

The thickness of the ventral midline of the spinal cord in human embryos during the fifth week

W. Woźniak, J. Łupicka, A. Sroka, M. Bruska, A. Pytel

Department of Anatomy, University of Medical Sciences, Poznań, Poland

[Received 21 July 2008; Accepted 21 July 2008]

The thickness of the ventral midline of the spinal cord was determined in 9 human embryos aged five weeks (developmental stages 13–15). This part of the spinal cord consists of floor plate, mantle and marginal layers. The floor plate ependymal cells form pseudostratified columnar epithelium. The thickness of the investigated structure varied from 20 to 50 micrometers at different levels of the spinal cord. (Folia Morphol 2008; 67: 205–208)

Key words: human neuroembryology, spinal cord, ventral midline cells, floor plate

INTRODUCTION

Differentiation of neuronal types and the establishment of appropriate connections between neurons and their targets are critical phases of the early development of the nervous system, and these events form the basis for the proper operation of the mature nervous system. Cells at the midline of the neural tube control axon projections and influence motor neurons and several types of interneuron differentiation [2, 12, 16].

The formation of precise neuronal circuits depends upon the ability of the growth cone, which guides axons, to navigate over a long distance. The guidance cues exist in either diffusible or cell surface-associated forms [7, 21]. Growth cone-associated cell surface receptors interpret these signals as positive/attractive (netrin) or negative/repulsive (slit and semaphorin proteins) forces [15, 24].

The midline of the developing central nervous system represents an important choice point for pathfinding axons [8]. Recent experiments show that axial mesoderm can stimulate formation of specific ventral cell types in the spinal cord, including floor plate and motor neurons. Generation of these cell types is dependent on the secreted signalling mole-

cule Sonic hedgehog (Shh), which is produced by axial mesodermal midline cells of the notochord. Shh signalling from the notochord induces development of the ventral midline cells and the floor plate which also expresses Shh together with other genes [4].

The ependymal lining floor plate has been investigated in ultrastructural and cytochemical studies on the human spinal cord by Gamble [3], Malinsky and Brichova [13] and Tanaka et al. [20]. In the present study we investigated the thickness and histological structure of the ventral midline of the spinal cord in human embryos aged five weeks.

Although morphological criteria do not permit delimitation of the ventral midline cells and the floor plate in the transverse plane, it is possible to measure the thickness of the ventral midline cells of the spinal cord in the sagittal plane.

MATERIAL AND METHODS

A study was made on human embryos aged five weeks (developmental stages 13, 14 and 15, Table 1). The embryos were from the Collection of the Department of Anatomy of the University of Medical Sciences in Poznań. All embryos were embedded *in toto* in paraplast and serially sectioned in the

Table 1. Crown-rump length, developmental stage, and age in days of the embryos examined

| Catalogue number | C-R length [mm] | Developmental stage | Age in postovulatory days | Plane of section |
|------------------|-----------------|---------------------|---------------------------|------------------|
| B 171 | 4 | 13 | 32 | Horizontal |
| B 194 | 6 | 13 | 32 | Horizontal |
| B 218 | 4.5 | 13 | 32 | Horizontal |
| A 19 | 7 | 14 | 32 | Horizontal |
| B 207 | 6 | 14 | 32 | Horizontal |
| P 41 | 5 | 14 | 33 | Horizontal |
| B 115 | 8 | 15 | 33 | Horizontal |
| PJK 20 | 7 | 15 | 33 | Horizontal |
| PJK 21 | 10 | 15 | 37 | Horizontal |

horizontal plane. Measurements of the ventral midline of the spinal cord were made on sections from various levels of the spinal cord with the aid of a Leica Olympus Microscope using program AxioVision LE with the Canon Module. The thickness of the ventral midline was determined together with the floor plate.

The structure of the ventral midline of the spinal cord was also investigated in histological sections.

RESULTS

In embryos at stage 13 the mantle layer of the ventral midline of the spinal cord consists of 4 rows of cells with oval nuclei. The ependyma at this part is formed of columnar cells with basally located nuclei. The marginal layer is clearly visible (Fig. 1). The thickness of the ventral midline in the embryos studied varied from 20 to 40 μm at different levels of the spinal cord and was greater in the cervical and upper thoracic segments than in the lumbar segment of the spinal cord (Table 2).

In embryos at stage 14 the mantle layer of the ventral midline is thicker than that of embryos in stage 13 and consists of 5 or 6 rows of cells (Fig. 2). The ependymal layer consists of pseudostratified columnar epithelium. In the mantle layer the crossing commissural axons are visible (Fig. 3). The thickness of this ventral midline cells varies from 30 to 50 μm (Table 2).

In embryos at stage 14 the central canal of the spinal cord diminishes and the anterior horns are well developed. The structure and thickness of the ventral midline of the spinal cord are similar to that of embryos at stage 14 (Fig. 4).

DISCUSSION

The ventral midline cells of the central nervous system are important in the neural tube, playing a role in the differentiation part of motor neurons

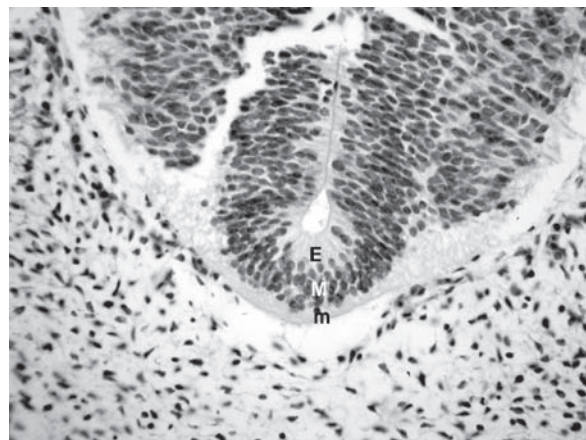


Figure 1. Horizontal section of the spinal cord of an embryo at stage 13. Staining with toluidine blue, $\times 400$; E — ependyma of the floor plate, M — mantle layer, m — marginal layer.

and the establishment of commissural projections [2, 14]. The differentiation of neurons in the ventral neural tube is dependent on inductive signals from the notochord and floor plate, which is formed of specialized glia in a narrow strip at the ventral midline [1, 14, 17, 18, 22].

