

# Localisation of NADPH-diaphorase-positive structures in the thymus of the rat, mouse and rabbit

Emil Švický<sup>1</sup>, Miloslav Ondrašovič<sup>1</sup>, Ján Danko<sup>1</sup>, Olga Ondrašovičová<sup>1</sup>, Andrej Jenča<sup>2</sup>, Norbert Pospieszny<sup>3</sup>, Michal Toropila<sup>1</sup>

<sup>1</sup>University of Veterinary Medicine of Košice, Slovak Republic

<sup>2</sup>Faculty of Medicine UPJŠ of Košice, Slovak Republic

<sup>3</sup>Department of Veterinary Anatomy and Histology, Agricultural Academy of Wrocław, Poland

[Received 6 January 2003; Revised 20 May; Accepted 20 May 2003]

*Localisation of the diaphorase activity of nicotinamide adenine dinucleotide phosphate (NADPH-d), acting as a marker of nitric oxide synthesis (NOS), was studied in the thymus of the rat, mouse and rabbit. The NADPH-d-active cells observed in the rat thymus were irregular in shape with numerous projections and were located on the boundary between the cortex and the medulla. The NADPH-d-active cells in the thymus of the mouse were located predominantly in the medulla. They varied in coloration and their shape was oval, round, or irregular. NADPH-d-positive nerve fibres were located perivascularly. The rabbit thymus displayed a lightly stained cortex, whereas the medulla was seen as a rounded complex of intensively stained cells, without sharp demarcation between them. In the rat thymus, the NADPH-d-positive nerve fibres were not evident, whereas NADPH-d-positive nerve fibres were seen in the perivascular topography of the mouse and the rabbit thymus.*

*These results suggest that NO may participate in the regulation of the thymic function in the species. In summary, the present results reveal the distribution of NADPH-diaphorase-d-positive structures in the rat, mouse and rabbit thymus.*

**key words:** NADPH-d, thymus, rat, mouse, rabbit

## INTRODUCTION

The nicotinamide dinucleotide hydrogenphosphate diaphorase (NADPH-d) histochemical reaction consists in the transfer of hydrogen from the NADPH-substrate to tetrazolium salt which changes to insoluble deep-blue formazan [19, 21]. The importance of this reaction increased after the discovery that the similarity of neuronal NADPH-d and nitric oxide synthase (NOS) is such that the two were identical [8, 16]. Nitric oxide (NO) is synthesised from L-arginine by an enzyme nitric oxide synthase (NOS), which can exist in a number of isoforms one of them being the neuronal NOS. It has been found that the neuronal

NOS, present in the brain and peripheral organs, is identical to NADPH-d [16]. With regard to the increasing importance of NO in the organism, the histochemical method for NADPH-d also appears suitable for the study of NOS-possessing nerve structures. Numerous studies have demonstrated that both forms of NOS, the endothelial and the neuronal, are located, together with NADPH-d, in tissues fixed with 4% paraformaldehyde. An increasing amount of data indicates that NO mediates certain functions in the autonomous nervous system [4, 14].

The aim of the work was to study and compare the NADPH-d activity of cells in the thymuses of var-

ious species of rodents and to identify their shape differences and localisation in individual components of the parenchyma.

## MATERIAL AND METHODS

Experiments were carried out on 10 mice of both sexes, of 40–50 g body weight, 8 male (Wistar) rats weighing 250–300 g and 9 Chinchilla rabbits, body weight 2.5–3.0 kg of both sexes. Before experiments, the animals were reared in the Central Animal Husbandry quarters under veterinary care. The animals were anaesthetised by pentobarbital and killed by intracardial perfusion of saline solution and subsequent perfusion of 4% paraformaldehyde containing 0.1% glutaraldehyde in 0.1 M phosphate buffer of pH 7.4. The solutions were prepared immediately before perfusion. After the perfusion, the thymuses were removed surgically and stored in identical fixative solution for 3–4 hours. They were then transferred to 30% saccharose solution in the same phosphate buffer and stored overnight at 4°C. Sections of 43–45  $\mu$ m were prepared by means of a cryomicrotome.

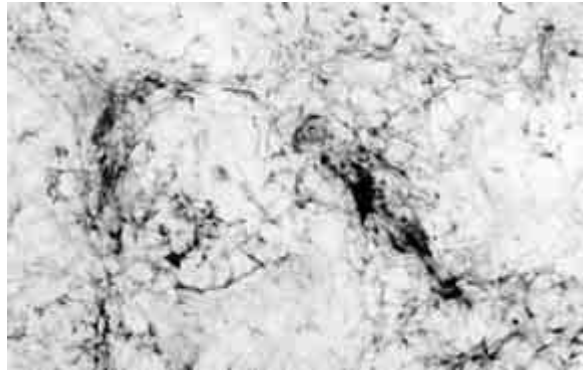
A histochemical reaction modified according to Scherer-Singler et al. [21] was used to visualise NADPH-d activity. The sections were incubated in a solution of 1.5 mM tetrazolium nitroblue (NBT, Sigma Chemicals, N-6876), 1.0 mM beta nicotinamide adenine dinucleotide phosphate (NADPH, Sigma Chemicals, N-1630), 10.0 monosodium maleate (maleic acid, Sigma Chemicals, N-1125) and 0.5 Triton X-100, dissolved in 0.1 M phosphate buffer, pH 8.0, at 37°C for hour. After the incubation, the sections were washed with 0.1 M phosphate buffer (pH 7.4), mounted in Entellan and evaluated under a light microscope.

