

Folia Morphol. Vol. 80, No. 4, pp. 870–880 DOI: 10.5603/FM.a2020.0130 Copyright © 2021 Via Medica ISSN 0015–5659 eISSN 1644–3284 journals.viamedica.pl

Concomitant administration of sitagliptin and rutin improves the adverse hepatic alterations in streptozotocin-induced diabetes mellitus in albino rats: an overview of the role of alpha smooth muscle actin

R.A. Attia, S. Abdel Fattah, M.M. Nasralla

Department of Anatomy and Embryology, Faculty of Medicine, Cairo University, Cairo, Egypt

[Received: 18 September 2020; Accepted: 7 October 2020; Early Publication date: 26 October 2020]

Background: Diabetes mellitus (DM), one of the commonest worldwide metabolic conditions, is believed to be associated with an imbalance between oxidants and antioxidants. Sitagliptin is an oral anti-hyperglycaemic drug that blocks dipeptidyl peptidase 4 (DPP4). Rutin is a polyphenolic natural flavonoid which has antioxidant and anti-proliferative activity. The aim of the present work is to elucidate the concomitant effect of sitagliptin and rutin on the deleterious alterations in the liver of experimentally induced diabetes in rats.

Materials and methods: Fifty adult male albino rats, weighing 170–200 g were used. Rats were randomly divided into five groups (n = 10): group 1 (control group), the other four groups (groups II, III, IV and V) received a single i.p. injection of streptozotocin, 65 mg/kg body weight to induce diabetes; group II (diabetic), group III (diabetic and rutin administered), group IV (diabetic and sitagliptin administered), and group V (diabetic with sitagliptin and rutin concomitantly administered). Haematoxylin and eosin, Masson trichrome, periodic acid Schiff, immune-histochemistry: α -smooth muscle actin (α -SMA), histomorphometric analysis, liver enzymes and oxidants/anti-oxidants; malondialdehyde/glutathione and were done.

Results: Distorted hepatic architecture, dilatation, congestion of sinusoids and central veins as well as cytoplasmic vacuolations were remarkable changes in the diabetic group. There was extravasation of blood, diffuse fibrous tissue formation, increase in the mean values of liver enzymes, oxidative markers and α -SMA expression in the same group. The aforementioned changes were ameliorated in groups III and IV. Concomitant administration of sitagliptin and rutin resulted in marked enhancement of these hepatic alterations.

Conclusions: Combination of sitagliptin and rutin has an ameliorating effect on the hepatic deterioration induced by diabetes, which is better than either sitagliptin or rutin alone. (Folia Morphol 2021; 80, 4: 870–880)

Key words: streptozotocin, liver, diabetes, alpha smooth muscle actin, sitagliptin, rutin

Address for correspondence: S. Abdel Fattah, MD, Department of Anatomy and Embryology, Faculty of Medicine, Cairo University, Cairo, Egypt, e-mail: dr_shery.fattah@yahoo.com

This article is available in open access under Creative Common Attribution-Non-Commercial-No Derivatives 4.0 International (CC BY-NC-ND 4.0) license, allowing to download articles and share them with others as long as they credit the authors and the publisher, but without permission to change them in any way or use them commercially.

INTRODUCTION

Diabetes mellitus (DM) is a metabolic syndrome categorized by elevated blood glucose levels and typical symptoms; polydipsia, polyuria and polyphagia [26]. Abnormal function of chief body organs comprising the liver can be a consequence of the upsurge of blood glucose levels [31]. The pivotal role of reactive oxygen species in the progress and exacerbation of DM has been discussed for several epochs [18, 25]. Lipid peroxidation disturbs all lipid-encompassing structures in cells, resulting in cytopathological consequences [29].

Rutin is a flavonoid compound that exists in various plants and possesses several pharmacological functions: blood glucose reducer, insulin release regulator, and dyslipidaemic modifier. Moreover, it owns anti-inflammatory and anti-tumour properties as well as attenuates the damage caused by reactive oxygen species (ROS) [13, 15, 27, 33].

Many studies have revealed that rutin has a robust therapeutic influence on liver injury triggered by different reasons for instance: biliary obstruction and high fatty diet. Nevertheless, the mechanism of rutin in DM-induced liver injury was not very distinct [24, 34]. Some researchers suggest that the safety of rutin in diabetic liver can be attributed to its anti-inflammatory properties, impeding lipogenesis [16].

Sitagliptin is an antidiabetic prescription drug taken orally that blocks dipeptidyl peptidase 4 (DPP4). Suppression of DPP4 increases insulin sensitivity and hence, lowers blood glucose concentrations [32]. Sitagliptin definitely has been approved by the Food and Drug Administration, Health Canada, as well as the European Commission for the treatment of diabetes. It can be used alone or with either metformin or glitazone [2, 4].

The objective of this study was to elucidate the effects of the combination of sitagliptin and rutin on the pathological alterations in the liver in experimentally induced diabetes in rats.

