis, has been demonstrated in studies on PCOS [8–10].

Endocan, endothelial cell-specific molecule (ESM-1), is a soluble 50 kDa molecular weight proteoglycan expressed in vascular endothelium [11]. It has been reported that endocan levels increase in many cancers, sepsis, obesity, and a variety of inflammatory pathologies by playing a critical role in vascular injury in organ-specific pathologies and being a potential novel marker in endothelium-dependent pathologic disorders [11, 12]. Although endothelial dysfunction in PCOS has been extensively studied, studies on the relationship between PCOS and endocan are limited [13, 14].
Therefore, in this study, we aimed to evaluate endocan levels of patients with PCOS in comparison to healthy women in order to clarify the relationship between PCOS and endothelial injury.

**MATERIAL AND METHODS**

**Study design and population**

This was a cross-sectional case-control study conducted between January 2017 and April 2017 in the Department of Obstetrics and Gynecology, Gaziosmanpasa University Hospital, Tokat, Turkey. Eighty-eight patients with PCOS (mean age, 22.06 ± 4.24 years; body mass index [BMI], 23.9 ± 4.74 kg/m²) and 87 age- and BMI-matched subjects (mean age, 23.71 ± 4.42 years; BMI, 22.15 ± 3.03 kg/m²) with normal menstrual cycles who presented to our outpatient clinic were included into the study. The selection criteria for the patients with PCOS were the absence of any significant abnormalities on physical examination except hirsutism; no lipid lowering, hypoglycemic, antihypertensive or hormone replacement therapy; normal thyroid function and prolactin level; absence of history or evidence of metabolic, cardiovascular, respiratory or hepatic disease. The diagnosis of PCOS was made in accordance with the criteria proposed at the Rotterdam revised consensus meeting (The Rotterdam European Society for Human Reproduction and Embryology [ESHRE]/American Society for Reproductive Medicine [ASRM]-Sponsored PCOS Consensus Workshop Group, 2004) [15]. All eligible patients met at least 2 of the 3 following criteria: [a] oligomenorrhea or amenorrhea, [b] clinical and/or biochemical signs of hyperandrogenism (hirsutism and acne), and [c] polycystic ovaries. Subjects with possible ovarian tumors, congenital adrenal hyperplasia or BMI greater than 30 kg/m² were excluded from the study. All patients and control subjects were non-smoker, normotensive (< 120/80 mm Hg in two measurements), had regular daily activity and not regular consumer of alcoholic beverages. None of the subjects were pregnant during the study procedures.

The study was approved by the Clinical Research Ethics Committee of Gaziosmanpasa University School of Medicine (16-KAEK-038), and all patients and control subjects gave written informed consent before enrollment. The study was conducted according to the principles of the latest version of Helsinki Declaration.

**Study procedures**

All the participants were evaluated in one study visit, during which data on medical history, physical examination including anthropometric measurements, laboratory test, and pelvic ultrasonography (US) were recorded. BMI (kg/m²) was used as an estimate of overall adiposity in both groups. To evaluate the severity of hirsutism, hirsutism score, which includes clinical signs of hyperandrogenism was used. A hirsutism score of more than 7 was considered as severe hirsutism [16]. The presence of PCO on US was recorded. All the measurements were made by the same physician.

For laboratory tests, venous blood samples were drawn on the third to fifth day of menstrual cycle between 08:00 and 10:00 a.m. after a 12-hour fasting. Glucose, total cholesterol, triglyceride, high-density lipoprotein (HDL)-cholesterol, and C-reactive protein (CRP) levels were measured by enzymatic colorimetric method using Olympus AU 600 auto analyzer (Olympus Diagnostics, GmbH, Hamburg, Germany). Low-density lipoprotein (LDL)-cholesterol level was estimated by the Friedewald formula [LDL-cholesterol = (totalcholesterol)–(HDL-cholesterol)–(triglyceride)/5]. Luteinizing hormone (LH), follicle stimulating hormone (FSH), estradiol (E2), total testosterone, dehydroepiandrosterone sulphate (DHEA-S), and insulin were measured in serum by chemiluminescence, using Modular E-170 Immunologic Analyzer (Roche Diagnostics, Osaka, Japan).

