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Assessment of selected parameters of placental microstructure in patients with intrahepatic cholestasis of pregnancy

Milena Gruszczynska-Losy¹, Ewa Wender-Ozegowska², Przemyslaw Wirstlein², Malgorzata Szczepanska²

¹Gynecologic and Obstetrical University Hospital in Poznan, Poland
²Division of Reproduction, Department of Obstetrics, Gynaecology and Gynaecologic Oncology,
Poznan University of Medical Sciences, Poznan, Poland

ABSTRACT

Objectives: Intrahepatic cholestasis of pregnancy (ICP) is the most common liver disorder during pregnancy. Cholestasis is associated with increased risk of fetal complications: prematurity, perinatal hypoxia and meconium stained amniotic fluid, and sudden intrauterine fetal death. The exact mechanisms associated with cholestasis fetal sequelae are not fully understood. The aim of the study was the histopathological evaluation of placentas from patients with cholestasis and healthy pregnant women to establish whether cholestasis is accompanied by changes in placental microstructure.

Material and methods: The effect of cholestasis on placental microstructure was investigated using placental tissue from patients with cholestasis treated with ursodeoxycholic acid (UDCA) and from uncomplicated pregnancies. Five placental histopathological features were analyzed: number of syncytial knots, number of capillaries per villous, structure of stroma, presence of Hofbauer cells, and villitis of unknown etiology.

Results: There were no statistically significant differences in any of the studied parameters between cholestasis-affected and healthy control groups.

Conclusions: There are no diffrences in placental microstructure in cholestasis patients treated with UDCA and in patients with uncomplicated pregnancy.

Key words: intrahepatic cholestasis of pregnancy; ursodeoxycholic acid; placenta; histology

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INTRODUCTION

Intrahepatic cholestasis of pregnancy is the most common liver disorder during pregnancy, occurring in 1% of pregnancies. Very frequently ICP develops during the late second and third trimesters. Pregnant women with cholestasis present characteristic symptoms: pruritus, especially during the night, associated with abnormal liver function. These signs usually resolve after delivery. Among others, the most important biochemical feature of ICP is elevated serum bile acid levels (> 10 mmol/L) [1]. Cholestasis is associated with increased risk of fetal complications: prematurity, perinatal hypoxia and meconium stained amniotic fluid, and sudden intrauterine fetal death [2]. The increased risk of detrimental perinatal results correlates with high bile acid concentration (> 40 mmol/L) [3]. The exact mechanisms associated with

cholestasis fetal sequelaes are not fully understood. In the normal course of pregnancy, the total concentration of bile acids in fetal serum is only sligthly higher than in maternal serum. Because the fetal hepatobiliary and renal systems are not fully developed and cannot eliminate bile acids, the latter must be transferred across the placenta to be eliminated by the maternal liver. In patients with cholestasis, as a result of high levels of bile acids in the serum, the transplacental bile acid gradient is reversed, causing impairment placental transport [1, 4].

Bile acids induce vasoconstriction of the chorionic vessels in the placenta that causes impaired fetal-maternal transport across the placenta, damage of the placental structure and reduced transport of nutrients and oxygen to the fetus.

Corresponding author:

Przemyslaw Wirstlein

Division of Reproduction, Department of Obstetrics, Gynaecology and Gynaecologic Oncology, Poznan University of Medical Sciences, 33 Polna St, 60—535 Poznan, Poland e-mail: abys@wp.pl

Little is known about the effect of intrahepatic cholestasis of pregnancy on the placental structure. Most reports involving sudden intrauterine fetal death are not connected with prominent morphological changes in the placenta or the features of its chronic insufficiently [5, 6].

The aim of the study was to evaluate the histopathological changes of placentas from patients with cholestasis and healthy pregnant women to establish whether cholestasis affects placental microstructure.

MATERIAL AND METHODS

Twenty patients with intrahepatic cholestasis and 40 women as controls with physiological pregnancy who delivered in 2018 in the Gynecologic and Obstetrical University Hospital, Poznan, Poland were qualified to this prospective case-control study. Recognision of cholestasis was determined after excluding other liver diseases (viral hepatitis, acute fatty liver of pregnancy, pre-eclampsia, primary biliary cirrhosis and HELLP syndrome), on the basis of unexplained itching associated with increased bile acid (> 10 mmol/L) and liver dysfunctions. We also excluded patients with complications of pregnancy other than ICP. We included patients with physiological pregnancy in the control group. Patients in the control group did not have cholestasis in their previous pregnancy or pruritus in the previous or current pregnancy. Inclusion criteria for pregnant women with cholestasis and controls demanded that all patients have single, live-born newborn.

