Biochemical and histological study on the effect of levetiracetam on the liver and kidney of pregnant albino rats

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Running Head: Assessment of Levetiracetam on pregnant rat

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Abstract

Background: Levetiracetam is a broad-spectrum antiseizure agent and one of the most commonly prescribed drugs for epilepsy. The aim of this work was assess the effect of levetiracetam at its therapeutic range on the liver and kidney of pregnant albino rats.

Material and methods: Forty pregnant rats were divided equally into two groups (I–II), Rats in the group I were gavaged 1.5 ml/day distilled water in two divided doses throughout pregnancy. Rats in the group II were gavaged 1.5 ml/day distilled water (containing 36 mg levetiracetam) in two divided doses throughout pregnancy. At the end of the experiment, blood samples were taken and the sera were separated and used for biochemical analysis. The Kidneys and livers of both groups were excised and used for light and electron microscopic examination.

Results: Treatment with levetiracetam induced undesirable histopathological changes in the liver and kidney of pregnant albino rats. These changes were in the form of distortion of the hepatic architecture, dilatation of the central and the portal veins, widening of the Bowman’s spaces, thickening and disruption of the glomerular basement membrane, fusion and effacement of secondary foot processes, cytoplasmic vacuolation, and swollen...
mitochondria with loss of their cristae. Such changes were confirmed by alteration of certain biochemical parameters related to the liver and kidney functions.

**Conclusions:** Authors concluded that levetiracetam induced deleterious effects on the liver and kidney of pregnant albino rats. Further investigations is recommended to clarify the mechanism of levetiracetam toxicity.

**Key words:** levetiracetam, liver, kidney, pregnant rats

**INTRODUCTION**

Epilepsy is a common sustained disorder affecting approximately more than one million women of childbearing age (9). The prevalence of epilepsy has been estimated to be 0.3–0.5% in pregnant women (33). Pregnant women with epilepsy are advised to maintain antiepileptic drugs (AED) to diminish maternal and fetal disturbance joined with seizures (22).

Pharmacological treatment options of epilepsy comprise conventional AED (valproate, phenobarbital and carbamazepine), in addition to the newer second generation agents (levetiracetam, gabapentin and topiramate) (15). Conventional AED had good therapeutic effects and low treatment cost. However, some undesired effects associated with these drugs, such as memory deterioration and Stevens–Johnson syndrome (37). The effectiveness of the second generation drugs, seems to be similar to that of the conventional AED. However, the newer drugs such as levetiracetam, seem to offer a lower risk of interactions with other medications, simpler titration, and administration regimens, and improved tolerability (15).

Levetiracetam was developed as an analogue of piracetam, a drug used to improve cognitive function (24). Levetiracetam is recognized for adjunct therapy of focal onset, primary generalized tonic–clonic, and myoclonic seizures in children and adults (25). The correlation between binding affinity of levetiracetam analogs and their potency toward audiogenic seizures suggests that a synaptic vesicle protein, SV2A, mediates the antiseizure effects of levetiracetam (26). Oral absorption of levetiracetam is rapid and nearly complete, with peak plasma concentrations in 1.3 hours. Two-thirds of the drug is
excreted unchanged in the urine and the remainder as 2-pyrrolidone-N-butyric acid (inactive deaminated metabolite). There is no metabolism of levetiracetam in the liver as its metabolism takes place in the blood. (27).

Some authors reported a significant increase in gamma glutamyl transferase in an epileptic patient on levetiracetam monotherapy (5). Study reported some adverse effects associated with levetiracetam therapy such as sleepiness, headaches, vomiting, inflammation of the nose and throat, and irritability (28).

Data on the safety of levetiracetam during pregnancy is still limited (3). So, this work aimed to assess the effect of levetiracetam on the liver and kidney of pregnant albino rats.

MATERIALS AND METHODS

Drug

Levetiracetam (Tiratam) was purchased from Al-Andalous for Pharmaceuticals-Egypt Industries. It was available in the form of tablets 500 mg each. Adult dosing can begin with 1000 mg twice daily, increased to a maximum dosage of 3000 mg twice daily (23). The chosen dose was 2000mg/day. The equivalent dose for adult rat weighting about 200 gm is 36 mg/day According to (20).