**Blood endocan level measurements**

The serum samples for endocan, i.e. ESM-1, were first stored for coagulation and then centrifuged for 15 min at 4000 xg at + 4°C. The serum samples obtained divided into aliquots and stored in a deep freeze at - 80°C until analysis. ESM-1 levels in serum samples were measured using ELISA with a “Human ESM-1 ELISA Kit” (Elabscience Biotechnology Co. Ltd., Lot: AK0016DEC07042, Wuhan, China). It uses a double-antibody sandwich enzyme-linked immunosorbent assay. Samples like serum and standards were pipetted into the 96 well microplate being coated on with a monoclonal antibody (also known as Capture Antibody that is specific for C terminal of human ESM-1), and incubated for 90 min. ESM-1 present within a sample is bound by the Capture Antibody. After washing away of any unbound molecules, a secondary monoclonal antibody specific for N terminal of ESM-1 that has been biotinylated, was added to the wells to incubate for another 1 hour. After a washing step, streptavidin-HRP (biotin-binding protein conjugated with polymers of horseradish peroxidase) was added and allowed to incubate for 30 min. Unbound material was washed away. Chromogen solution was added and incubated for 15 min (protected from light) for the conversion of the colorless solution to blue solution, the intensity of which was proportional to the amount of ESM-1 in the sample. As an effect of the acidic stop solution, the color has become yellow. The colored reaction product was measured using an automated ELISA reader at 450 nm. The results were expressed as picograms per milliliter (pg/mL). The performance characteristics of this kit from the manufacturer’s sheet are given as: The measuring ranges, sensitivity, intra-assy precision and inter-assy precision of the endocan kit were 15.63 pg/mL to 1000 pg/mL.

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9.375 pg/mL, < 8 and < 10 %, respectively. Values beneath the detection limit of 15.63 pg/mL were regarded as ‘0’.

Statistical analysis
The power analysis was performed using G Power 3.1 for Windows (Heinrich-Heine-Universität Düsseldorf, Düsseldorf, Germany) [17]. Sample size calculation was based on the mean endocan levels. Mean endocan levels were reported as 621.82 ± 345.41 pg/mL and 391.40 ± 234.97 pg/mL in our pilot study. According to our pilot study results for circulating endocan levels, the minimum required size of the study population was calculated to be 70 subjects (α = 0.05 and the study power = 0.90).

Quantitative data were summarized using mean and standard deviation. Independent samples t test or one way analysis of variance (ANOVA) test were used to compare normal data between groups. For post-hoc comparisons between the pair-wise groups, the Tukey HSD test was used. Mann Whitney U test was used to compare non-normal data between groups. Pearson correlation coefficients were used to test inter-variable relationships. A p value < 0.05 was considered statistically significant. Analyses were performed using SPSS 19 (IBM SPSS Statistics 19, SPSS Inc., Somers, NY, USA).

RESULTS
Clinical findings
The mean age of all the participants was 22.88 ± 4.40 years, and BMI was 23.03 ± 4.06 kg/m². The frequency of PCO finding on US, oligomenorrhea, and hyperandrogenism was 73 (83.9%), 69 (78.4%), and 68 (77.3%), respectively. Age, BMI, and hirsutism score of PCOS patients were significantly higher than control group (p < 0.05) (Tab. 1). According to BMI, of all the study population, 32 (18.3%) and 15 (8.5%) were overweight (25–29.9 kg/m²) and obese (≥ 30 kg/m²), respectively. On the other hand, among PCOS patients, 20 (22.7%) and 14 (15.9%) were overweight and obese, respectively.

Biochemical findings
Biochemical and hormonal profiles of the control group were within normal ranges. Among biochemical parameters, triglyceride, total cholesterol, LDL, fasting blood glucose and CRP were significantly higher in PCOS group compared to control group (p < 0.05) (Tab. 1). For hormonal parameters, LH, E2, DHEA-S and total testosterone were significantly higher in PCOS group compared to control group (p < 0.05), as FSH and TSH were comparable between groups (Tab. 1).

