This is a provisional PDF only. Copyedited and fully formatted version will be made available soon.



ORGAN POLSKIEGO TOWARZYSTWA GINEKOLOGICZNEGO THE OFFICIAL JOURNAL OF THE POLISH GYNECOLOGICAL SOCIETY

ISSN: 0017-0011

e-ISSN: 2543-6767

# Expression and diagnostic value of IncRNA HCG18 in gestational diabetes mellitus

Authors: Lin Chen, Jie Jiang, Yong Tian

**DOI:** 10.5603/gpl.95298

Article type: Research paper

**Submitted:** 2023-04-25

**Accepted:** 2023-05-29

Published online: 2023-10-09

This article has been peer reviewed and published immediately upon acceptance. It is an open access article, which means that it can be downloaded, printed, and distributed freely, provided the work is properly cited.

Articles in "Ginekologia Polska" are listed in PubMed.

#### ORIGINAL PAPER / GYNECOLOGY

# Expression and diagnostic value of lncRNA HCG18 in gestational diabetes mellitus

Lin Chen, Jie Jiang, Yong Tian

Central Hospital of Enshi Tujia and Miao Autonomous Prefecture, Enshi, China

# **Corresponding author:**

Yong Tian

Central Hospital of Enshi Tujia and Miao Autonomous Prefecture, Enshi, China

e-mail: tianyongdr@163.com

#### **ABSTRACT**

**Objectives:** Gestational diabetes mellitus (GDM) is the first type of diabetes induced by abnormal maternal glucose metabolism after pregnancy. Long non-coding RNA (lncRNA) has been found to be of great value in the study of its pathogenesis and treatment. This study aimed to explore the expression and diagnostic value of lncRNA HCG18 in GDM. Material and methods: The expression levels of HCG18 in serum of participating GDM patients and healthy controls were detected by polymerase chain reaction (RT-qPCR). The correlation between the expression of HCG18 and the blood glucose level was clarified by the detection of blood glucose levels in GDM patients. The receiver operating characteristic curve (ROC) was used to evaluate the clinical diagnostic value of HCG18 for GDM. Furthermore, multivariate logistic analysis was used to verify the diagnostic value of HCG18 in GDM. Results: This study concluded that the expression level of HCG18 was upregulated in the serum of GDM patients compared with the control group. ROC curve showed that the AUC was 0.916, the sensitivity was 80.5%, the specificity was 90.2%, and multivariate logistic regression analysis verified that HCG18 (OR = 6.984, 95% CI = 3.751–13.005, p < 0.001) was significantly associated with GDM, which suggesting that HCG18 has diagnostic significance for GDM. In addition, the expression of HCG18 was positively correlated with fasting blood glucose, 1h blood glucose and 2 h blood glucose of patients.

**Conclusions**: LncRNA HCG18 was elevated in patient serum and might serve as a diagnostic biomarker for GDM.

#### INTRODUCTION

Gestational diabetes mellitus (GDM) is the first diagnosed during pregnancy in patients with normal glucose metabolism or latent hypoglycemia before pregnancy [1]. Data show that GDM accounts for 80 percent of all diabetes-related pregnant women [2]. Gestational diabetes mellitus may cause maternal infection, increased amniotic fluid, fetal hyperglycemia, excessive insulin secretion, and complications such as hypertension and neonatal hypoglycemia [3–5]. The prevalence of GDM has increased significantly in recent years, due to changes in lifestyle and personal dietary habits. Currently, dietary therapy and drug therapy are the main treatment methods. Reasonable dietary control not only needs to ensure basic energy needs, but also avoid postprandial hyperglycemia or starvation ketosis to ensure the normal growth and development of the fetus [4]. If the blood sugar level cannot be effectively controlled in a short period of time, insulin therapy needs to be added according to the specific condition of the pregnant woman [6]. Therefore, it is necessary to explore new ways to detect and diagnose GDM in time.