Animals

Sixty adult albino rats (twenty males and forty females) of local strain weighing about 200 g were obtained for this study. Care of animals and practices were provided with the Animal House, Faculty of Medicine (Girls), Al-Azhar University, Cairo, according to the “Guide of the Care and Use of Laboratory Animals (11). Each male rat was located with 2 virgin female rats in a cage for 24 hours. In the morning, the first day of pregnancy was determined by the presence of the vaginal plug (16). Each pregnant female was placed in an individual cage until the end of study.

Experimental design

The forty pregnant rats were divided equally into two groups:
— **Group I (control)**: each pregnant rat was gavaged 1.5 ml/day distilled water in two divided doses throughout pregnancy.

— **Group II (treated)**: each pregnant rat was gavaged 1.5 ml/day distilled water (containing 36 mg levetiracetam) in two divided doses throughout pregnancy.

At the end of the experiment, blood samples were taken on the first day after delivery from the retro-orbital vein of all rats after ether inhalation (36). The sera were separated by centrifugation of blood at 3000 rpm for 15 minutes, then preserved at −20°C until used for biochemical analysis. Rats were sacrificed by cervical dislocation to avoid any chemical injury (6), their abdominal cavities were exposed by a midline incision, and the Kidneys and livers were excised and used for light and electron microscopic examination.

**Histopathological preparation**

**Preparation for light microscopic examination**

Pieces of liver and kidney tissues from the control and treated groups were washed and fixed for three days in Bouin’s solution. Later, the tissues were dehydrated in ascending graded series of ethyl alcohol and processed for paraffin wax embedding. Liver and Kidney tissues blocks were cut using microtome into serial sections at a thickness of 7μm. The sections were placed on glass slides and stained with hematoxylin and eosin, Masson’s trichrome for detecting collagen fibers and PAS (periodic acid Schiff’s reaction) for verifying the contents of carbohydrates(2). For histopathological examination of liver and kidney tissue a microscope (LEICA DM 750) and digital camera were used.

**Preparation for Electron Microscopic examination**

Small slices of the liver and kidney tissues were fixed quickly in 2.5% phosphate-buffered glutaraldehyde (pH 7.4). The specimens were then post-fixed with 1% osmium tetroxide in the same buffer at 41C, dehydrated in graded ethanol and embedded in epoxy resin (Bancroft and Gamble, 2008). Using LKB ultra-microtome, semithin sections (0.5μm thick) were cut and stained with toluidine blue for light microscopic examination. Ultrathin sections were cut (80 nm), stained with uranyl acetate and lead citrate (10) and examined
and photographed using electron microscope (JEOL1010 EX II, Japan) at the Regional Mycology and Biotechnology Center, Al-Azhar University, Cairo, Egypt.

Biochemical indices

The level of some parameters of liver and kidney functions were measured using (BioSystems- BTS 310; Barcelona, Spain) which was calibrated for probes from rats.

Liver function indices

Alanine aminotransferase (ALT) and aspartate aminotransferase (AST) enzymes levels were measured in IU/L using the methods of (32).

Kidney function indices

Creatinine level in mg/dl was determined using the method of (8). Blood urea nitrogen measurement in mg/dl was assessed according to (18).

Statistical analysis

Statistical analysis of the levels of ALT, AST, urea and creatinine in the studied groups were carried out using the SPSS statistical package, the data were analyzed and statistical significance was determined by using unpaired Student t- test (21).

RESULTS

Histopathological results

Liver

Light microscopy

Examination of H - E stained liver transverse sections of group I (control group) revealed that the liver was built up of cords of hepatocytes emitting from the central vein and spaced with blood sinusoids (Figs. 1A and 1B). The portal tract was built up of a branches of portal vein, bile duct and lymph vessel (Fig. 1B). Masson's trichrome staining showed minimal collagen fibers around the central vein (Fig. 2A) and the portal tract (Fig.
Periodic Acid Schiff reaction revealed deposition of PAS positive material in the hepatocytes (Fig. 3A).

Inspection of H - E stained liver transverse sections of group II (treated group) revealed that the hepatic cords did not have the normal radial arrangement, and the central vein was dilated (Fig. 1D). Most hepatocytes had cytoplasmic vacuoles, and some of their nuclei were small, irregular and deeply stained (Figs. 1D and 1E). The portal area showed a markedly dilated portal vein (Fig. 1E). Masson's trichrome staining showed a markedly higher level of collagen-fiber distribution around the central vein (Fig. 2D) and the elements of the portal tract as compared with sections from control group (Fig.2E). Periodic Acid Schiff reaction showed apparent reduction of the distribution of PAS positive material in the hepatocytes (Fig. 3C).