MATERIALS AND METHODS

Chemicals

Streptozotocin (STZ) (trade name Zanosar) was purchased from Sigma Chemical Company, St. Louis Missouri, USA, in the form of 1 g vials. The drug was dissolved in 0.1 M sodium citrate (pH adjusted to 4.5).

Sitagliptin (in the form of Januvia 100 tablet). Each tablet was ground and dissolved in 10 mL solution of 0.5% carboxymethyl cellulose, and afterwards shaken to obtain a suspension form (10 mg/mL). Rutin was purchased from Sigma Chemical Company, St. Louis, Missouri, USA, in the form of powder and dissolved in saline.

Experimental animals

Fifty adult male Sprague-Dawley albino rats, weighing 170–200 g were used. They were kept in the animal house of Kasr Al-Aini Faculty of Medicine, Cairo University, Egypt. The rats had free access to standard rat chow and water. They were maintained according to the standard guidelines of the Institutional Animal Care and Use Committee, subsequent to the Institutional Review Board approval. The rats were permitted to accustom for 2 weeks prior to the experiment.

The rats were randomly divided into five groups (n = 10).

Group 1 (control) received an intraperitoneal (i.p.) injection of 0.1 mol/L sodium citrate buffer (pH 4.5).

The other four groups (groups II, III, IV, V) received a single i.p. injection of STZ, 65 mg/kg body weight [10], freshly dissolved in 0.1 M citrate buffer (pH 4.5). Diabetes mellitus was verified by measuring blood glucose levels (after overnight fast) with the use of glucose oxidase reagent strips (Lif3 scan, Milpitas, CA, USA). Rats with blood sugar level > 250 mg/dL were used as the diabetic group. In order to monitor blood glucose levels, blood glucose was tested every week for 4 weeks.

Group II (diabetic): diabetic rats received no treatment during the course of the study.

Group III (diabetic + rutin): diabetic rat received rutin at a dose of 10 mg/kg/day dissolved in saline orally for 4 weeks [34].

Group IV (diabetic + sitagliptin): diabetic rats received oral sitagliptin at a dose of 100 mg/kg/day sitagliptin via gastric gavage for 4 weeks [28].

Group V (diabetic + rutin+ sitagliptin): diabetic group receiving oral sitagliptin at a dose of 100 mg/ /kg/day concomitantly with rutin at a dose of 10 mg/ /kg/day orally via gastric gavage for 4 weeks.

All animals were clinically monitored and weighed on a weekly basis. After 4 weeks a blood sample was withdrawn from the tail vein, using a fine heparinized capillary tube, for the assessment of liver function. Formerly, the rats of each group were sacrificed with an overdose of intraperitoneal phenobarbital sodium (40 mg/kg). The rats were dissected. The liver of each animal was excised and prepared for light microscopic study. Liver specimens were fixed in 10% formalin dehydrated in ethyl alcohol, cleared in xylol and embedded in paraffin wax. Sections of 5 μ m thickness were cut and mounted on glass slides. Other sections were mounted on positive charged slides for immunohistochemistry. These sections were subjected to the following.

Light microscopic study

- Haematoxylin and eosin (H&E) stain to study the changes in histopathological architecture.
- Masson's trichrome stain to demonstrate collagen fibres.

Histochemical study

Periodic acid Schiff (PAS) reaction to demonstrate the glycogen. The paraffin sections were dewaxed, rehydrated and then oxidized in 1% of periodic acid (5 min). Formerly, they were wash away with distilled water, pickled with Schiff's reagent for 15 min, rinsed in tap water for 5–10 min, counterstained in haematoxylin, discerned in 1% acid-alcohol, cleansed in tap water, dehydrated in ascending degrees of alcohol, cleared in xylene, and mounted in Canada balsam. Glycogen and other reactive carbohydrates appeared magenta.

Immunohistochemical staining

Alpha smooth muscle actin (α -SMA) to evaluate the fibrosis processes [5]. The segments from each paraffin block were incubated with primary antibody α -SMA antibody [(ab7817) 1:100]. Next, the sections were incubated with Goat anti-Rabbit IgG H&L (HRP) (ab205718) for 20 min at 37°C. Each phase was tailed by satisfactorily wash with phosphate buffered saline (PBS).

Morphometric study

Using Leica image analysis computer system (software Qwin 500, Switzerland), the following parameters were assessed:

- area percentage of collagen fibres in Masson's trichrome;
- mean optical density of PAS reaction in PAS;
- area percentage of immune reaction of α -SMA.

Stained sections were inspected by magnification $\times 400$ and measured within a field of standard measuring frame. This was completed in 10 non overlapping microscopic fields of each specimen and their mean values were acquired.

Biochemical study

Liver function assay. Retro-orbital blood samples were extracted from each rat for valuation of liver enzymes. Alkaline phosphatase (ALP), aspartate aminotransferase (AST) and alanine aminotransferase (ALT) enzymes levels were assessed using specific kits provided by Bio-diagnostic Company (Bio-diagnostic eka@lycos.com and info@bio-diagnostic. com). These measurements were done in the Biochemistry Department, Faculty of Medicine, Cairo University, Egypt.