## RESULTS

The NADPH-d histochemistry of the thymus of rodents showed a different distribution of diaphorase-positive reactivity in the medulla and the cortex, which appeared as deep blue stained cellular structures.

### Thymus of rats

The diaphorase-positive cells exhibited only slight intensity of coloration and diffuse distribution throughout the entire parenchyma of the cortex. The distribution of NADPH-d positive cells in the thymus medulla of rats is different. Intensively coloured cells were observed predominantly in the corticomedullary area where they occurred as irregular agglomer-



**Figure 1.** NADPH-d-positive cells on the border of the cortex and medulla in the rat thymus. They are of different shape and of intensive coloration. Primary magnification:  $\times 120$ .

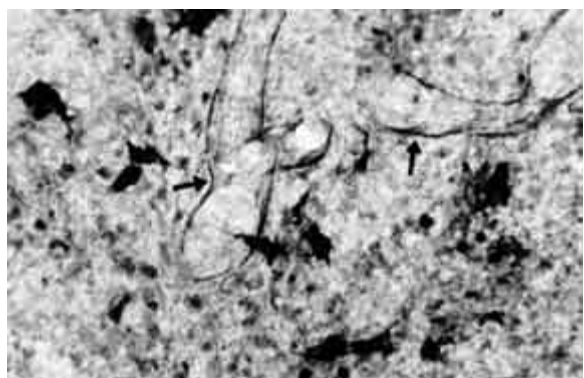
ates (Fig. 1). They were only sporadically dispersed in the inner cortex. In comparison with the cells of the thymus medulla in mice they exhibited a high degree of irregularity with numerous projections without NADPH-d-positive nerve fibres.

### Thymus of mice

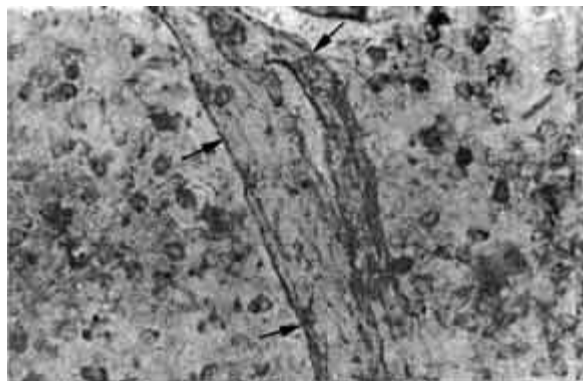
The NADPH-d-positive cortex cells occurred in low numbers and diffuse dispersion. Individual cells were oval and exhibited a low intensity of diaphorase-positive coloration.

No projections were observed in these cells. Examination of preparations are of better visibility only for radially oriented vessels.

The thymus medulla of mice contained more NADPH-d positive cells (Fig. 2). The shape of these cells was similar to that observed in the cortex. They were mostly oval, round and also irregular in shape. They varied in the intensity of their coloration. Medullary cells showed a marked intensity of NADPH-d activity. They were of irregular shape and dark colour.



**Figure 2.** NADPH-d-positive cells in the medulla of the mouse thymus. Fine NADPH-d-positive nerve fibres are observed in the perivascular area. Primary magnification:  $\times 120$ .



**Figure 3.** Detailed view of NADPH-d-positive cells of the rabbit. NADPH-d-positive nerve fibres arranged in perivascular topography. Primary magnification:  $\times 120$ .

The cells had short projections and were frequently located close to the vessels. Blood vessels located in the medulla and septa showed signs of NADPH-d-positivity of the endothelial cells. NADPH-d-positive nerve fibres were observed in perivascular localisation.

### Thymus of rabbits

The NADPH-d-positive cells were present in the cortex and the medulla of the thymus, but there were more of these cells in the medulla. The cells of the cortex were rounded or irregular in shape and were only slightly labelled, solitary and scattered. The medulla was characterised by the presence of a large quantity of cells with varying density of staining (Fig. 3). In general, they were similar in form and the oval, rounded, monopolar type of cell bodies in particular were observed to lack any apparent processes. Blood vessels localised in the septa showed signs of NADPH-d-positivity of the endothelial cells. NADPH-d-positive fibres were observed in perivascular localisation.