All participants gave written informed consent. The obtained data included concentration of transaminases and bile acid at the time of diagnosis and delivery, gestational week at diagnosis and delivery, the highest concentration of bile acid and transaminases, neonatal birth weight, umbilical pH value, Apgar score, placental weight, and placental/neonatal birth weight ratio (Tab. 1). All women with ICP were treated with ursodeoxycholic acid (UDCA). All the participants were nonsmokers.

The study was approved by the Ethical Committee of Poznan University of Medical Sciences.

Collection of samples and slide preparation

After delivery placentas were weighed and fixed in 4% buffered formalin for 10 days. After fixation, the samples were dehydrated using in sequence: 70–100% series of ethyl alcohol dilutions, xylene, and embedded in 58°C paraffin. Samples were cut at 5 mm using a microtome (Leica SM 2010R) and mounted on slides. The slides were incubated for 2 h at 58°C to remove excess paraffin and ensure laminar adhesion. Deparaffinization was performed in three changes of xylene for 10 min each. Next, the slides were rehydrated through a 100–70% series of ethyl alcohol dilutions and washed in distilled water. Finally, deparaffinized samples were stained with hematoxylin and eosin (HE) and examined with a light microscope with camera attachment (Axioskop 40 ZEISS) with 100x and 400x magnification.

Histology

Histological analysis of the preparations (n = 20 ICP patients, n = 40 control patients) was performed by pathologist unaware of gestational age and disease status.

Evaluation of syncytial knots

The definition of syncytial knots was described by Geenes et al. [7] as the presence of at least 10 aggregated syncytiotrophoblast nuclei that were not in direct contact with communicating villi surfaces. They illustrate areas of intensified apoptosis. In pregnancies with a pathological course complicated by intrauterine growth restriction and pre-eclampsia the amount of syncytial knots enhances. This phenomenon is a result of increased placental apoptosis. Exposure placental explant patches to influence of hyperoxia, hypoxia or reactive oxygen species in vitro causes creation of syncytial knots [7].

Table 1. Characteristics of maternal and delivery outcomes: values expressed as median (range) or mean (± SD)					
	ICP (n = 20)	Control (n = 40)	P value		
Maternal characteristics					
Age [years]	30 (± 5)	30 (± 5)	0.869		
Gravity	1 (1–6)	(1–4)	0.402		
Parity	0–5	0–3	0.415		
Fetal characteristics					
Gestation age at delivery [weeks]	37 (± 2)	38 (± 2)	0.698		
Birth weight [g]	2892 (± 683)	3623 (± 2763)	0.043		
Apgar score 5 minutes	10 (5–10)	10 (8–10)	0.058		
Umbilical artery pH	7.3 (7.2–7.5)	7.3 (7.1–7.5)	0.516		
Placental weight [g]	529 (± 42)	544 (± 68)	0.164		
Placenta/naonatal birth weight ratios	0.16 (0.02-0.038)	0.18 (0.13-0.62)	0.001		

In each placental sample, in three fields of view, the number of syncytial knots was counted manually. A count of syncytial knot density per mm² of villous tissue was allowed by the determination the surface area of villous tissue in the frames.

Evaluation of Hofbauer cells (HBCs)

Hofbauer cells are fetal origin [8]. They are placental villous macrophages, which emerge in placental tissue from 18 days after conception and persist to the end of pregnancy. By the fourth to fifth month of pregnancy, their identification becomes difficult as villous stroma becomes compressed [9]. Functions of tissue macrophages are phagocytosis of cellular debris and antigen presentation in response to infectious agents and inflammation [10]. Due to villitis of unknown etiology (VUE) and varial infection HBCs proliferation or hyperplasia is observed [11, 12]. In placentas from complicated pregnancies (gestational diabetes mellitus, intrauterine growth restriction, pre-eclampsia), the number of HBCs seems to increase while in placentas from uncomplicated pregnancies, HBCs either disappear or become less in number after the fourth month of pregnancy [13].

HBCs were identified as round or ovoid cells with eccentric nuclei and granular cytoplasm. The HBCs were counted in two high-power fields per slide at 400×10^{-5} magnification from three different fields in each section. The mean value per villous was calculated (Tab. 2).

Villitis

Villitis arises due to hematogenous infection of the placenta by TORCH infection. In contrast, majority of cases are immune mediated and are not an effect of infection [14]. There are two types of villitis: acute and chronic. In the first type of villitis, which is often a result of infection, polymorphonuclear leukocytes infiltrate the villi with or without associated necrosis. Whereas in the second type of villitis

the tissue is infiltrated by macrophages and lymphocytes usually with concomitant fibrosis and cellular proliferation of the villi [15, 16].