Assays of oxidative/antioxidative markers. We measured malondialdehyde (MDA) using a commercial kit (Biodiagnostic, Cairo, Egypt), according to the manufacturer's instructions. Concisely, roughly 10-20 mg liver tissue was homogenised in 1 mL PBS, pH 7.0, utilising a micropestle in a micro tube. At that point, 20% (w/v) trichloroacetic acid was supplemented to the homogenate to precipitate the protein, and centrifuged at 12,000 g for 15 min. Afterwards, 0.8% thiobarbituric acid solution was added to the liver homogenate to precipitate the protein. After boiling for 10 min in a water bath, the absorbance was measured at 405 nm using a spectrophotometer. The concentration of MDA was calculated per milligram protein using a standard curve. The standard curve was prepared as follows. We dissolved 25 μ L 1,1,3,3--tetraethoxypropane (TEP) in 100 mL water to attain a 1 mM stock solution. We organized a working standard by hydrolysis of 1 mL TEP stock solution in 50 mL 1% sulfuric acid and incubation for 2 h at room temperature. The resultant MDA 20 nmol/mL standard was diluted with 1% sulfuric acid to yield the final concentrations of 10.5, 2.5, 1.25 and 0.625 nmol/mL to prepare a standard curve for estimating total MDA. Then, 0.250 mL standard were mixed with $25 \,\mu\text{L}$ DNPH solution and incubated for 10 min. A 20 μ L volume of the reaction mixture was injected directly onto HPLC system.

Glutathione (GSH) the antioxidant stress marker was measured using a commercial kit to detect glutathione (Biodiagnostic, Cairo, Egypt), according to the producer's instructions. The measurement was based on reduction of 5,5-dithiobis-2 nitrobenzoic acid, with reduced glutathione to create yellow compound. The reduced chromogen was directly proportional to GSH levels, and the ultimate reaction product was assayed spectrophotometrically by quantifying its absorbance at 405 nm.



Figure 1. A photomicrograph of sections of liver. A, B (control); A. Hepatic lobules and central vein (C); B. The portal triad consisting of branch of portal vein (P), bile ductule (B) and branch of hepatic artery (A); C, D, E (diabetic); C. Loss of hepatic architecture with marked dilatation of central vein (C), cytoplasmic vacuolations in the hepatocytes (arrows) and vacuoles (V) in between the hepatocyte; D. Dilatation of blood sinusoids (arrows) and degeneration of hepatocyte (arrow head); E. Marked congestion of portal vein (P) with mononuclear cell infiltration (arrows); F (diabetic and rutin-treated): Marked dilatation of central vein (C) and vacuolated areas (V); G (diabetic and sitagliptin-treated): Dilatation of central vein with minimal vacuolation (arrows); H (diabetic + rutin + sitagliptin-treated): Minimal dilatations of central vein (H&E ×400).

Statistical analysis

The statistical package for the social science (SPSS) was used for data analysis. The data obtained from image analyser were summarised as means and standard deviations and compared using one-way analysis of variance (ANOVA). P values < 0.05 were considered statistically significant, while p < 0.01 was considered statistically highly significant.

RESULTS

Light microscopic study

Haematoxylin and eosin results. Liver sections obtained from rats of the control group showed classic hepatic lobules consisting of intersecting plates of liver cells (hepatocyte) radiating outwards from a central vein to the periphery of the lobules. Narrow blood sinusoids were seen intervening between cords of hepatocyte (Fig. 1A). The portal area at the periphery of the lobules was seen formed of the bile ductule, a branch of portal vein with thin wall and wide lumen and branch of hepatic artery which appeared narrower in lumen and thicker in wall (Fig. 1B). The diabetic group (group II) revealed distortion of the parenchymal architecture. The hepatocytes exhibited marked pathologic affection where the cytoplasm displayed marked degree of cytoplasmic vacuolations. The central vein was dilated and congested. There was marked congestion of portal vein with mononuclear cell infiltration in the portal area (Fig. 1C-E). The liver in H&E sections in the diabetic and rutin-treated group (group III) revealed dilatation of central vein with vacuolated areas of degenerated hepatocytes (Fig. 1F). The diabetic and sitagliptin-treated group (group IV) exposed dilatation of central vein with small vacuolated areas between the hepatocytes (Fig. 1G). Concomitant administration of rutin and sitagliptin (group V) resulted in an apparently normal hepatic architecture as that of the control group apart from



Figure 2. Photomicrographs of sections of liver. **A** (control): Minimal amount of collagen deposition (arrow) around the central vein; **B** (diabetic): Increased collagen fibres deposition (arrows) in the portal area. Note the marked engorged portal vein (P); **C** (diabetic and rutin-treated): Increased collagen fibres deposition (arrows); **D** (diabetic and sitagliptin-treated): Moderate collagen fibres deposition (arrows); **E** (diabetic + rutin- + sitagliptin-treated): Minimal collagen fibres deposition (arrows); **F**. Area percentage of collagen content in liver following administration of rutin and/or sitagliptin to diabetic rats; *Significantly different from control at p < 0.05; #Noticeably different from diabetic p < 0.05(Masson's trichrome $\times 400$).

mild affections. Most hepatocytes were apparently normal with eosinophilic cytoplasm and rounded nuclei. There was mild dilatation of central vein (Fig. 1H).