Electron microscopy

Examination of ultrathin sections of the liver of the group I (control group) revealed that the hepatocytes had intact plasma membranes and nuclei with demarcated nuclear envelopes. The nuclei contained prominent nucleoli and both euchromatin and heterochromatin. Their cytoplasm contained rounded or oval mitochondria, free ribosome, rough endoplasmic reticulum and glycogen granules (Fig.4A).

Examination of ultrathin sections of the liver of the group II (treated group) showed histopathological changes in the hepatocytes. Some nuclei had irregular nuclear envelopes and absent nucleoli. The cytoplasm contained vacuolated areas, few glycogen granules, swollen mitochondria with loss of the cristae and matrix, rough endoplasmic reticulum and ribosomes (Fig. 4B).

Kidney

Light microscopy

Examination of H - E stained kidney sections of group I (control group) showed the renal cortex was built up of Malpighian corpuscles and tubules (proximal convoluted tubules (PCT), distal convoluted tubules (DCT) and collecting tubules). The Malpighian corpuscles was formed of glomerulus surrounded by Bowman’s capsule (Fig.1C). The
PCT and DCT were lined by cuboidal cells. The PCT had narrow Lumina with brush border on their luminal surfaces; however, the DCT exhibited more number of lining cells, and wide Lumina with no brush border. The collecting tubules were lined by 10-20 cuboidal epithelial cells (Fig.1C).

Masson’s trichrome stained sections revealed normal distribution of collagen fibers in some elements of renal cortex (Fig.2C). PAS-stained sections revealed a PAS-positive reaction in some elements of renal cortex (Fig.3B).

Examination of H - E stained kidney transverse sections of group II showed renal corpuscles containing some shrunken glomeruli with wide capsular spaces. Some of the glomerular tuft nuclei were small, irregular and deeply stained. Some of the epithelial cells lining the renal tubules (PCT, DCT and collecting tubules) showed cytoplasmic vacuolization, intraluminal exfoliation as well as small darkly stained irregular nuclei. The brush border of the some PCT became disrupted (Fig.1F). Masson’s trichrome stained sections showed an apparent increase in the collagen fibers deposition in the parietal layer of Bowman's capsule, around the blood vessels, and in the basal lamina of some renal tubules (Fig.2F). PAS-stained sections revealed a weak reaction in the parietal layer of Bowman’s capsule, in the basal lamina of the renal tubules and in the brush border of the PCT (Fig.3D).

Electron microscopy

Examination of ultrathin sections of the renal cortex of the group I (control group) showed that the renal corpuscles was formed of glomerular capillaries lined by fenestrated endothelium and wrapped by podocytes. The podocytes had large irregular euchromatic nuclei, primary processes and secondary processes. Filtration slits were located between the secondary processes. The glomerular basement membrane appeared as a trilaminar structure (Fig.5A). The epithelial cells lining the PCT were rested on a regular basal lamina that displayed basal infoldings ran perpendicular to it. Numerous rod like mitochondria were lodged between the basal infoldings and a noticeable number of mitochondria of different sizes and shapes were situated on the upper portion of the cells (Fig.5B). In addition to mitochondria, the cytoplasm harbors endoplasmic reticulum, ribosomes, Golgi
complex and lysosomes. The apex of the epithelial cells had tall, closely packed projections called microvilli. The nuclei of these epithelial cells were large, euchromatic with well demarcated nuclear membrane. The nuclei contained clumps of heterochromatin and nucleoli. (Fig. 5B). The epithelial cells lining the DCT were resting on a regular basement membrane that exhibited basal infoldings ran perpendicular to it. Numerous elongated mitochondria were lodged between the basal infoldings (Fig. 5C).