Table 1. Distributions of variables in the study groups

<table>
<thead>
<tr>
<th></th>
<th>Control group (n = 87)</th>
<th>PCOS patients (n = 88)</th>
<th>p</th>
</tr>
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<tbody>
<tr>
<td>Demographic and clinical findings</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Age [years]</td>
<td>23.71 ± 4.42</td>
<td>22.06 ± 4.24</td>
<td>0.012a</td>
</tr>
<tr>
<td>BMI [kg/m²]</td>
<td>22.15 ± 3.03</td>
<td>23.9 ± 4.74</td>
<td>0.004a</td>
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<tr>
<td>Hirsutism score</td>
<td>0.39 ± 0.6</td>
<td>9.95 ± 4.23</td>
<td>&lt; 0.001b</td>
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<tr>
<td>Hormonal parameters</td>
<td></td>
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<tr>
<td>FSH</td>
<td>5.59 ± 1.68</td>
<td>5.54 ± 1.9</td>
<td>0.830a</td>
</tr>
<tr>
<td>LH</td>
<td>8.2 ± 3.3</td>
<td>10.95 ± 7.47</td>
<td>0.002a</td>
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<tr>
<td>DHEA-S</td>
<td>92.62 ± 36.37</td>
<td>240.36 ± 122.01</td>
<td>&lt; 0.001b</td>
</tr>
<tr>
<td>Total testosterone</td>
<td>0.06 ± 0.1</td>
<td>0.56 ± 0.28</td>
<td>&lt; 0.001b</td>
</tr>
<tr>
<td>E2</td>
<td>59.99 ± 15.79</td>
<td>67.78 ± 69.51</td>
<td>0.037b</td>
</tr>
<tr>
<td>TSH</td>
<td>1.68 ± 0.58</td>
<td>1.82 ± 0.98</td>
<td>0.252a</td>
</tr>
<tr>
<td>Biochemical parameters</td>
<td></td>
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</tr>
<tr>
<td>Fasting blood glucose</td>
<td>79.45 ± 6.6</td>
<td>88.66 ± 9.57</td>
<td>&lt; 0.001a</td>
</tr>
<tr>
<td>Triglyceride</td>
<td>72.56 ± 12.75</td>
<td>110.28 ± 50.79</td>
<td>&lt; 0.001a</td>
</tr>
<tr>
<td>Total cholesterol</td>
<td>106.3 ± 16.35</td>
<td>167.51 ± 32.22</td>
<td>0.001a</td>
</tr>
<tr>
<td>HDL</td>
<td>53.44 ± 3.66</td>
<td>51.64 ± 9.71</td>
<td>0.107a</td>
</tr>
<tr>
<td>LDL</td>
<td>90.13 ± 9.43</td>
<td>110.11 ± 25.14</td>
<td>&lt; 0.001a</td>
</tr>
<tr>
<td>CRP</td>
<td>3.84 ± 1.6</td>
<td>5.33 ± 4.72</td>
<td>&lt; 0.001b</td>
</tr>
</tbody>
</table>

Data are shown as mean ± standard deviation; * Mann Whitney U test; PCOS — polycystic ovary syndrome; BMI — body mass index; FSH — follicle stimulating hormone; LH — luteinizing hormone; DHEA-S — luteinizing hormone; E2 — estradiol; TSH — thyroid stimulating hormone; HDL — high-density lipoprotein; LDL — low-density lipoprotein; CRP — C-reactive protein

Table 2. Bivariate correlations of variables with endocan in all participants and PCOS and control groups