LncRNAs have been found to act as biotherapeutic and prognostic targets in many diseases. For example, lncRNA SNHG5 can be used as a molecular therapeutic target for patients with various novel cancers [7], as well as MIR22HG [8]. LncRNA HLA complex group 18 (HCG18) belongs to the HLA complex group [9], HCG18 played an oncogenic role in osteosarcoma via miR-148b/ETV5, promoting osteosarcoma cell proliferation and metastasis in previous studies [10]. Exosome-mediated HCG18 promoted the polarization of M2 macrophages in gastric cancer by decreasing miR-875-3p in macrophages [11]. In addition, HCG18 has also been demonstrated to have regulatory ability in diabetic peripheral neuropathy, cardiovascular disease [12], that expect to clear cell renal cell carcinoma [13], hepatocellular carcinoma [14], and nasopharyngeal carcinoma [15]. However, the role of HCG18 in GDM remains unclear and requires in-depth exploration.

The current study aims to explore the expression and diagnostic value of HCG18 in GDM, to understand the potential of HCG18 as a diagnostic biomarker for GDM, and to provide theoretical reference for the subsequent treatment of GDM patients.

#### MATERIAL AND METHODS

## **Information of participants**

The clinical sample was 230 participants from Central Hospital of Enshi Tujia and

Miao Autonomous Prefecture, including GDM patients (n = 118) and controls (n = 112). Referring to the Chinese diagnostic criteria of GDM, fasting blood glucose at 5.1 mmol/L, 1 hour after taking glucose at 10.0 mmol/L, and 2 hours after taking glucose at 8.5 mmol/L were taken as the critical blood glucose. Gestational diabetes mellitus was diagnosed when the fasting blood glucose level or the blood glucose level diagnosed by 75 g OGTT after 24 weeks of gestation was greater than the critical blood glucose level [16]. In addition to GDM, the participants were routinely examined to exclude diseases such as multiple pregnancy, and the participants' age, body mass index (BMI), pregnancy period, blood glucose and other clinical data were recorded and counted. All participants were aware of this study and signed informed consent. Meanwhile, blood samples of participants in the fasting state, 1h and 2 h after glucose ingestion were collected, and serum samples were obtained after centrifugation and stored for subsequent assays.

#### Real-time quantitative PCR assay

TRIzol reagent (Invitrogen, USA) was obtained to extract total RNA from the serum of participants in the fasting state, and cDNA was obtained using the SuperScript II reverse transcriptase kit (Invitrogen, USA). With the help of SYBR® Green PCR kit (TaKaRa, Japan), the reaction system was configured with primers and cDNA as templates, and the detection was performed in 7500 RT PCR system (Applied Biosystems, USA). Glyceraldehyde phosphate dehydrogenase (GAPDH) was used as an internal reference, and the expression of HCG18 was calculated by the  $2^{-\Delta\Delta Ct}$  method.

## Blood glucose and biochemical index detection

Fasting blood glucose (FBG), 1 h or 2 h blood glucose were measured by glucose oxidase method and continuous glucose monitoring concentration system (CGMS). Triacylglycerol (TG), total cholesterol (TC), and low-density lipoprotein (LDL) were performed using automated standard routine enzymatic methods (Abbott Aeroset, USA).

## Statistical analysis

Data analysis was performed by SPSS 17.0 and GraphPad Prism 7.0 software, measurement data were expressed as mean  $\pm$  standard deviation, and differences between the two groups were tested by Student's t test. Receiver operating characteristic curve (ROC) to evaluate the clinical diagnostic value of HCG18 in GDM. Multivariate logistic analysis was used to verify the relationship between GDM and various indicators. P < 0.05 was considered

statistically significant.

#### **RESULTS**

#### Clinical data of participants

Table 1 recorded the relevant clinical data of GDM patients and healthy controls. The results showed that there was no significant difference in age, gestational age, fetal birth weight, and LDL among the participants (p > 0.05). Besides, there were significant differences in BMI, FBG, 1h blood glucose, 2h blood glucose, TG and TC between the two groups (p < 0.05).

