

GC-MS as a tool for reliable non-invasive prenatal diagnosis of Smith-Lemli-Opitz syndrome but essential also for other cholesterolopathies verification

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ABSTRACT

Rare multiple congenital malformations/developmental disorders are challenging in clinical diagnosis. The introduction of next-generation sequencing (NGS) has revolutionized this diagnostic by offering multigene panels or whole-exome sequencing. However, if there is no possibility to perform NGS or if we are facing prenatal ultrasound results, clinical diagnostics is even more difficult. For a selected group of congenital metabolic disorders, resulting from defects in cholesterol biosynthesis (called cholesterolopathies), application of gas chromatography-mass spectrometry (GS-MS) may provide or orientate diagnostics. The most common of these is Smith-Lemli-Opitz syndrome (SLOS), but in this publication, we also want to introduce other cholesterolopathies and draw attention to the possibility of non-invasive prenatal diagnosis of SLOS.

Key words: prenatal diagnosis; GC-MS; prenatal ultrasound; Smith-Lemli-Opitz; cholesterol biosynthesis

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INTRODUCTION

Cholesterol (cholest-5-en-3beta-ol) is a chemical compound belonging to specific lipids called steroids, which common feature is the presence of a carbon skeleton, composed of four coupled rings (steran) (Fig. 1).

In the human body, it occurs in plasma, blood and tissues in both free and fatty acid esterified forms. Endogenous cholesterol is mainly synthesized in the liver, intestines and

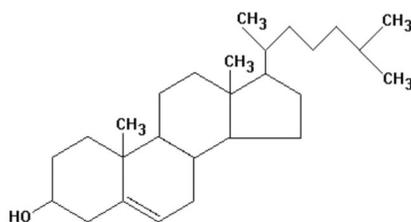


Figure 1. Cholesterol (C₂₇H₄₆O) with marked side groups (hydroxyl and methyl)

skin, as well as in the central nervous system [1]. The human brain contains 23% of the cholesterol circulating in the body. Most of it is synthesized in oligodendrocytes and accumulated in myelinated axon sheaths, but it is also found in the cells of neurons and astrocytes. The blood-brain barrier prevents the penetration of cholesterol from the bloodstream, which excludes the regulation of cholesterol levels in the brain through supplementation [1]. The optimal amount of cholesterol in the human body is determined by the correct course of the process of its biosynthesis, which may be disturbed in discussed below syndromes.

Although the developing fetus attempts to synthesize cholesterol, in the first weeks of pregnancy, it uses mainly maternal cholesterol. Its transport through the **membranes of the secondary yolk sac (during the first eight weeks of gestation), and later the placenta (when trophoblast takes over its nutritional role) to the fetal circulation,** determines the proper development of the embryo and

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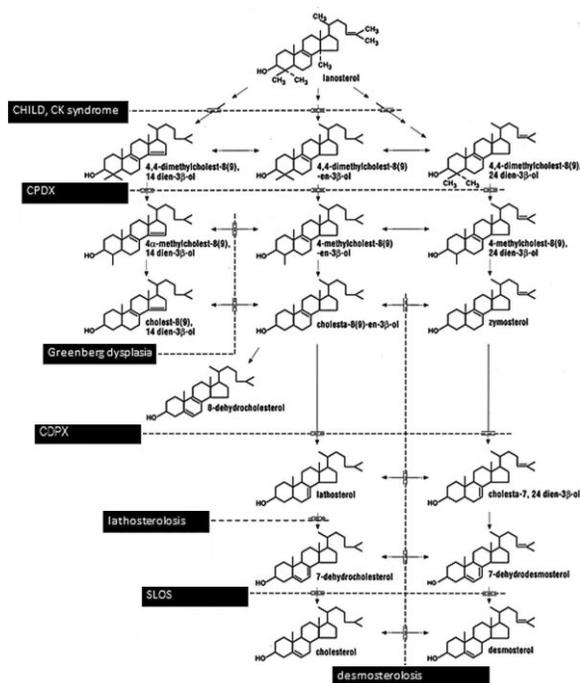