## DISCUSSION

In our study we described the NADPH-d-positive structures in rodent thymuses. The NADPH-d activity could be regarded as a marker of NOS, the enzyme responsible for the synthesis of NO [5, 8, 22]. The proof of activity of this enzyme in the nervous system of mammals was presented by the distribution of NADPH-d-positive structures described in the thymus of rats and chickens [1, 3, 13–15, 26]. Dorko et al. [11] studied the localisation of NADPH-d positive cells in the thymuses of mice and pheasants.

Our observation of the thymus of rats agrees with the results mentioned above. The NADPH-d-positive rat cells were observed as irregular agglomerates on the boundary of the cortex and the medulla. Indi-

vidual cells had irregular shapes and numerous projections. In our observations of NADPH-d-positive cells in the thymus of mice we have demonstrated differences in the number and type of these cells in the cortex and the medulla. While oval, diffuse, dispersed cells of a low intensity of coloration predominated in the cortex, the medulla contains a higher number of various types of NADPH-d-positive cells. They occurred in numerous agglomerations with varying intensity of coloration. Attention should be paid to cells that are dark in colour, of irregular shape and which contain small projections. These were frequently located in close proximity to vessels such as NADPH-d-positive nerve fibres. The cortex of the rabbit thymus showed only weak NADPH-d staining, similar to the thymic cortex of other animal species [14]. In contrast, the intensity of NADPH-d-positive labelling was greater in the medulla. Various types of cells were observed, recognisable according to the density of NADPH-d staining. This unequal staining may suggest an unequal utilisation of NO by these cells. The data presented by [2] indicates that lymphocyte functions appear to be dependent on L-arginine, the NO pathway. NADPH-d-positive cells of the medulla formed rounded oval groups, easily distinguishable at low magnification. One type of these cells was morphologically similar to those weakly stained in the cortex.

In this study our interest was focused mostly on the presence of some NADPH-d-positive nerve structures. However, these nerve fibres ran alongside blood vessels. The innervation of the thymus has previously been clearly demonstrated [6]. The non-adrenergic and non-cholinergic (MANC) neurotransmission in the peripheral nervous system has been discussed by other authors [7, 27]. The results of [20] suggest that NO may participate in the haemodynamic control of the pancreas, due to NADPH-d activity observed in both its endocrine and exocrine aspects. Similarly, in the thyroid gland NADPH-d-labelled neuronal bodies, nerve fibres and vascular endothelium were discovered [25].

By analogy with other peripheral targets, the possibility exists of the presence of NANC fibres among thymic innervation. Our results are able to support this possibility in the case of the mice and rabbit thymus, although no NADPH-d-positive nerve fibres have been demonstrated in the rat and chick thymus [15]. Our observation of the NADPH-d activity of nerve fibres in the mice and rabbit thymus disagrees with those of others, who have investigated NADPH-d activity in the above-mentioned animal

species. There is no clear explanation for this discrepancy, except for interspecies differences. In support of this view [20] it can be stated that interspecies differences were evident in the distribution of NADPH-d-positive nerve fibres in the pancreas, suggesting that NO may exert different modes of action on the pancreas from species to species. It should be noted, that for final proof of thymic NANC fibres, it will be necessary to carry out more experiments using other methods.

In summary, the present results reveal the distribution of NADPH-diaphorase-positive structures in the rat, mice and rabbit thymus. Especially remarkable were the blood vessels with clearly visible endothelial cells and with fine perivascular nerve fibres. These findings suggest, that NO may participate in the regulation of the thymic function.