Masson's trichrome results. Histological examination of sections of the liver from rats of group I showed minimal amount of collagen fibres in the form of a thin layer of collagen fibres around the central vein and hepatic sinusoids (Fig. 2A). Group II exhibited an increase in the amount of dense collagen fibres around the portal tract and the blood sinusoids (Fig. 2B) and in between hepatocytes (Fig. 2B). Meanwhile, group III and IV displayed moderate increase in the amount of dense collagen fibres around the portal tract and the blood sinusoids (Fig. 2C, D). On the other hand, group V showed mild increase in the amount of collagen fibres around the central vein and in the portal tract (Fig. 2E).



Figure 3. Photomicrographs of sections of liver. **A** (control): Strong positive PAS reaction of the hepatocyte around the central vein (C); **B** (diabetic): Weak positive PAS reaction of the hepatocyte; **C** (diabetic and rutin-treated): Moderate positive PAS reaction of the hepatocyte; **D** (diabetic and sitagliptin-treated): Moderate positive PAS reaction of the hepatocyte; **E** (diabetic + rutin- + sitagliptin-treated): Strong positive PAS reaction of the hepatocyte around the central vein (C); **F**. Optical density in liver following administration of rutin and/or sitagliptin to diabetic rat; *Significantly different from control at p < 0.05; *Noticeably different from diabetic p < 0.05 (PAS ×400).

Histochemical study

PAS reaction results. In the control group the hepatocytic cytoplasm contained considerable amounts of glycogen and displayed strong positive PAS reaction in the form of small red granules filling the cytoplasm (Fig. 3A), while group III exposed very faint weak positive PAS reaction (Fig. 3B). Group IV displayed weak positive and group IV showed moderate positive PAS reaction (Fig. 3D). However, group V revealed strong positive PAS reaction in the cytoplasm of hepatocyte (Fig. 3E).

Immune-histochemical results

Examination of the liver sections of group I revealed negative immunoreactivity of α -SMA in the hepatocyte. There was localised immunoreactivity around the central vein (Fig. 4A). Group II and III showed strong positive immune-reactivity



Figure 4. Photomicrographs of sections of liver. **A** (control): Localised positive immune-reactivity around central vein (arrows) and negative immune-reactivity in other parts; **B**, **C** (diabetic): Strong positive immune-reactivity; **D** (diabetic and rutin-treated group): Positive immune-reactivity; **E** (diabetic and sitagliptin-treated): Moderate positive immunoreactivity; **F** (diabetic + rutin- + sitagliptin-treated): Weak positive immunoreactivity (arrows); **G**. Area percentage of alpha smooth muscle actin (α -SMA) in liver following administration of rutin and/or sitagliptin to diabetic rats; *Significantly different from control at p < 0.05; *Noticeably different from diabetic p < 0.05 (α -SMA ×400).

(Fig. 4B, C). Group IV exposed moderate positive PAS reaction and moderate positive immunoreactivity (Fig. 4D). Nevertheless, group V showed minimal positive immune-reactivity (Fig. 4E).

Morphometric results

Statistical study of the mean area percentage of collagen fibres. The mean area percentage of the collagen fibres of group II, III and IV showed highly significant increase in its value compared with the corresponding control group. However, the mean value of the above mentioned parameter of group V was non-significant compared with the corresponding control group. The mean value of the area percentage of the collagen fibres of group II, III and IV increased significantly in comparison with group V (Table 1, Fig. 2F).

Statistical study of the mean optical density of PAS reaction. A significant increase in the optical density was found in group II and III in comparison with the corresponding control group. On the other hand, a significant decrease in the optical density was found in group IV and V in comparison with the group II (Table 1, Fig. 3F).