<table>
<thead>
<tr>
<th></th>
<th>Endocan (pg/mL)</th>
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<tbody>
<tr>
<td></td>
<td>Total (n = 175)</td>
<td>PCOS group (n = 88)</td>
<td>Control group (n = 87)</td>
</tr>
<tr>
<td>BMI</td>
<td>r 0.210</td>
<td>-0.428</td>
<td>-0.090</td>
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<tr>
<td></td>
<td>p 0.005</td>
<td>&lt; 0.001</td>
<td>0.406</td>
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<tr>
<td>Hirsutism score</td>
<td>r 0.231</td>
<td>-0.199</td>
<td>0.512</td>
</tr>
<tr>
<td></td>
<td>p 0.002</td>
<td>0.063</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>LH</td>
<td>r 0.229</td>
<td>0.186</td>
<td>0.128</td>
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<tr>
<td></td>
<td>p 0.002</td>
<td>0.083</td>
<td>0.237</td>
</tr>
<tr>
<td>DHEA-S</td>
<td>r 0.273</td>
<td>0.038</td>
<td>0.279</td>
</tr>
<tr>
<td></td>
<td>p &lt; 0.001</td>
<td>0.723</td>
<td>0.009</td>
</tr>
<tr>
<td>Total testosterone</td>
<td>r 0.346</td>
<td>0.057</td>
<td>0.453</td>
</tr>
<tr>
<td></td>
<td>p &lt; 0.001</td>
<td>0.597</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Fasting blood glucose</td>
<td>r 0.328</td>
<td>-0.007</td>
<td>0.559</td>
</tr>
<tr>
<td></td>
<td>p &lt; 0.001</td>
<td>0.947</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Total cholesterol</td>
<td>r 0.299</td>
<td>-0.028</td>
<td>0.284</td>
</tr>
<tr>
<td></td>
<td>p &lt; 0.001</td>
<td>0.799</td>
<td>0.008</td>
</tr>
<tr>
<td>HDL</td>
<td>r 0.158</td>
<td>0.392</td>
<td>-0.309</td>
</tr>
<tr>
<td></td>
<td>p 0.037</td>
<td>&lt; 0.001</td>
<td>0.004</td>
</tr>
<tr>
<td>CRP</td>
<td>r -0.093</td>
<td>-0.424</td>
<td>0.648</td>
</tr>
<tr>
<td></td>
<td>p 0.222</td>
<td>&lt; 0.001</td>
<td>&lt; 0.001</td>
</tr>
</tbody>
</table>

r denotes Pearson correlation coefficient; PCOS — polycystic ovary syndrome; BMI — body mass index; LH: luteinizing hormone; DHEA-S — luteinizing hormone; HDL — high-density lipoprotein; CRP — C-reactive protein
Serum endocan levels

Serum endocan level was significantly higher in PCOS group than control group (540.9 ± 280.3 pg/mL vs. 355.5 ± 233.5 pg/mL, respectively; p < 0.001) (Fig. 1). The presence of PCO finding on US and oligomenorrhea in PCOS group did not produce significant effect on blood endocan levels (p > 0.05) (Fig. 1). However, overweight and obese PCOS patients had significantly lower blood endocan levels than normal weight PCOS patients (p = 0.002) (Fig. 1).

As BMI and LH were not correlated with endocan levels in control group; hirsutism score, DHEA-S, total testosterone, fasting blood glucose, total cholesterol, and CRP were positively and HDL was negatively correlated with endocan levels (Tab. 2). In PCOS group, there was a weak-to-moderate negative correlation between endocan levels and BMI and CRP, and positive correlation between endocan levels and HDL (Tab. 2).

**DISCUSSION**

In this case-control study, we primarily showed that serum endocan level is increased in patients with PCOS. Since BMI and CRP, the parameters related with inflammatory process, were negatively correlated with blood endocan level, hormonal rather than inflammatory/endothelial changes of PCOS are implicated for the increase in circulating endocan level. The presence of PCO finding on US or oligomenorrhea did not affect the endocan level, suggesting that there is no relation between imaging findings and menstrual symptoms of PCOS and endocan level.

Endocan is a circulating endothelium-derived proteoglycan. An increase in tissue or blood endocan level shows endothelial activation and neovascularization, which are indicators of inflammation and tumor progression [18]. Therefore, blood endocan level has been used as a biomarker for various cancers including ovarian or endometrial carcinoma, pre-eclampsia, and inflammation-related diseases such as cirrhosis, sepsis, hypertension, and Behcet’s disease [18–24]. As well as presence and severity of inflammation-related diseases, these markers indicate cardiovascular and metabolic risk of patients [25]. Furthermore, some studies proposed endocan as a potential target for treatment of these diseases [26, 27].