## REFERENCES

1. Aimi Y, Fujimura M, Vincent SR, Kimura H (1991) Localization of NADPH-diaphorase containing neurons in sensory ganglia of the rat. *J Comp Neurol*, 306: 382–392.
2. Albina JR, Henry WL (1991) Suppression of lymphocyte proliferation through the nitric-oxide synthesizing pathway. *J Surg Res*, 110: 327–334.
3. Anderson CR (1992) NADPH-diaphorase positive neurons in the rat spinal cord include a subpopulation of autonomic preganglionic neurons. *Neurosci Lett*, 139: 280–284.
4. Boeckstaens GE, Pelckmans PA, Bult H, De Man JG, Herman AG, Van Maercke N (1992) The arginine-nitric oxide pathway mediates non-adrenergic non-cholinergic neurotransmission in gastrointestinal tissue. *Guanidino Compounds in Biology and Medicine*, Chapter 13, John Libbey Company Ltd, 89–96.
5. Bredt DS, Glatt CE, Hwang P M, Fotuhi M, Dawson TM, Snyder SH (1991) Nitric oxide synthase protein and mRNA are discretely localized in neuronal populations of the mammalian CNS together with NADPH diaphorase. *Neuron*, 7: 615–624.
6. Bulloch K, Pomerantz W (1984) Autonomic nervous system innervation of thymic-related lymphoid tissue in wildtype and nude-mice. *J Comp Neurol*, 228: 57–68.
7. Burnett AL, Lownstein CJ, Bredt DS, Chang TS, Snyder SH (1992) Nitric oxide: a physiologic mediator of penile erection. *Science*, 257: 401–403.
8. Dawson TM, Bredt DS, Fotuhi M, Hwang PM, Snyder SH (1991) Oxide synthase and neuronal NADPH-diaphorase are identical in brain and peripheral tissues. *Proc Natl Acad Sci, USA* 88: 7797–7801.
9. Dorko F, Gomboš A, Kocišová M, Gregor A (1993) Innervation of the thymus in the prenatal period. *Funct Develop Morphol*, 3; 1: 32–33.
10. Dorko F, Kocišová M, Gregor A, Dorko E (1996) The thymus — primary lymphatic organ. *Slov Lek*, 7: 15–17.
11. Dorko F, Kocišová M, Gregor A, Dorko E (1998) Localization of NADPH-d positive cells in thymuses of pheasant and mice. *Brat Lek Listy*, 99, 2: 104–107.
12. Dorko F, Kocišová M, Sirotáková M, Schmidtová K, Lovásová K, Dorko E (2000) The localization of NADPH-d positive cells and autonomic innervation of the rat thymus. *Pakistan J Biol Sci*, 5: 759–762.
13. Downing JEG (1994) Multiple nitric oxide synthase systems in adult rat thymus revealed using NADPH-diaphorase histochemistry. *Immunology*, 82: 659–664.
14. Grozdanovic Z, Baumgarten HG, Bruning G (1992) Histochemistry of NADPH-diaphorase, a marker for neuronal nitric oxide synthase, in the peripheral autonomic nervous system of the mouse. *Neuroscience*, 48: 225–235.
15. Gulati P, Chan AS, Leong SK (1993) NADPH-diaphorase positive cells in the chick and rat thymus. *Thymus*, 22: 117–124.
16. Hope BT, Michael GJ, Knigge KM, Vincent SR (1991) Neuronal NADPH-diaphorase is a nitric oxide synthase. *Proc Natl Acad Sci USA*, 88: 2811–2814.
17. Kiechle FL, Maliski T (1993) Nitric oxide biochemistry, pathophysiology, and detection. *Am J Clin Pathol* 100: 567–573.
18. Kocišová M, Gomboš A, Gregor A, Bánková E, Dorko F (1992) Innervation of the rat thymus. *Folia Fac Med Univ Šafarik Cass XLIX*: 53–58.
19. Mizukawa K (1990) Reduced nicotinamide adenine dinucleotide phosphate-diaphorase histochemistry: Light and electron microscopic investigations. In: *Methods in Neuroscience*. 3 Ed. Academic Press, Inc. 457–472.
20. Shimosegawa T, Takashi A, Akihiko S, Reishi A, Yoshifumi K, Masaru K, Takayoshi T (1993) NADPH-diaphorase activity in neurons of the mammalian pancreas: Coexpression with vasoactive intestinal polypeptide. *Gastroenterology*, 105: 999–1008.
21. Scherer-Singler U, Vincent SR, Kimura H, McGeer EG (1983) Demonstration of a unique population of neurons with NADPH-diaphorase histochemistry. *J Neurosci Methods*, 8: 229–234.
22. Schmidt HHHW, Gague GD, Nakane M, Pollock JS, Miller MF, Murad F (1992) Mapping of neural NO synthase in the rat suggests frequent localization with NADPH-diaphorase but not soluble guanylyl cyclase and novel paraneural functions for nitrinergic signal transduction. *J Histochem Cytochem* 40: 1439–1456.
23. Singh U (1984) Sympathetic innervation of fetal mouse thymus. *Eur J Immunol*, 14: 757–759.
24. Sirotáková M, Stopek D, Kocišová M, Dorko F (1997) Development of adrenergic and ACHE-positive innervation in bursa cloacalis (Fabricii) in pheasants: a histochemical study. *Folia Fac Med Univ Šafarik Cass*, 54: 71–81.
25. Syed MA, Leong SK, Chan AS (1994) Localization of NADPH-diaphorase reactivity in the chick and mouse thyroid gland. *Thyroid*, 4: 475–478.
26. Valtchanoff JG, Weinberg RJ, Rustioni A (1992) NADPH-diaphorase in the spinal cord of rats. *J Comp Neurol* 321: 209–222.
27. Vanderwinden JM, Mailleuz P, Schiffmann SN, Vanderhaeghen JJ, De Laet MH (1992) Nitric oxide synthase activity in infantile hypertrophic pyloric stenosis. *N Engl J Med* 327: 511–515.