Statistical study of the area percentage of immune reaction of α -SMA. The mean area percentage of α -SMA immune-positive cells showed no significant difference among the control group and group V. While diabetic group (group II) resulted in significant

Groups	Area % of collagen fibres	Optical density	Area % of α -SMA
Group I (control)	1.89 ± 0.16	0.812 ± 0.065	1.36 ± 0.12
Group II (diabetics)	15.74ª ± 1.17	$0.113^{a} \pm 0.011$	$7.45^{\circ}\pm0.47$
Group III (rutin)	$10.99^{a,b}\pm 1.06$	$0.339^{a,b}\pm 0.016$	$5.86^{a,b} \pm 0.43$
Group IV (sitagliptin)	$7.19^{a,b}\pm 0.49$	$0.565^{\mathrm{a,b}}\pm0.031$	$3.44^{a,b} \pm 0.21$
Group V (rutin + sitagliptin)	$3.19^{\text{b}} \pm 0.21$	$0.765^{\text{b}} \pm 0.041$	$1.44^{\text{b}} \pm 0.11$

Table 1. Area percentage of collagen fibres, optical density and area percentage of alpha smooth muscle actin (α-SMA) immune-reaction in different experimental groups

Data is shown as mean \pm standard error of the mean, n = 10. Multiple comparisons were made using ANOVA one-way test followed by Tukey-Kramer as a post-hoc test. *Certainly different from control p < 0.05; *Significantly different from diabetic p < 0.05

Table	2.	Mean	values	of liver	enzymes	in	different	experimental	arour	าร
Tubic	~ .	wicun	vuluuu	01 11 001	Chizynnos		uniterent	experimental	group	10

Groups	AST [IU/L]	ALT [IU/L]	ALP [IU/L]
Group I (control)	89.24 ± 4.1	36.18 ± 2.3	71.22 ± 6.2
Group II (diabetics)	$152.16^{a} \pm 8.2$	$74.14^{a} \pm 3.4$	$162.32^{a} \pm 8.4$
Group III (rutin)	$125.99^{a,b} \pm 1.06$	$59.39^{\mathrm{a,b}}\pm0.016$	$115.86^{a,b}\pm 0.43$
Group IV (sitagliptin)	$112.15^{a,b} \pm 0.49$	$51.55^{\mathrm{a,b}}\pm0.031$	$89.44^{a,b} \pm 0.21$
Group V (rutin + sitagliptin)	$94.12^{b} \pm 4.1$	$40.18^{\text{b}} \pm 2.6$	$78.28^{b} \pm 9.2$

Data is shown as mean \pm standard deviation, n = 10. Multiple comparisons were made using ANOVA one-way test followed by Tukey-Kramer as a post-hoc test. *Certainly different from control p < 0.05; *Significantly different from diabetic p < 0.05; AST — aspartate aminotransferase; ALT — alanine aminotransferase; ALP — alkaline phosphatase

increase) in the mean area percentage of the α -SMA immunoreactivity as compared with groups I and V (Table 1, Fig. 4G).

Biochemical assay

Liver enzymes

Biochemical assay of the liver enzymes of the diabetic group revealed a marked increase in AST mean value which was statistically significant compared with the mean values of group I and groups IV and V. Meanwhile, treatment with sitagliptin and rutin in group V displayed a decrement in AST mean value which was statistically non-significant compared with the corresponding values in the control group.

Group II (diabetic group) showed marked increases in ALT mean value, which was statistically significant compared with the mean values of the control group and sitagliptin- and rutin-treated groups. Meanwhile, treatment with sitagliptin and rutin in group V showed reduction in ALT mean value which was insignificant in relation with the control group.

The diabetic group showed a marked increase in ALP mean value which was statistically significant compared with the mean value of the control, and sitagliptin- and rutin-treated groups. On the other hand, treatment with sitagliptin and rutin in group III showed a reduction in ALP mean value which was statistically significant compared with diabetic group and non-significant compared with the value in the control group (Table 2).

Assay of oxidant and antioxidant markers

Malondialdehyde. The liver homogenates of diabetic group demonstrated a marked upsurge in MDA mean value which was statistically significant compared with the mean value of the control group. Meanwhile, treatment with sitagliptin and rutin showed a reduction in MDA mean value compared with the same values in group II, and was statistically non-significant compared with the values in the control group (Table 3).

Glutathione. The liver homogenates of group II demonstrated a marked decrease in GSH mean value which was statistically significant compared with the mean value of the control group. On the other hand, treatment with sitagliptin and rutin demonstrated an increase in the mean value, which was statistically non-significant compared with the value of the control group (Table 3).

Table 3.	Mean values	of oxidative/	antioxidative/	markers in
different	experimental	groups		

Groups	Malondialdehyde [nmol/g protein]	Glutathione [µmol/g protein]
Group I (control)	18.62 ± 2.4	0.28 ± 0.02
Group II (diabetics)	$42.18^{\circ} \pm 3.6$	$0.12^{\text{a}}\pm0.02$
Group III (rutin)	$30.99^{a,b}\pm 1.06$	$0.19^{\scriptscriptstyle a,b}\pm0.016$
Group IV (sitagliptin)	$26.14^{\scriptscriptstyle a,b}\pm0.49$	$0.20^{\scriptscriptstyle a,b}\pm0.031$
Group V (rutin + sitagliptin)	$20.16^{\text{b}}\pm1.8$	$0.22^{\text{b}}\pm0.01$

Data is shown as mean \pm standard deviation, n=10. Multiple comparisons were made using ANOVA one-way test followed by Tukey-Kramer as a post-hoc test. *Certainly different from control p<0.05; *Significantly different from diabetic p<0.05