# **LncRNA HCG18 expression was upregulated**

Compared with the control group, the relative expression level of HCG18 in the serum of GDM patients was increased via RT-qRCR assays (Fig. 1, p < 0.001).

## **Diagnostic value of HCG18**

The diagnostic value of HCG18 in GDM was evaluated by ROC curve. As shown in Figure 2, the area under the curve (AUC) was 0.916, the sensitivity was 80.5%, the specificity was 90.2%, and the 95% confidence interval (CI) was 0.880–0.952, suggesting that HCG18 had a significant performance in the diagnosis of GDM (p < 0.001). Multivariate logistic regression analysis verified the relationship between GDM and various indicators, as shown in Table 2 for details. Among all indicators, HCG18 (OR = 6.984, 95% CI = 3.751–13.005, p < 0.001) and TG (OR = 1.880, 95% CI = 1.002–3.527, p = 0.049) were significantly associated with GDM, which suggested that HCG18 may be a diagnostic factor in GDM patients.

# HCG18 expression was positively correlated with blood glucose

The relative expression of HCG18 was positively correlated with fasting blood glucose (r = 0.6146, p < 0.0001), 1h blood glucose (r = 0.6979, p < 0.0001) and 2 h blood glucose (r = 0.7418, p < 0.0001) in GDM patients (Fig. 3A–C).

#### DISCUSSION

The complex changes in glucose metabolism during pregnancy, including increased glucose requirements, increased insulin resistance, and relative insulin insufficiency, contribute to GDM in some pregnant women [17]. After discussion, it is known that the pregnant women's

age, excessive obesity, family genetic history, adverse pregnancy and birth history, hypertension and other factors will be the inducing factors of GDM, resulting in glucose metabolism disorder in pregnant women, affecting normal pregnancy and fetal health [18, 19]. With the application of molecular biology in diseases, it is not uncommon for lncRNAs to be involved in the regulation as diagnostic and prognostic factors. Li et al. [20] explored that RPL13P5 forms a co-expression chain with TSC2 gene through the PI3K-Akt signaling pathway to promote insulin resistance in GDM patients. Another report showed OIP5-AS1 as a potential biomarker of GDM and a regulator of trophoblasts [21]. As mentioned above, HCG18 can play a key regulatory role in a variety of tumors, while its predictive and diagnostic role in GDM has not been clearly reported. However, it has been reported that HCG18 is involved in the regulation of insulin, which is a key factor causing GDM [22].

Collecting the serum of participants for HCG18 expression detection in this study, we found that HCG18 was significantly elevated in GDM patients compared with normal group. Similarly, Ren et al. [23] found that HCG18 is highly expressed in a model of diabetic peripheral neuropathy. In previous GDM studies, MEG8 was also up-regulated and predicted kidney damage [24], SOX2OT was highly expressed in GDM and affected adverse events such as preterm birth and intrauterine death [25], which suggested that the overexpression of HCG18 in GDM has reference value. Fetal macrosomia is one of the most common complications of GDM. Shi and his colleagues [26] confirmed that abnormally expressed lncRNAs may play a partial or key role in the development of GDM macrosomia, providing a potential biological target for the treatment of macrosomia. Besides, excessive obesity can also aggravate the production of GDM, and high expression of lncRNA has been confirmed to be associated with BMI and obesity [27, 28]. We evaluated the significant role of HCG18 in the diagnosis of GDM by ROC curve, and the multivariate logistic regression analysis also verified the close correlation between HCG18 and GDM, suggesting that HCG18 has the diagnostic value of GDM.

Glucose level was used to reflect glucose metabolism level in GDM patients, among which fasting blood glucose, 1-hour blood glucose level and 2-hour blood glucose level are all important diagnostic criteria [29, 30]. By analyzing the relationship between the relative expression of HCG18 and blood glucose in GDM patients, we learned that HCG18 was positively correlated with fasting blood glucose, 1h blood sugar, and 2h blood glucose. It has been reported that miR-195-5p is positively correlated with fasting blood glucose, 1-hour blood glucose and 2-hour blood glucose in GDM patients, which is consistent with the trend of our study [31]. Based on this, it was confirmed that high expression of HCG18 could

reflect blood glucose level, and the expression of HCG18 was correlated with the occurrence and development of GDM.