Statistical Analysis

For statistical analysis, SigmaStat version 3.5 software (Systat Software, Inc., Point Richmond, CA, USA) was used. The analysis of the results was based on the Student's t-test for variables with parametric distributions. For variables with non-parametric distributions, the Mann–Whitney rank sum test was used. The Fisher exact test was used for assessment of the villitis distribution. P < 0.05 was considered statistically significant.

RESULTS

Biochemical characteristics of the ICP patient's population is shown in the Table 3. There were no differences in neonatal,maternal and obstetrical outcomes. Nevertheless, there was a statistically significant difference in birth weights $(2892 \pm 683 \text{ g vs } 3623 \pm 2763 \text{ g}, p = 0.043)$ and placenta/neonatal birth weight ratios (0.16 vs 0.18, p = 0.001) in neonates born to women with and without cholestasis, respectively (Tab. 1).

The histological slides of 20 patients with ICP and 40 healthy controls were examined. Placentas from both groups were found to be appropriate for their gestational age. Five placental histopathological features were analyzed: number of syncytial knots, number of capillaries per villous, structure of stroma, presence of HBCs, and VUE. The placentas from both groups exhibit normal microstructure: compact stroma and appropriate number of capillaries per villous (< 10).

There were no statistically significant differences in any of the remaining three parameters between the cholestasis and healthy control groups (Tab. 3). Representative images are shown in Figure 1.

Table 2. Placental histology of the study population						
	ICP (n = 20)	Control (n = 40)	P value			
Hofbauer cells/villous [median (range)]	2 (0–5)	1 (0-3)	0.397			
Syncytial knots [median (range)]	21 (12–34)	15 (9–29)	0.143			
Structure of stroma	compact	compact	-			
Villitis of unknown etiology	1 (5%)	3 (7.5%)	0.999			
Number of capillaries per villous	< 10	< 10	-			

Table 3. Biochemical characteristic of ICP patient population, median (range)							
Sample	Week of delivery [week]	Bile acid (highest value) [mmol/L]	Alat [U/L]	Aspat [U/L]	Bile acid at delivery [mmol/L]		
N = 40	37 (± 2)	37.5 (11–171.3)	188.1 (13.5–1228.9)	101.2 (16.7–695.2)	22.4 (10.8–102)		

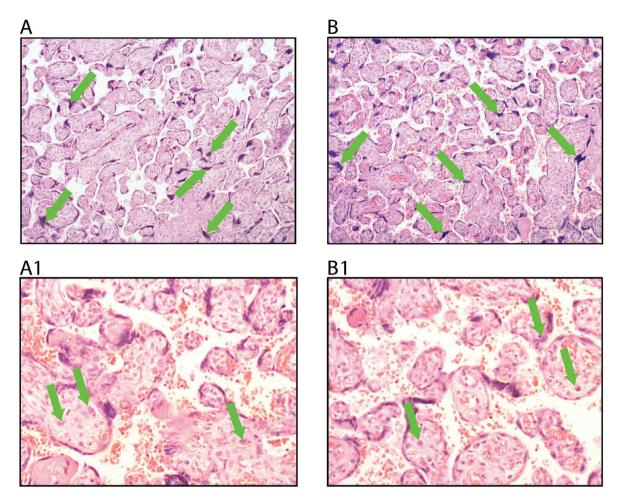


Figure 1. Representative images villi of trophoblast in a $20 \times (A, B)$ and $40 \times (A1, B1)$ lens magnification microscope stained by H + E obtained from women with normal pregnancy (A, A1) and with pregnancy complicated by cholestasis (B, B1) treated with ursodeoxycholic acid. The green arrows indicate the syncytial knots (A, B) and Hofbauer cells (A1, B1).

DISCUSSION

In this paper, we analyzed the histopathological images of placentas of patients with pregnancy complicated with cholestasis, and non-complicated, phisiological pregnancy. All pregnant women with cholestasis were treated with UDCA which is considered the first-line treatment for ICP, because it normalizes the transplacental bile acid gradient, significantly reducing fetal and maternal bile acid levels [17]. Five placental histopathological features were analyzed: number of syncytial knots, number of capillaries per villous, structure of stroma, presence of HBCs and VUE. In our study, we did not find any differences in the microstructure of the placenta of patients with cholestasis relative to patients with normal pregnancy.