DISCUSSION

In the present study, manifestations of the pathological effects of diabetes on the liver were observed. These histopathological effects include distorted hepatic architecture, dilatation and congestion of central veins, and hepatocytic degeneration in the form of cytoplasmic vacuolation. These alterations in the liver were attributed to cellular necrosis and inflammation, which might be a result of amplified mitochondrial oxidative stress. The later stress could be a consequence of triglyceride metabolism and the establishment of free radicals in peroxisomes [17, 22]. The cytoplasmic vacuolation of hepatocytes is due to deprivation of the ATP energy stocks, which is a prerequisite for sustaining ionic and fluid homeostasis [30]. The aforementioned mechanism results in reduced activity of the energy-dependent sodium pump plasma membrane. The failure of this active transportation system grounds sodium to cross the threshold and accumulate within the cells and potassium to blowout followed by gain of water, triggering cellular swelling [23]. In addition, high deliberations release of ROS caused by suppressed oxidative phosphorylation predictably contribute to depletion of ATP [28].

Histomorphometric studies exhibited diminished PAS reaction (glycogen content) in the hepatocytes of the STZ-treated animals. Glycogen dislodgment in the cytoplasm of the hepatocytes might be due to the accumulation of lipid droplets [6].

In the present study, fibrosis is an obvious manifestation in the diabetic group in the form of increased collagen fibres around central vein and portal area and increased expression of α -SMA. Beta-oxidation of fatty might occur due to inadequate insulin, and this leads to accumulation of hydrogen peroxide in tissues [8, 12, 21]. The existence of collagen in the presinusoidal spaces may impede blood supply to liver cells and reduce metabolic exchange, probably leading to hepatocellular dysfunction and necrosis [11]. The deposition of collagen in the liver can be accredited to hepatic stellate cells (HSCs) that are involved in pathogenesis by liver damage-dependent activation. Activated HSCs differentiate into myofibroblasts [7]. Myofibroblasts blasts exhibit a synthesis profile that lead to increase in their deposition in the extracellular matrix. This process helps them to proliferate, alters their morphology, and increases contractility by activating fibre formation of smooth muscle actin. Then, this contributes to the constriction of sinusoidal blood flow and increases collagen fibres synthesis and release.

In the existing research, tests for liver function were conducted to observe the effects of STZ-induced diabetes on the liver at 4 weeks after STZ treatment. Compared with control rats, levels of AST, ALT, and ALP were increased in diabetic rats. AST and ALT are both enzymes that are established principally in liver mitochondria [32]. If there is liver impairment the enzymes are released into the bloodstream after death of the liver cells [14]. Peculiarly, high levels of AST and ALT are pivotal indicators of hepatic injury [29].

The present work revealed increment in MDA and decreased GSH in liver homogenates of diabetic rats. Recent studies have shown that the cause of DM advance and its complications is lipid peroxidase which leads to ROS formation [19]. An increase in ROS generation and a decrease in antioxidant system activity result in an imbalance that leads to oxidative hassle [20]. The high blood glucose levels in diabetes lead to oxidative stress and diminish the capacity of endogenous antioxidants. This is due to the production of many reducing sugars over both the glycolytic and polyol pathways [20].

In experimentally induced diabetic rats treated with oral intake of rutin (group III) or sitagliptin (group IV), microscopic examination of the liver disclosed variable microscopic changes in the form of dilated congested central veins and cellular infiltrations at the region of portal tract and some vacuolated areas. The allocations of collagenous fibres were slightly decreased compared with the control group.

In the current study co-administration of sitagliptin and rutin in group V significantly improved the histological picture of the liver and the severity of liver damage was lower as compared with the group treated with either sitagliptin or rutin alone.

Sitagliptin is an oral anti-diabetes drug known as an inhibitor of DPP4 used to treat type II DM [3]. DPP4 inhibition was proposed to reduce hepatic lipogenesis by several mechanisms: down-regulating the gene expression of sterol regulatory factor binding protein-1c (SREBP-1c), constraining fatty acid synthase, lowering the serum levels of very low density lipoprotein and low density lipoprotein cholesterol, subsequently decreasing hepatic lipid accumulation and steatosis [24]. In addition, DPP4 inhibition enhanced glycaemic regulation; moreover, it modifies cholesterol synthesis, lipoproteins [9] and liver function enzymes (ALT, AST, ALP). Hence, all the previous mechanisms lead to amelioration of hepatic histopathological features in clinical trials of patients with type 2 diabetes [33].

Previous studies have shown that rutin has a significant outcome on blood glucose control, and also has a very significant effect on the safety of liver cells [1, 6, 7]. DM-related hepatic cellular damage is strictly interrelated to free-radical burden; therefore, rutin's antioxidant activity can shield the liver cells [15, 34].