Figure 2. Part of the cholesterol biosynthesis pathway (modified from Nowaczyk MJM, Cunniff C. 2012. Smith–Lemli–Opitz syndrome and other disorders of cholesterol biosynthesis: An introduction. *Am J Med Genet Part C Semin Med Genet* 160C: 239–241).

the development of most organs. Cholesterol molecules collected by the embryo participate in signalling pathways crucial for embryonic development — they regulate the function of sonic hedgehog (SHH) proteins [2]. These proteins determine the survival and migration of nerve cells and important nuclear receptors, such as the transcription factor for alpha-fetoprotein, which, by binding to DNA GATA sequences, enables the transcription of numerous genes involved in the development of crucial organs [3].

Congenital defects in enzymes of the cholesterol biosynthesis pathway have recently emerged as significant causes of congenital anomalies. Patients with these metabolic diseases present with different malformations that involve many organs and systems [4–6]. To date, nine disorders due to enzymatic defects in post-squalene cholesterol biosynthesis have been identified (Fig. 2). These are:

1. Smith–Lemli–Opitz syndrome (SLOS, OMIM: 270400),
2. X-linked dominant chondrodysplasia punctata type 2 (CDPX2, OMIM: 302960) and MEND (male EBP disorder with neurological defects; OMIM: 300960),
3. Congenital hemidysplasia with ichthyosiform erythroderma and limb defects syndrome (CHILD syndrome, OMIM: 308050),
4. Sterol-C4-methyloxidase-like deficiency (SC4MOL deficiency, OMIM: 607545),
5. CK syndrome [named for the initials of the original proband] (OMIM: 300831),

6. Greenberg/HEM dysplasia (hydrops-ectopic calcification-moth-eaten skeletal dysplasia, OMIM: 215140), (this phenotype is likely due to a laminopathy, but is usually discussed with inborn errors of cholesterol synthesis, see Table 1 for causative details),
7. Antley–Bixler syndrome with ambiguous genitalia (cytochrome P450 oxidoreductase deficiency, POR deficiency, OMIM: 201750),
8. Desmosterolosis (OMIM: 602398),
9. Lathosterolosis (OMIM: 607330).

SMITH-LEMLI-OPITZ SYNDROME

The best-known among the listed diseases is Smith–Lemli–Opitz syndrome caused by a low activity of 3β-hydroxysteroid-D7-reductase (7-dehydrocholesterol reductase, DHCR7). Its role is to convert 7-dehydrocholesterol (7DHC) into cholesterol in the Kandutsch-Russell pathway and 7-dehydrodesmosterol into desmosterol in the Bloch pathway (Fig. 3).

Desmosterol is mainly present in the brain [7]. SLOS, first described in 1964, is also the first multiple malformation syndrome attributed to an inborn error of sterol synthesis [8, 9]. The consequence of a malfunctioning enzyme is primarily the accumulation of 7-DHC in blood and tissue and probably 7-dehydrodesmosterol in the brain [10]. Besides, acting in cells Δ7,Δ8-isomerase converts 7-dehydrocholesterol into 8-dehydrocholesterol, and, similarly, 7-dehydrodesmosterol in 8-dehydrodesmosterol [11]. Studies have shown that patients with milder symptoms have normal cholesterol levels in the membranes of nerve cells, which is probably due to its local synthesis. At the same time, it seems that disease symptoms may also be caused by the accumulation of 7,8-dehydrodesmosterol or its oxidised metabolites [10].