It has been known that PCOS is a low-level chronic inflammation with increased serum level of inflammatory markers such as interleukin-1, interleukin-6, CRP, and tumor necrosis factor alpha [28, 29]. Accordingly, in the present study biochemical and hormonal profiles of patients with PCOS showed significantly higher level of cardiovascular and inflammatory parameters regardless of PCO finding on US, oligomenorrhea, and BMI. The low-grade inflammation in PCOS is commonly associated with endothelial dysfunction regardless of BMI [30, 31]. Since increased level of circulating endocan is associated with endothelial dysfunction in
inflammatory conditions, which was suggested as an early sign of atherogenesis, endocan may have a potential role as a prognostic biomarker in PCOS and indicator of cardiovascular risk [32, 33].

In a recent study, Bicer et al. reported an increased level of circulating endocan levels in women with PCOS compared with controls (5.99 ± 2.37 ng/mL vs. 3.66 ± 1.79 ng/mL, p < 0.001) [14]. Similarly, in our study, which is the second study in the literature evaluating endocan level in PCOS, serum endocan level was significantly higher in PCOS group than the control group (540.9 ± 280.3 pg/mL vs. 355.5 ± 233.5 pg/mL, respectively; p < 0.001).

Lean women populations with PCOS are a specific group and have different phenotypic, metabolic, and hematologic characteristics than obese women with PCOS and there is still an ongoing debate regarding this category of patients. It has been shown that obesity, and not PCOS status per se, was a major determinant of the circulating inflammatory markers TNF-α, soluble type 2 TNF receptor, IL-6, and hs-CRP [34]. While the source of excess circulating TNF-α in PCOS is likely to be adipose tissue in the obese but remains unknown in lean women with the disorder [35]. Therefore studies suggest that obesity is not independently associated with these markers of inflammation.

While they did not stratify PCOS subjects based on BMI, Bicer et al. also found that endocan is an independent predictor for carotid intima media thickness, which indicates risk of cardiovascular events, thus suggested that blood endocan level can be used to predict increased cardiovascular risk in PCOS patients [14]. However, in our study we interestingly found that overweight and obese PCOS patients had significantly lower serum endocan levels than normal weight PCOS patients, and in PCOS group, there was a weak-to-moderate negative correlation between endocan levels and BMI and CRP, and positive correlation between endocan levels and HDL, indicating that endocan is not a direct marker of inflammation or cardiovascular risk in PCOS. We suggest that the increased endocan levels in lean PCOS patients compared to obese women with PCOS in our study may be explained by [1] the intracellular signalling pathways that govern the relationship between inflammatory response and circulating endocan levels may be more impaired in lean women with PCOS than obese women with PCOS or [2] endocan production may be regarded as part of a protective mechanism in the crosstalk of multiple signalling mediators, fighting with the state of increased inflammation. We think our study contributes to our understanding on the presence of PCOS in lean women while future studies in this field should define the Rotterdam phenotype of subjects, stratify subjects based on BMI, use clinically relevant cut offs rather than comparison of means and ultimately provide longitudinal data on changes in biomarkers and their correlation with metabolic and clinical manifestations of PCOS.

Furthermore, in our study the presence of PCO finding on ultrasonography and oligomenorrhea in PCOS group did not produce significant effect on serum endocan levels. Therefore, we suggest that there is no relation between symptoms of PCOS and endocan level. On the other hand, in control group of women with normal menstrual cycles; hirsutism score, DHEA-S, total testosterone, fasting blood glucose, total cholesterol, and CRP were positively and HDL was negatively correlated with endocan levels.

The main limitation of our study was its cross-sectional design, which limits the interpretation of cause-effect relation of increased level of circulating endocan level and PCOS. Nevertheless, this is among the few studies on the blood endocan level in patients with PCOS. Further studies are needed to clarify the exact mechanism underlying the increase in blood endocan level in PCOS.

CONCLUSIONS

Serum endocan level is increased in patients with PCOS, which seems to result from hormonal or other factors which need to be evaluated rather than inflammatory/endothelial changes of PCOS. Further studies are needed to conclude on mechanism of increased endocan level in PCOS, and to evaluate the clinical value of endocan as a biomarker for inflammation and cardiovascular risk, and potential therapeutical target in PCOS.

Acknowledgements

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Conflict of interest

Authors declare no conflicts of interest.

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