Examination of ultrathin sections of the renal cortex of group II showed histopathological changes. The renal corpuscle revealed thickening of the glomerular basement membrane and loss of its trilaminar structure, some of the secondary foot processes were effaced or fused (Fig. 5D). The epithelial cells lining some PCT had small nuclei with increased heterochromatin contents and irregular ill demarcated nuclear envelope. Their mitochondria were longitudinally arranged with loss of cristae or swollen. Their cytoplasm exhibited primary lysosomes, and their microvilli were less abundant (Figs. 5E). The epithelial cells lining some DCT showed small nuclei with increased heterochromatin. Their cytoplasm exhibited rarefaction, loss of basal infoldings, disorganized mitochondria with loss of their cristae (Fig. 5F).

**Biochemical results**

*Liver indices*

Statistical analysis of the mean levels of ALT and AST in groups I and II revealed significant (P < 0.05) increase in group II as compared with group I (Table 1).

*Kidney indices*

Statistical analysis of the mean levels of Urea and creatinine in groups I and II revealed significant (P < 0.05) increase in group II as compared with group I (Table 1).

**DISCUSSION**

The results of the current study revealed that treatment of pregnant albino rats with levetiracetam induced degenerative changes in their livers, specifically in the form of distortion of the hepatic architecture. Levetiracetam treatment also led to dilatation of the
central and the portal veins, cytoplasmic vacuolation and swollen mitochondria with loss of their cristae, such histopathological changes were confirmed by elevation of liver enzymes.

These results are in line with (31) who reported an episode of acute liver injury immediately following the treatment with levetiracetam in a patient with traumatic brain injury and restoration of normal liver function following stoppage of levetiracetam. Some studies (35, 2) reported elevated serum alkaline phosphatase (ALP) level after levetiracetam treatment in children with epilepsy. Some authors observed that the serum ALP level returned to normal after the cessation of levetiracetam administration and concluded that the liver was the main cause of enzyme elevation after exclusion of other causes (35). Some authors reported an elevation of the liver enzymes in a patient following levetiracetam administration for seizure prophylaxis. The enzymes returned to the normal level after levetiracetam discontinuation (27).

The current study showed that treatment of pregnant albino rats with levetiracetam induced nuclear changes in their liver cells. Such results were confirmed by a study (7) whose authors evaluated the genotoxic effect of levetiracetam (at doses 300 and 600 mg/kg b. w) in the liver and brain tissues of treated pregnant rats and their fetuses during pregnancy. They reported a significant increase of DNA damage as detected by alkaline Comet assay, and an appearance of both apoptotic laddered and smeared DNA in the tissues tested.

On the other hand, the results of the present study were contrary to study whose authors evaluated the levetiracetam tolerability in patients (fourteen) with liver disease and epilepsy. After 6 months of supervision, they reported that none of the patients showed decline of liver function on the base of blood analysis (4). Based on these observations, they assumed that levetiracetam was an agreeable treatment option in epileptic cases with liver impairment. The results of the present study were also contrary to some authors (17) who reported that chronic administration of levetiracetam to rats at a therapeutic dose (54mg/day) revealed no histopathological differences in their liver as compared to those of control. Some authors (34) mentioned that levetiracetam did not interact with the cytochrome P450 system and was the best drug of choice for cases with liver impairment.
In the current study, Masson’s trichrome staining showed higher level of collagen-fiber distribution around the central vein and the elements of the portal tract in levetiracetam treated group as compared with sections from the control group. Some authors mentioned that excessive collagen deposition in the liver was mediated by death of hepatocytes and activation of hepatic stellate cells. They added that lipid peroxidation including the generation of reactive oxygen species, could be implicated as a cause of excessive collagen deposition (29).

The current study revealed that treatment of pregnant albino rats with levetiracetam induced degenerative changes in their renal cortex. The changes were in the form of widening of the Bowman’s spaces, pyknosis of some of the glomerular tuft nuclei, thickening and disruption of the glomerular basement membrane, and fusion and effacement of secondary foot processes. Levetiracetam treatment also led to cytoplasmic vacuolization, intraluminal exfoliation, and disorganized mitochondria in the cells lining the renal tubules. In addition, Levetiracetam treatment led to an increased heterochromatin contents of some nuclei of the cells lining the renal tubules, such pathological changes were confirmed by significant elevation of the levels of urea and creatinine.