Although its clinical presentation may vary, SLOS is usually characterized by prenatal and postnatal growth retardation, microcephaly, moderate to severe intellectual disability, and multiple major and minor malformations, including characteristic facial features, cleft palate, cardiac defects, postaxial polydactyly, 2–3 toe syndactyly, hypospadias, and undervirilization of the genitalia in males [9, 12, personal observation]. There are also data suggesting higher intrauterine mortality of SLOS-affected fetuses [13]. The reported manifestations of Smith–Lemli–Opitz syndrome during the prenatal period (presented in Tab. 1) include, among others: intrauterine growth retardation [14, 15], low maternal unconjugated estriol (MSuE3) [16], or a combination of very rare congenital anomalies, such as ulnar hypoplasia, vertebral segmentation anomalies, congenital pulmonary adenomatoid malformation, fused lungs, laparoschisis, holomyelia, and hypothalamic hamartoma [17].

Table 1. Clinical presentation and diagnostic procedures in disorders with impaired cholesterol biosynthesis

Disorder	Possible prenatal; typical postnatal findings	Causative gene/enzyme	Biochemical markers (pre- and postnatal)
Smith–Lemli–Opitz syndrome	increased nuchal translucency (NT > 3 mm); delayed growth, microcephaly, CNS malformations (i.a., holoprosencephaly), cleft palate, cardiac defects (i.a., atrioventricular canal defect, AVCD), renal and genital anomalies (especially sex reversal in 46,XY fetuses), polydactyly [27]	<i>DHCR7</i> 7-dehydrocholesterol reductase	elevated 7-dehydrocholesterol and 8-dehydrocholesterol; very low to normal cholesterol; prenatally also: 7-dehydropregnantriol and 8-dehydroestriol in maternal urine
X-linked dominant chondrodysplasia punctata type 2 (CDPX2 or Conradi–Hunermann–Happle syndrome)	affects females; asymmetric shortening of the long bones, stippling of the epiphyses, clubfoot, joint contractures, heart and renal defects [23]; postnatal skin manifestation (ichthyosis following the Blaschko's lines, scarring alopecia), cataract	<i>EBP</i> /Emopamil binding protein (3b-hydroxysteroid-D8,D7-sterol isomerase)	elevated cholesta-8(9)-en-3β-ol (and 8-dehydrocholesterol)
Emopamil binding protein (EBP) deficiency in males (X-linked recessive syndrome of multiple congenital abnormalities and intellectual disability, allelic to X-linked dominant chondrodysplasia punctate); MEND syndrome (male EBP disorder with neurological defects)	12 cases reported; CNS malformations (corpus callosum agenesis, Dandy–Walker malformation, hydrocephalus), heart defect, postaxial polydactyly; postnatally facial dysmorphism, epiphyseal stippling [19]	Hypomorphic mutations of the <i>EBP</i> gene <i>EBP</i> /Emopamil binding protein (3b-hydroxysteroid-D8,D7-sterol isomerase)	elevated cholesta-8(9)-en-3β-ol (and 8-dehydrocholesterol)
Congenital hemidysplasia with ichthyosiform erythroderma and limb defects syndrome (CHILD syndrome)	unilateral limb reduction defects; poly- oligo-, ectrodactyly, unilateral renal malformations, congenital heart defects (10–20% of cases) and punctate epiphyseal calcifications [28]	<i>MSDHL</i> /3β-hydroxysteroid dehydrogenase	elevated 4α-monomethyl and 4,4′α-dimethyl sterols or elevated 4α-carboxymethylcholest-8(9)-en-3β-ol (not in plasma, only tissue)
Sterol-C4-methyloxidase-like (SC4MOL) deficiency	5 cases reported; microcephaly in 4 [29]	<i>MSMO1</i> (SC4MOL)/Methylsterol monooxygenase 1 (sterol-C4-methyloxidase)	elevated 4α-monomethyl and 4,4′α-dimethyl sterols; low to normal cholesterol (postnatally)
CK syndrome	24 affected males from three unrelated families; microcephaly in all; postnatally: facial dysmorphism and severe developmental delay [28]	Hypomorphic mutations of the <i>MSDHL</i> /3β-hydroxysteroid dehydrogenase	elevated 4α-monomethyl and 4,4′α-dimethyl sterols (to a lower degree than in CHILD s.) (not in plasma, only tissue)
Greenberg dysplasia/ Hydrops-ectopic calcification-moth-eaten dysplasia (HEM dysplasia)	lethal prenatally; hydrops fetalis (non-immune), severe limb shortening with disorganized cartilaginous/bony architecture, small thorax	<i>LBR</i> / Lamin B receptor (has sterol-D14-reductase activity)	elevated cholesta-8,14-dien-3β-ol and cholesta-8,14,24-trien-3β-ol
Congenital adrenal hyperplasia due to P450 oxidoreductase deficiency	numerous cases reported (5 in Poland) <i>skeletal, craniofacial, and urogenital anomalies</i> : craniosynostosis, exophthalmos, radio-humeral synostosis, bowing of the femur/ulna, multiple contractures (resemblance to Antley-Bixler syndrome due to <i>FGFR2</i> mutations), ambiguous genitalia in 46,XX and 46,XY fetuses	<i>POR</i> / P450 oxidoreductase (genetics are summarised by Baronio et al. [30])	elevated lanosterol and dihydrolanosterol + presence of markers of adrenal/gonadal CYP21A2, CYP17A1, CYP19A1 deficiency (elevated serum 17α-hydroxyprogesterone, 21-deoxy-cortisol, corticosterone, progesterone, diminished androgens and estrogens)
Desmosterolosis	10 cases reported; agenesis of the corpus callosum with white matter atrophy with/without ventriculomegaly, retromicrognathia with/without cleft palate, distal arthrogyposis, and delay in growth and development [31]	<i>DHCR24</i> / 3b-hydroxysteroid-D24-reductase	elevated desmosterol (and 7-dehydrocholesterol)
Lathosterolosis	5 cases reported with spectrum of congenital anomalies overlapping with those in Smith–Lemli–Opitz s: microcephaly, postaxial polydactyly, horseshoe kidney (1 case), meningocele (2 case) (summarized by Anderson et al. [24]; 1 another with progressive liver involvement [32])	<i>SCSCL</i> / 3b-hydroxysteroid-D5-desaturase (sterol-C5 desaturase)	elevated lathosterol (and raised concentration of 8(9)-cholestenol)