CONCLUSION AND RECOMMENDATIONS

Concomitant administration of sitagliptin and rutin has an outstanding ameliorating role on diabetes-induced hepatic histopathological and biochemical alterations. This is better than either rutin or sitagliptin alone. Therefore, the use of both sitagliptin and rutin give outstanding results in liver protection against diabetic changes. It is recommended to use rutin on higher doses to test its effect on diabetes-induced and other hepatic injuries.

Conflict of interest: None declared

REFERENCES

- Ahmed OM, Moneim AA, Yazid IA, et al. Antihyperglycemic, antihyperlipidemic and antioxidant effects and the probable mechanisms of action of Ruta graveolens infusion and rutin in nicotinamide-streptozotocin-induced diabetic rats. Diabetol Croat. 2010; 39(1): 15–35.
- Ahrén B. Clinical results of treating type 2 diabetic patients with sitagliptin, vildagliptin or saxagliptin: diabetes control and potential adverse events. Best Pract Res Clin Endocrinol Metab. 2009; 23(4): 487–498, doi: 10.1016/j. beem.2009.03.003, indexed in Pubmed: 19748066.
- Amori RE, Lau J, Pittas AG. Efficacy and safety of incretin therapy in type 2 diabetes: systematic review and meta-analysis. JAMA. 2007; 298(2): 194–206, doi: 10.1001/ jama.298.2.194, indexed in Pubmed: 17622601.

- Aschner P, Kipnes MS, Lunceford JK, et al. Sitagliptin Study 021 Group. Effect of the dipeptidyl peptidase-4 inhibitor sitagliptin as monotherapy on glycemic control in patients with type 2 diabetes. Diabetes Care. 2006; 29(12): 2632–2637, doi: 10.2337/dc06-0703, indexed in Pubmed: 17130196.
- Bancroft JD, Gamble M. Theory and Practice of Histological Techniques. 7th ed. Staining methods. Churchill Livingstone,, Edinburgh, London, Madrid, Melbourne, New York and Tokyo 2008.
- Bertolani C, Marra F. The role of adipokines in liver fibrosis. Pathophysiology. 2008; 15(2): 91–101, doi: 10.1016/j. pathophys.2008.05.001, indexed in Pubmed: 18602801.
- Geerts A. History, heterogeneity, developmental biology, and functions of quiescent hepatic stellate cells. Semin Liver Dis. 2001; 21(3): 311–335, doi: 10.1055/s-2001-17550, indexed in Pubmed: 11586463.
- Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res. 2010; 107(9): 1058–1070, doi: 10.1161/ CIRCRESAHA.110.223545, indexed in Pubmed: 21030723.
- Giampietro O, Giampietro C, Bartola LD, et al. Sitagliptin as add-on therapy in insulin deficiency: biomarkers of therapeutic efficacy respond differently in type 1 and type 2 diabetes. Drug Des Devel Ther. 2013; 7: 99–104, doi: 10.2147/DDDT.S38346, indexed in Pubmed: 23439744.
- Haidara MA, Ibrahim MI, Sit EI, et al. Effect of a-tocopherol on glucose uptake and contractility in rat skeletal muscles. Med Sci Monv. 2003; 9(5): 214–217.
- Horn T, Junge J, Christoffersen P. Early alcoholic liver injury: changes of the Disse space in acinar zone 3. Liver. 1985; 5(6): 301–310, doi: 10.1111/j.1600-0676.1985.tb00253.x, indexed in Pubmed: 4088004.
- Hramiak IM, Finegoodk DT, Adams PC. Factors affecting glucose tolerance in hereditary hemochromatosis. Clin Invest Med. 1997; 20: 110–118.
- Hsu CY, Shih HY, Chia YC, et al. Rutin potentiates insulin receptor kinase to enhance insulin-dependent glucose transporter 4 translocation. Mol Nutr Food Res. 2014; 58(6): 1168–1176, doi: 10.1002/mnfr.201300691, indexed in Pubmed: 24668568.
- Jeschke MG. The hepatic response to thermal injury: is the liver important for postburn outcomes? Mol Med. 2009; 15(9-10): 337–351, doi: 10.2119/molmed.2009.00005, indexed in Pubmed: 19603107.
- Júnior II, Barbosa Hd, Carvalho DCR, et al. Brazilian attenuated hyperglycemia, dyslipidemia, and prooxidant status in alloxan-induced diabetic rats. Scientific World J. 2017; 2017: 5275813, doi: 10.1155/2017/5275813, indexed in Pubmed: 28567440.
- Liu Q, Pan R, Ding L, et al. Rutin exhibits hepatoprotective effects in a mouse model of non-alcoholic fatty liver disease by reducing hepatic lipid levels and mitigating lipid-induced oxidative injuries. Int Immunopharmacol. 2017; 49: 132–141, doi: 10.1016/j.intimp.2017.05.026, indexed in Pubmed: 28577437.
- Manaras K, Jongdee N, Uraporn V, et al. Effect of glabridin on collagen deposition in liver and amelioration of hepatocyte destruction in diabetes rats. Exp Ther Med. 2019; 18(2): 1164–1174, doi: 10.3892/etm.2019.7664, indexed in Pubmed: 31316610.