Some GDM patients will return to normal blood sugar levels after pregnancy under reasonable management and treatment, but there are still patients who will develop type 2 diabetes or other complications, and the probability is increasing. This study used the expression level of HCG18 to effectively predict and diagnose GDM, which can provide a certain theoretical basis for the clinical treatment of GDM. However, if it is to be truly applied to the clinic, it is necessary to increase the number of samples for experimentation and verification, and we will also pay attention and implement it in the follow-up research.

#### **CONCLUSIONS**

In general, this study stated that the expression of lncRNA HCG18 is upregulated in GDM serum and confirmed that HCG18 has the potential to diagnose GDM, speculating that HCG18 may become a biomarker for GDM.

#### Article information and declarations

## Data availability statement

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

#### **Ethics statement**

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. The study was approved by the Central Hospital of Enshi Tujia and Miao Autonomous Prefecture. All participants were aware of this study and signed informed consent.

## **Author contributions**

All authors designed this study. LC and JJ conducted the experiment, analyzed the data and wrote the manuscript. YT revised the manuscript. All authors reviewed and approved for publication.

#### **Funding**

None.

## Acknowledgments

None.

## **Conflict of interest**

There is no conflict of interest in this study.

## Supplementary material

None.

#### REFERENCES

- 1. He Y, Wu Na. Research Progress on Gestational Diabetes Mellitus and Endothelial Dysfunction Markers. Diabetes Metab Syndr Obes. 2021; 14: 983–990, doi: 10.2147/DMSO.S295737, indexed in Pubmed: 33688231.
- 2. Zhang H. Mechanism associated with aberrant lncRNA MEG3 expression in gestational diabetes mellitus. Exp Ther Med. 2019; 18(5): 3699–3706, doi: 10.3892/etm.2019.8062, indexed in Pubmed: 31656536.
- 3. Cao YM, Wang W, Cai NN, et al. The Impact of the One-Day Clinic Diabetes Mellitus Management Model on Perinatal Outcomes in Patients with Gestational Diabetes Mellitus. Diabetes Metab Syndr Obes. 2021; 14: 3533–3540, doi: 10.2147/DMSO.S316878, indexed in Pubmed: 34385828.
- 4. Zhang X, Wu Y, Miao L. Study on the Effects of Individualized Nutritional Intervention on Pregnancy Outcome and Neonatal Immune Function in Patients with Gestational Diabetes Mellitus. Biomed Res Int. 2022; 2022: 3246784, doi: 10.1155/2022/3246784, indexed in Pubmed: 35036430.
- 5. Matsumoto Y, Yamada H, Yoshida M, et al. Background Factors Determining the Introduction and Dosage of Insulin in Women With Gestational Diabetes Mellitus. J Clin Med Res. 2019; 11(6): 447–451, doi: 10.14740/jocmr3824, indexed in Pubmed: 31143312.
- 6. Wang TS, Gao F, Qi QR, et al. Dysregulated LIF-STAT3 pathway is responsible for impaired embryo implantation in a Streptozotocin-induced diabetic mouse model. Biol Open. 2015; 4(7): 893–902, doi: 10.1242/bio.011890, indexed in Pubmed: 26002932.
- 7. Li YH, Hu YQ, Wang SC, et al. LncRNA SNHG5: A new budding star in human cancers. Gene. 2020; 749: 144724, doi: 10.1016/j.gene.2020.144724, indexed in Pubmed: 32360843.
- 8. Zhang Le, Li C, Su X. Emerging impact of the long noncoding RNA MIR22HG on