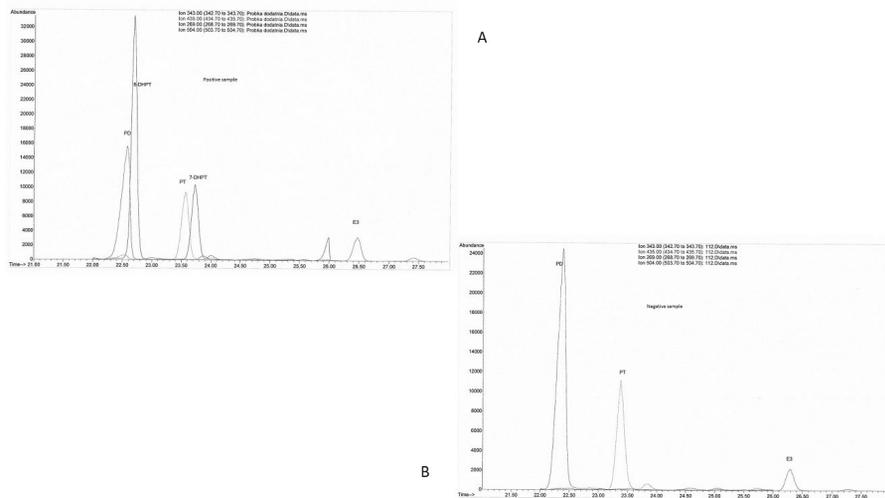


Figure 4. GC/MS results of maternal urine sample in pregnancy; A. affected with Smith-Lemli-Opitz syndrome (note elevated 7- and 8-DHPT); B. healthy child