Our results are in accordance with Patel et al. [18] who compared the placentas of 24 pregnant women with ICP and 30 healthy women. They found no differences in maternal, neonatal, and obstetrical outcomes. They found a significant

difference between the groups only in the gestation age at delivery. Patients with cholestasis delivered two weeks earlier than the controls. The authors analyzed 17 placental histopathological parameters and didn't find statistically significant differences in any of these features between patients with cholestasis and healthy pregnant women. Comparison of the placentas from women, treated and not treated with UDCA, demonstrated a statistically significant reduction in VUE (9% vs 53%, p = 0.03), which implies that UDCA has an anti-inflammatory impact on the placenta. However, the exact mechanism by which UDCA decreases placental inflammation is not known.

Guven et al. [19] investigated the alterations in the architecture of the umbilical cord and the placenta in selected pregnancy complications. They stated a significant enhancement in the amount of syncytial knots in placentas from women with pre-eclampsia, oligohydramnions, polyhydramnions, and repeated cesarean sections, but not in ICP

and control groups. Furthermore, they found a significant reduction in the diameter and volume of the arterial lumen of an umbilical artery in the ICP group compared to the control group. The arterial tunica intima and tunica media were thicker in ICP than in the control group.

The available data of the histopathological analysis of placenta in patients with cholestasis and its comparison with that of the group of healthy pregnant women indicates morphological differences. These changes include higher amount of syncytial knots and enhancement surface of terminal villi, and were observed in pregnant women with cholestasis untreated with UDCA.

In the article of Geenes et al. [20] slides prepared from 28 ICP patients and 12 healthy controls were subjected to histological examination. In the placentas achieved from pregnant women with cholestasis a couple of morphological abnormalities were found to be more frequent as compared to the placentas from physiological pregnancies. These contained chorionic villi that had dense fibrotic stroma and were small for the gestational age, focally-thickened amniotic basement membranes, increased in number of syncytial knots and stricture of intervillous space. The number of syncytial knots was significantly higher in placentas from untreated women with ICP compared to those from uncomplicated pregnancies (p = 0.02). However, there was no significant difference in the amount of syncytial knots in the placentas achieved from women with cholestasis treated with UDCA compared to healthy pregnant women.

Wikström Shemer et al. [21] in a prospective case-control study using the computerized stereology method, examined placentas from 10 untreated and 10 UDCA-treated patients with cholestasis, and eight healthy pregnant women for morphological differences. They analyzed five histopathological features of placentas (volume fraction of collagen, surface area of terminal villi and capillaries, chorangiosis, anumber of syncytial knots and volume of placenta).

The results of their research demonstrated that cholestasis influences the microarchitecture of placenta by enhancement the number of syncytial knots and terminal villous and capillary surface area. The UDCA treatment had protective effect on placental micro architecture. The analyses of placentas of UDCA-treated cholestasis, compared to untreated ICP, showed statistically significant differences in amount of syncytial knots, capillary surface area and terminal villous. The statistical differences were not detected in any analyzed parameters in placentas between UDCA-treated cholestasis patients and healthy pregnant women.

Increased capillary growth in terminal villi and syncytial knots were both described as a sign of hypoxia which is commonly evident in placentas coexisting with diseases with reduced blood perfusion (diabetes mellitus and pre-eclampsia).

Conclusions from the two abovementioned cited studies indicate that ICP is associated with a couple of anomalies of the placenta microstructure, including an enhancement in the amount of syncytial knots. However, in placentas of women treated with UDCA, the number of syncytial knots was comparable to those in placentas from uncomplicated pregnancies [20, 21].

The studies of Geenes et al. [20], Wikström Shemer et al. [21], and Patel et al. [18] were published in 2011, 2012, and 2014 respectively and, probably for that reason, some patients did not receive treatment with UDCA which is currently the first-line treatment for cholestasis.

The final answer to the question, whether treatment with UDCA in women with cholestasis improves obstetric outcomes, will be obtained after the completion of the triple-masked, placebo-controlled, randomised trial — PITCHES (Phase III trial in IntrahepaTic CHolestasis of pregnancy to Evaluate urSodeoxycholic acid in improving perinatal outcomes) which is currently randomized [22].

Our study has a number of limitations. Our work was carried out on a relatively small group of patients, but the real cholestasis, confirmed with biochemical data is rather rare in our population. To reduce statistical error, the histological examination of placentas should be carried out on a larger and an independent cohort. All patients with cholestasis included in this study, as well as all patients with cholestasis who delivered in 2018 in Gynecologic and Obstetrical University Hospital in Poznan, were treated with UDCA before delivery. UDCA treatment of pregnant women with cholestasis is standard in our hospital; therefore it was not possible to compare placental microstructure between patients with pregnancy complicated by cholestasis treated and not treated with UDCA.

CONCLUSIONS

Based on obtained results, we can conclude that the histopathological structure of the placenta of patients with cholestasis treated with UDCA does not differ from women with normal pregnancy.

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