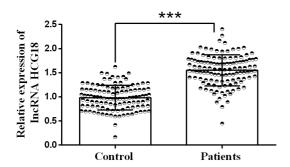
- proliferation and apoptosis in multiple human cancers. J Exp Clin Cancer Res. 2020; 39(1): 271, doi: 10.1186/s13046-020-01784-8, indexed in Pubmed: 33267888.
- 9. Lu Y, Guo J, Zhu S, et al. LncRNA HCG18 is critical for vascular smooth muscle cell proliferation and phenotypic switching. Hum Cell. 2020; 33(3): 537–544, doi: 10.1007/s13577-020-00366-2, indexed in Pubmed: 32449112.
- 10. Zheng Z, Lin K. LncRNA HCG18 promotes cell multiplication and metastasis by miR-148b/ETV5 regulation in osteosarcoma. Am J Transl Res. 2021; 13(7): 7783–7793, indexed in Pubmed: 34377255.
- 11. Xin L, Wu Y, Liu C, et al. Exosome-mediated transfer of lncRNA HCG18 promotes M2 macrophage polarization in gastric cancer. Mol Immunol. 2021; 140: 196–205, doi: 10.1016/j.molimm.2021.10.011, indexed in Pubmed: 34735868.
- 12. Liu Xu, Qiao K, Zhu K, et al. Long Noncoding RNA HCG18 Promotes Malignant Phenotypes of Breast Cancer Cells the HCG18/miR-103a-3p/UBE2O/mTORC1/HIF-1 $\alpha$ -Positive Feedback Loop. Front Cell Dev Biol. 2021; 9: 675082, doi: 10.3389/fcell.2021.675082, indexed in Pubmed: 34976998.
- 13. Yang Yu, Gong P, Yao D, et al. LncRNA Promotes Clear Cell Renal Cell Carcinoma Progression by Targeting to Upregulate . Cancer Manag Res. 2021; 13: 2287–2294, doi: 10.2147/CMAR.S298649, indexed in Pubmed: 33732021.
- 14. Zou Y, Sun Z, Sun S. LncRNA HCG18 contributes to the progression of hepatocellular carcinoma via miR-214-3p/CENPM axis. J Biochem. 2020; 168(5): 535–546, doi: 10.1093/jb/mvaa073, indexed in Pubmed: 32663252.
- 15. Li L, Ma TT, Ma YH, et al. LncRNA HCG18 contributes to nasopharyngeal carcinoma development by modulating miR-140/CCND1 and Hedgehog signaling pathway. Eur Rev Med Pharmacol Sci. 2019; 23(23): 10387–10399, doi: 10.26355/eurrev 201912 19678, indexed in Pubmed: 31841193.
- 16. Xu D, You J, Chen G, et al. Changes of Serum Zinc-2-Glycoprotein Level and Analysis of Its Related Factors in Gestational Diabetes Mellitus. J Diabetes Res. 2021; 2021: 8879786, doi: 10.1155/2021/8879786, indexed in Pubmed: 33681385.
- 17. Yang S, Lin R, Si L, et al. Cod-Liver Oil Improves Metabolic Indices and hs-CRP Levels in Gestational Diabetes Mellitus Patients: A Double-Blind Randomized Controlled Trial. J Diabetes Res. 2019; 2019: 7074042, doi: <a href="https://doi.org/10.1155/2019/7074042">10.1155/2019/7074042</a>, indexed in Pubmed: 31956660.
- 18. Bień A, Rzońca E, Kańczugowska A, et al. Factors Affecting the Quality of Life and the Illness Acceptance of Pregnant Women with Diabetes. Int J Environ Res Public Health. 2015;