- Maritim AC, Sanders RA, Watkins JB. Diabetes, oxidative stress, and antioxidants: a review. J Biochem Mol Toxicol. 2003; 17(1): 24–38, doi: 10.1002/jbt.10058, indexed in Pubmed: 12616644.
- Masarone M, Rosato V, Dallio M, et al. Role of oxidative stress in pathophysiology of nonalcoholic fatty liver disease. Oxid Med Cell Longev. 2018; 2018: 9547613, doi: 10.1155/2018/9547613, indexed in Pubmed: 29991976.
- 20. Meng XM, Chung ACK, Lan HY. Role of the TGF-β/BMP-7/ Smad pathways in renal diseases. Clin Sci (Lond). 2013; 124(4): 243–254, doi: 10.1042/CS20120252, indexed in Pubmed: 23126427.
- Moustafa Hassan I, El-Gharabawy G, Moustafa AG. The effect of Sitagliptin (Januvia) on the liver of adult Albino rats in cases of experimental diabetes mellitus(Microscopic and laboratory studies). Egyptian J Hosp Med. 2012; 47(1): 260–278, doi: 10.21608/ejhm.2012.16296.
- Nakashima O, Kurogi M, Yamaguchi R, et al. Unique hypervascular nodules in alcoholic liver cirrhosis: identical to focal nodular hyperplasia-like nodules? J Hepatol. 2004; 41(6): 992–998, doi: 10.1016/j.jhep.2004.08.014, indexed in Pubmed: 15582133.
- Newmeyer DD, Ferguson-Miller S. Mitochondria: releasing power for life and unleashing the machineries of death. Cell. 2003; 112(4): 481–490, doi: 10.1016/s0092-8674(03)00116-8, indexed in Pubmed: 12600312.
- Pan PH, Lin SY, Wang YY, et al. Protective effects of rutin on liver injury induced by biliary obstruction in rats. Free Radic Biol Med. 2014; 73: 106–116, doi: 10.1016/j.freeradbiomed.2014.05.001, indexed in Pubmed: 24815012.
- Rahimi R, Nikfar S, Larijani B, et al. A review on the role of antioxidants in the management of diabetes and its complications. Biomed Pharmacother. 2005; 59(7): 365–373, doi: 10.1016/j. biopha.2005.07.002, indexed in Pubmed: 16081237.
- Rother KI. Diabetes treatment bridging the divide. N Engl J Med. 2007; 356(15): 1499–1501, doi: 10.1056/ NEJMp078030, indexed in Pubmed: 17429082.

- Soares JM, Pereira Leal AE, Silva JC, et al. Influence of flavonoids on mechanism of modulation of insulin secretion. Pharmacogn Mag. 2017; 13(52): 639–646, doi: 10.4103/ pm.pm 87 17, indexed in Pubmed: 29200726.
- Tian S, Bilin Xu, Taolei C, et al. Sitagliptin reduces insulin resistance and improves rat liver steatosis via the SIRT1/ /AMPKα pathway. Exp Ther Med. 2018; 16(4): 3121–3128, doi: 10.3892/etm.2018.6554, indexed in Pubmed: 30214535.
- Uchida K. Future of toxicology: lipid peroxidation in the future: from biomarker to etiology. Chem Res Toxicol. 2007; 20(1): 3–5, doi: 10.1021/tx600304n, indexed in Pubmed: 17226920.
- Vanlangenakker N, Vanden Berghe T, Krysko DV, et al. Molecular mechanisms and pathophysiology of necrotic cell death. Curr Mol Med. 2008; 8(3): 207–220, doi: 10.2174/156652408784221306, indexed in Pubmed: 18473820.
- Wada J, Makino H. Inflammation and the pathogenesis of diabetic nephropathy. Clin Sci (Lond). 2013; 124(3): 139–152, doi: 10.1042/CS20120198, indexed in Pubmed: 23075333.
- 32. Yilmaz Y, Senates E, Yesil A, et al. Not only type 2 diabetes but also prediabetes is associated with portal inflammation and fibrosis in patients with non-alcoholic fatty liver disease. J Diabetes Complications. 2014; 28(3): 328–331, doi: 10.1016/j.jdiacomp.2014.01.013, indexed in Pubmed: 24602757.
- Yoo H, Ku SK, Baek YD, et al. Anti-inflammatory effects of rutin on HMGB1-induced inflammatory responses in vitro and in vivo. Inflamm Res. 2014; 63(3): 197–206, doi: 10.1007/ s00011-013-0689-x, indexed in Pubmed: 24292859.
- 34. Zargar S, Wani TA, Alamro AA, et al. Amelioration of thioacetamide-induced liver toxicity in Wistar rats by rutin. Int J Immunopathol Pharmacol. 2017; 30(3): 207–214, doi: 10.1177/0394632017714175, indexed in Pubmed: 28590141.