These findings were in line with a study (14) whose authors presented occurrence of acute kidney injury in Drug Reaction with Eosinophilia and Systemic Symptoms (DRESS) two months after starting levetiracetam to a 75-year-old man with glioblastoma. Also, the results of the present study were in line with some researchers (30) who reported occurrence of an acute kidney injury 1 day after the initiation of levetiracetam treatment for new-onset seizures in a healthy 23-year-old female patient. Based on the time course of the patient’s elevation in the level of serum creatinine and the exclusion of other causes, the authors suggested that levetiracetam treatment was contributed to the acute kidney injury.

On the other hand, the results of the current study were contrary to some researchers (13) who evaluated the clinical use of intravenous levetiracetam in patients with renal impairment and didn’t detect any major side effects any patients. So, they concluded that intravenous levetiracetam treatment was effective and safe in patients with renal impairment. The results of the present study also were contrary to a study (19) whose
authors presented two pediatric cases of levetiracetam overdose for one month (115, 300 mg/kg/day) and didn't observe remarkable severe side effects in their patients. So, they suggested that accidental treatment with an overdose of levetiracetam in children for long period was rarely accompanied with serious side effects.

In the current work, Masson's trichrome staining showed higher level of collagen-fiber distribution in the parietal layer of Bowman's capsule, and in the wall of some renal tubules in levetiracetam treated group as compared with sections from the control group. Such results may be caused by disturbance in oxidant/antioxidant system. Such suggestion was supported by some authors who investigated the association of reactive oxygen species/antioxidant systems with kidney fibrosis following Ischemic/Reperfusion injury (12). They concluded that ROS/oxidative stress was contributed to the progression of kidney fibrosis (12).

In the current study, the degenerative changes that were observed in the liver and kidney might be due to disturbance in the oxidant/antioxidant ratio. Such suggestion was supported by researchers (7) who explained that genotoxic potential of levetiracetam by decreased free radical neutralization and/or an increased production of free radicals.

CONCLUSIONS

Authors concluded that levetiracetam induced deleterious effects on the liver and kidney of pregnant albino rats. Continuous assessment of the liver and kidney functions during levetiracetam therapy is advised. In addition, further investigations is recommended to clarify the mechanism of levetiracetam toxicity.

Declaration of conflicts of interest: No conflicts of interest

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References


**Table 1.** Means and standard deviations of ALT, AST, Urea and creatinine of pregnant rats in groups’ I – II.

<table>
<thead>
<tr>
<th>Measurements</th>
<th>Pregnant rats at 1-day after delivery</th>
<th>T test</th>
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<tr>
<td></td>
<td>Group I (n = 20)</td>
<td>Group II (n = 20)</td>
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<tr>
<td>ALT (IU/ L)</td>
<td>Mean ± SD Range 17.2 ± 2.69 13–22</td>
<td>35.9 ± 4.817791 31–43</td>
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<td>AST (IU/ L)</td>
<td>Mean ± SD Range 12.6 ± 1.349897 11–14</td>
<td>26.9 ± 2.960856 24–31</td>
<td>2.160369</td>
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<td>Urea (mg/dl)</td>
<td>Mean ± SD Range 37.39 ± 1.066094 36–39</td>
<td>49.55 ± 1.480428 47–51</td>
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</tr>
<tr>
<td>Creatinine (mg/dl)</td>
<td>Mean ± SD Range 0.663 ± 0.021108 0.64–0.69</td>
<td>0.791 ± 0.008756 0.78–0.8</td>
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</tr>
</tbody>
</table>
**Figure 1.** Photomicrographs of liver (A, B, D, E) and renal cortex(C, F) transverse sections (H&E ×400 magnification):

1A: group I shows hepatocytes (H), central vein (CV), blood sinusoids(S) flat endothelial cells (E) and von Kupffer cells (K).

1B: group I shows a branch of portal vein (PV), bile duct (BD) and lymph vessel (L). The hepatocytes (H), vesicular basophilic nuclei (N).

1C: group I shows the glomerulus (G), Bowman’s space (S), the Partial (pl) and visceral (vl) layers of Bowman’s capsule, PCT (P), DCT (D) and collecting tubules (CT).

1D: group II shows dilated central vein (CV), vacuolated cytoplasm (V) of hepatocytes, irregular deeply stained nuclei (N*), congested blood sinusoids (S).

1E: group II shows dilated portal vein (PV), vacuolated cytoplasm (V) of hepatocytes, irregular deeply stained nuclei (N*), hepatic artery (HA) and bile duct (BD).