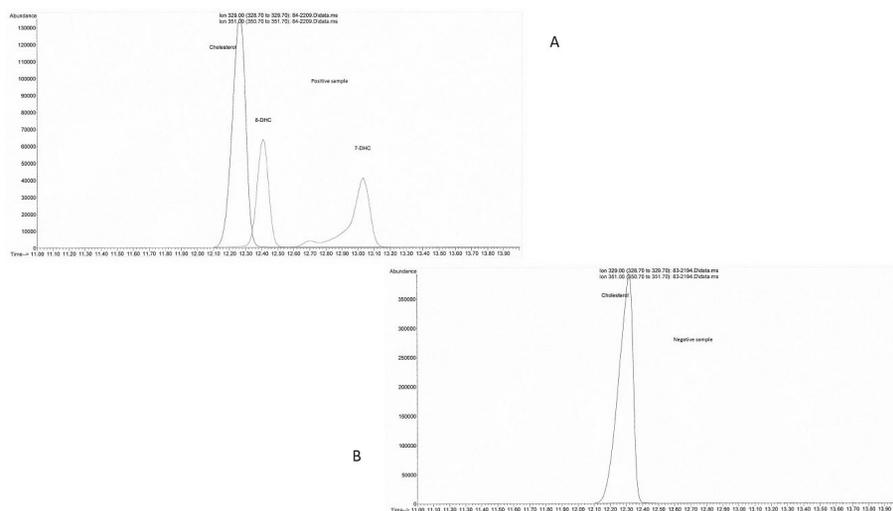


Figure 5. GC/MS of plasma sample from; A. patient affected with Smith-Lemli-Opitz syndrome (note elevated 7- and 8-DHC); B. healthy patients

in the *DHCR7* gene in the fetus. Morning urine samples of pregnant women taken between 13 and 24 weeks of gestation are preferred for analysis.

Another technique to detect *DHCR7* deficiency in the fetus is the analysis of amniotic fluid metabolites, where 7- and 8-dehydrocholesterol occurring in trace amounts in children without SLO syndrome are directly quantified. The same chemical compounds are determined in the child's plasma after birth, and their concentration is 50–1000 times higher than the reference norm, which at low cholesterol is a high diagnostic value in SLO syndrome (Fig. 5).

For verification of biochemical results or in cases when prenatal samples are not useful for biochemical tests (borderline normal results or lack of validation for specific markers), molecular analyses may be offered for prenatal

detection of the discussed syndromes. They are based on DNA isolated from chorionic villus, amniotic fluid or rarely umbilical blood. Results of biochemical analysis of maternal urine, amniotic fluid and direct CVS analysis are available within 2 or 3 days. Molecular studies of cultured amniocytes or villus cells require 2 to 3 weeks to complete.

CONCLUSIONS

We hope that this article will provide readers with knowledge about a group of disorders characterized by defects in cholesterol biosynthesis, thus increasing awareness in the medical community about their diagnosis, especially in the prenatal period with the application of non-invasive prenatal biochemical test (GC-MS analysis). The validated methodology allows for **cheap (comparing to molecular**

Table 2. Details concerning samples and shipment for testing toward Smith–Lemli–Opitz syndrome to the CMHI

SLOS markers/material	Sample requirements		Necessary forms
7,8-dehydro-cholesterol and cholesterol in amniotic fluid (AF) ; also latosterol, desmosterol (in 13-18 hbd samples)	1 ml AF	protect the tube from sunlight, wrap with dark film	Test Requisition Form available at: www.czdz.pl (refer to Pracownia Hormonów Steroidowych i Zaburzeń Metabolizmu)
estriol, 8-dehydroestriol, 7-dehydropregnantriol, pregnantriol, pregnandioli in maternal urine + creatinine	single morning urine sample of pregnant women ≥ 13 hbd	as above	as above

testing, including a panel of genes or exome sequencing), fast and reliable diagnosis of some of them. In Poland, it is offered at the Children's Memorial Health Institute, Department of Biochemistry, Radioimmunology and Experimental Medicine. The details of samples and shipment for Smith-Lemli-Opitz syndrome testing are given in the table below.

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Conflict of interest

None.

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