- 13(1): ijerph13010068, doi: 10.3390/ijerph13010068, indexed in Pubmed: 26703697.
- 19. Furukawa S, Kobayashi Y. Leaner Women with Impaired Insulin Secretion Accounts for about 40% of Gestational Diabetes Mellitus in Japan. J Pregnancy. 2019; 2019: 7578403, doi: 10.1155/2019/7578403, indexed in Pubmed: 31275653.
- 20. Li Y, Cheng X, Li D. LncRNA RPL13p5 gene expression promotes insulin resistance in patients with gestational diabetes. Ann Palliat Med. 2021; 10(10): 11024–11034, doi: 10.21037/apm-21-2940, indexed in Pubmed: 34763465.
- 21. Li Y, Liu L. LncRNA OIP5-AS1 Signatures as a Biomarker of Gestational Diabetes Mellitus and a Regulator on Trophoblast Cells. Gynecol Obstet Invest. 2021; 86(6): 509–517, doi: 10.1159/000520340, indexed in Pubmed: 34844256.
- 22. Xia Yu, Zhang Y, Wang H. Upregulated lncRNA HCG18 in Patients with Non-Alcoholic Fatty Liver Disease and Its Regulatory Effect on Insulin Resistance. Diabetes Metab Syndr Obes. 2021; 14: 4747–4756, doi: 10.2147/DMSO.S333431, indexed in Pubmed: 34887672.
- 23. Ren W, Xi G, Li X, et al. Long non-coding RNA HCG18 promotes M1 macrophage polarization through regulating the miR-146a/TRAF6 axis, facilitating the progression of diabetic peripheral neuropathy. Mol Cell Biochem. 2021; 476(1): 471–482, doi: 10.1007/s11010-020-03923-3, indexed in Pubmed: 32996080.
- 24. Zhang W, Cao D, Wang Y, et al. LncRNA MEG8 is upregulated in gestational diabetes mellitus (GDM) and predicted kidney injury. J Diabetes Complications. 2021; 35(1): 107749, doi: 10.1016/j.jdiacomp.2020.107749, indexed in Pubmed: 33189541.
- 25. Ran G, Zhu X, Qin Y. LncRNA SOX2OT is Upregulated in Gestational Diabetes Mellitus (GDM) and Correlated with Multiple Adverse Events. Diabetes Metab Syndr Obes. 2021; 14: 3989–3995, doi: 10.2147/DMSO.S319739, indexed in Pubmed: 34531671.
- 26. Shi Z, Zhao C, Long W, et al. Microarray Expression Profile Analysis of Long Non-Coding RNAs in Umbilical Cord Plasma Reveals their Potential Role in Gestational Diabetes-Induced Macrosomia. Cell Physiol Biochem. 2015; 36(2): 542–554, doi: 10.1159/000430119, indexed in Pubmed: 25997527.
- 27. Yuan Y, Cao X, Hu J, et al. The role and possible mechanism of lncRNA AC092159.2 in modulating adipocyte differentiation. J Mol Endocrinol. 2019; 62(3): 137–148, doi: 10.1530/JME-18-0215, indexed in Pubmed: 30753134.
- 28. Lekva T, Norwitz ER, Aukrust P, et al. Impact of Systemic Inflammation on the Progression of Gestational Diabetes Mellitus. Curr Diab Rep. 2016; 16(4): 26, doi: 10.1007/s11892-016-0715-9, indexed in Pubmed: 26879309.
- 29. Hu S, Yan J, You Y, et al. Association of polymorphisms in STRA6 gene with gestational

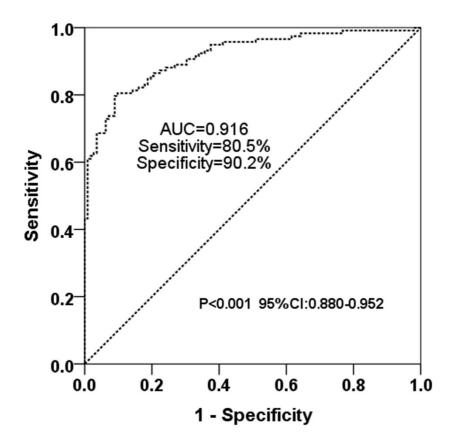
diabetes mellitus in a Chinese Han population. Medicine (Baltimore). 2019; 98(11): e14885, doi: 10.1097/MD.0000000000014885, indexed in Pubmed: 30882700.