1F: group II shows shrunken glomeruli (G), wide Bowman's space (BS), deeply stained glomerular cell nuclei (arrow), the lining epithelium of the PCT (P), DCT (D) and collecting tubules (CT), have vacuolated cytoplasm (V) and small deeply stained nuclei (arrow). The Lumina of some renal tubules contain cellular exfoliation (ex).

**Figure 2.** Photomicrographs of liver (A, B, D, E) and renal cortex(C, F) transverse sections (Masson Trichrome ×400 magnification):

2A: group I shows normal distribution of collagen fibers (CF) around the central vein (CV).

2B: group I shows normal distribution of collagen fibers (CF) around the portal tract (PT) and between the hepatocytes.

2C: group I shows the collagen fibers (cf) in the parietal layer (pl) of the Bowman’s capsule, glomerular capillaries (G) and in the basement membranes (BM) of renal tubules (T).

2D: group II shows a higher level of collagen fibers (CF) distribution is seen around the central vein (CV).
2E: group II shows a higher level of collagen fiber (CF) distribution is seen around the elements of the portal tract (PT).

2F: group II shows increase in the collagen fibers (cf) in the parietal layer (pl) of the Bowman’s capsule surrounding the glomerulus (G), in the basement membranes (BM) of renal tubules (T) and around the blood vessel (BV).

Figure 3. Photomicrographs of liver of liver (A, C) and renal cortex (B, D) transverse sections (PAS ×400 magnification)

3A: group I shows the normal distribution of PAS positive material in the hepatocytes.

3B: group I shows the PAS positive reaction in the parietal layer (pl) of Bowman’s capsule around the glomerulus (G), in the basement membranes (BM) of renal tubules and in the brush border (B) of the PCT (P)

3C: group II shows reduction in the distribution of PAS positive material in the hepatocytes.

3D: group II shows decrease the PAS positive reaction in the parietal layer (pl) of Bowman’s capsule around the glomerulus (G), in the basement membranes (BM) of renal tubules and in the brush border (B) of the PCT (P).

Figure 4. Photomicrographs of Transmission electron microscopy of liver sections (TEM ×10,000 magnification).

4A: groups I shows the cytoplasm of hepatocyte contains mitochondria (M), ribosomes (r), and glycogen granules (g). Notice the nucleus (N), nuclear envelope (ne), nuclear pores (arrow), heterochromatin (h), euchromatin (e) and prominent nucleolus (nu).

4B: groups II shows the nuclei of hepatocytes (N) have irregular nuclear envelopes (ne*) and absent nucleoli. The cytoplasm contains vacuolated areas (V), mitochondria with a loss of cristae (M), ribosomes.
**Figure 5.** Electron micrograph of the ultrathin renal cortex section (TEM ×10,000 magnification).

5A: group I shows a part of renal corpuscle. The glomerular capillary (C), fenestrated endothelium (F), the trilaminar glomerular basement membrane (BM), podocytes (PO) with their nuclei (N), primary cytoplasmic process (PP) and secondary processes (SP).

5B: group I shows a part of cell lining the PCT, it has euchromatic nucleus (N), well demarcated nuclear envelope (ne), containing nuclear pores (NP), numerous apical microvilli (MV), basal infoldings (arrow), elongated mitochondria (M), ribosomes (r) and rough endoplasmic reticulum (R).

5C: group I showing a part of cell lining the DCT, it has oval nucleus (N), well demarcated nuclear envelope (ne), nucleolus (Nu), basal infoldings (arrow), basement membrane (BM) and longitudinal arranged mitochondria (M).

5D: group II shows a part of renal corpuscle, the podocyte (Po), primary processes (PP), distorted and flat secondary foot processes (SP). The basement membrane (BM) lining the glomerular capillaries (C) is thickened and its trilaminar structure is lost.

5E: group II showing a part of cell lining the PCT, it has small nucleus (N) with increased heterochromatin, ill demarcated irregular nuclear envelope (ne), less abundant apical microvilli (MV) and elongated mitochondria (M) with loss of their cristae.

5F: group II showing a part of cell lining the DCT, it has small nucleus (N) with increased heterochromatin content and irregular ill-demarcated nuclear envelope (ne), loss of basal infoldings, disorganized mitochondria (M) with loss of their cristae, and rarefied cytoplasm (R).