30. Wei Q, Pu X, Zhang Li, et al. Expression of Dual-Specificity Phosphatase 9 in Placenta and Its Relationship with Gestational Diabetes Mellitus. J Diabetes Res. 2019; 2019: 1963178, doi: 10.1155/2019/1963178, indexed in Pubmed: 31772940.

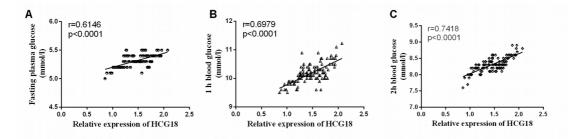
31. Wang J, Pan Y, Dai F, et al. Serum miR-195-5p is upregulated in gestational diabetes mellitus. J Clin Lab Anal. 2020; 34(8): e23325, doi: 10.1002/jcla.23325, indexed in Pubmed: 32301163.



**Figure 1.** The relative expression level of HCG18 in serum of gestational diabetes mellitus (GDM) patients was increased compared with healthy controls; p < 0.001, compared with the controls



**Figure 2.** Receiver operating characteristic curve (ROC) curve of HCG18; AUC — area under the curve; CI — confidence interval



**Figure 3.** HCG18 expression was positively correlated with blood glucose; **A.** The expression of HCG18 was positively correlated with fasting blood glucose (r = 0.6146, p < 0.0001); **B.** The expression of HCG18 was positively correlated with 1h blood sugar (r = 0.6979, p < 0.0001); **C.** The expression of HCG18 was positively correlated with 2 h blood sugar (r = 0.7418, p < 0.0001)

Table 1. Comparison of clinical data between gestational diabetes and healthy individuals

	T . 1 ( DDO)		
	Total (n = 230)	Total $(n = 230)$	
Paraments	Control	Patients	p value
	(n = 112)	(n = 118)	_
Age [years]	$29.57 \pm 0.97$	$29.48 \pm 0.93$	0.439
BMI [kg/m²]	$24.65 \pm 2.78$	$25.94 \pm 3.12$	0.001
Gestational age [weeks]	$26.89 \pm 1.03$	$27.05 \pm 1.46$	0.334
FBG [mmol/L]	$4.45 \pm 0.31$	$5.30 \pm 0.11$	< 0.001
1 h blood glucose	$6.32 \pm 0.20$	$10.20 \pm 0.30$	< 0.001
[mmol/L]			
2 h blood glucose	$5.51 \pm 0.40$	$8.29 \pm 0.20$	< 0.001
[mmol/L]			
Fetal birth weight [g]	3426.38 ±	$3448.92 \pm 178.91$	0.462
	276.75		
TG [mg/dL]	$1.00 \pm 0.28$	$1.34 \pm 0.33$	< 0.001
TC [mg/dL]	$5.45 \pm 0.34$	$5.89 \pm 0.45$	< 0.001
LDL [mg/dL]	$2.63 \pm 0.36$	$2.72 \pm 0.51$	0.110

BMI — body mass index; FBG — fasting blood glucose; TG — triglycerides; TC — total cholesterol; LDL — low density lipoprotein; Data are expressed as n or mean ± standard deviation (SD)

Table 2. Relationship between gestational diabetes mellitus and various indicators

Indicators	Multivariate logistic analysis			
	OR	95% CI	p value	
LncRNA HCG18	6.984	3.751-13.005	< 0.001	
Age [years]	1.410	0.758-2.621	0.277	
BMI [kg/m²]	1.733	0.922-3.257	0.088	
Gestational age [weeks]	1.173	0.623-2.211	0.621	
Fetal birth weight [g]	1.308	0.702-2.437	0.398	
TG [mg/dL]	1.880	1.002-3.527	0.049	
TC [mg/dL]	1.416	0.766-2.620	0.267	
LDL [mg/dL]	1.216	0.659-2.243	0.531	

CI — confidence interval; BMI — body mass index; TG — triglycerides; TC — total cholesterol; LDL — low density lipoprotein