According to O'Rahilly and Müller [16], the floor plate is formed by the ventromedial cells of the epinotochordal part of the neural plate or tube and is induced by the notochord. It differs regionally. Cells in the midbrain can induce the production of dopaminergic neurons, whereas those of the rhombencephalon develop into the septum medullae.

According to many authors, beginning with the descriptions of His [5], the floor plate is made up of columnar ependymal cells that span the width of the neural tube at its ventral midline. The anterior boundary of the floor plate has been placed at the hindbrain-midbrain junction [9, 10] but there are several

Table 2. Thickness of the ventral midline of the spinal cord in the embryos examined

| Catalogue number of embryo | Developmental stage | Level of section | Thickness in micrometers |
|----------------------------|---------------------|--|--------------------------|
| B 171 | 13 | Upper thoracic Lumbar | 40 30 |
| B 194 | 13 | Cervical Lower thoracic | 30 20 |
| B 218 | 13 | Upper thoracic Lower thoracic Lumbar | 20 20 20 |
| P 41 | 14 | Lower cervical Upper thoracic Lower thoracic | 40 40 30 |
| A 19 | 14 | Upper thoracic Lower thoracic | 30 40 |
| B 207 | 14 | Cervical Upper thoracic Lumbar | 50 50 40 |
| B 115 | 15 | Cervical Upper thoracic Lower thoracic | 40 40 40 |
| PJK 21 | 15 | Cervical Upper thoracic Lower thoracic | 30 30 30 |
| PJK 20 | 15 | Cervical Upper thoracic Lower thoracic | 50 40 40 |

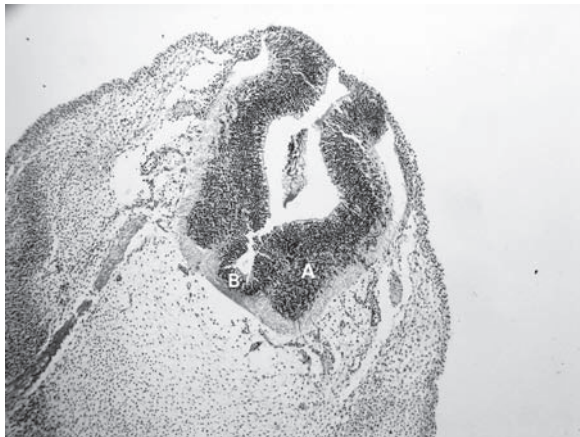


Figure 2. Horizontal section of the spinal cord in an embryo at stage 14. Bodian's protargol, $\times 100$; A — anterior horn, B — ventral midline cells.

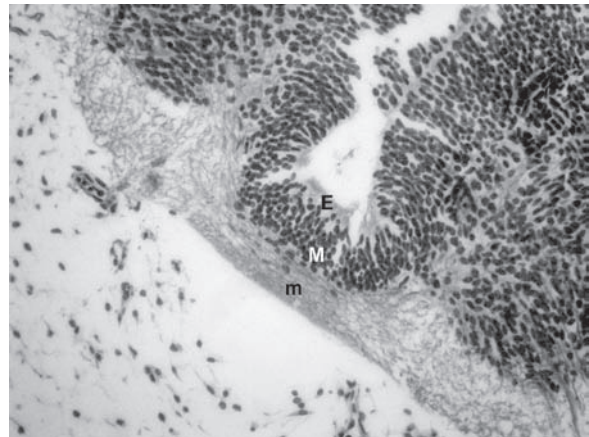


Figure 3. Horizontal section through the ventral part of the spinal cord in an embryo at stage 14. Bodian's protargol, $\times 400$; E — ependyma of the floor plate, M — mantle layer, m — marginal layer with commissural fibres.

reasons for believing that this plate extends through the midbrain into the posterior diencephalon and ends near the mammillary region [2, 11, 17, 18].

In the present study it was shown that in the early embryonic human spinal cord the ventral midline of

the spinal cord consists of ependymal lining cells, mantle, and marginal layers. The thickness of the part of the spinal cord under investigation slightly increases at stages 14 and 15, and during these stages commissural axons are visible crossing the marginal layer.

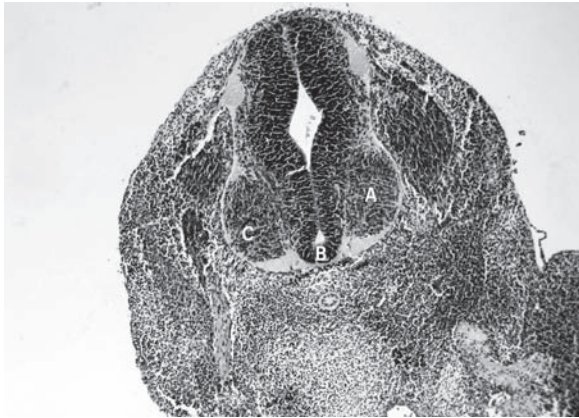


Figure 4. Horizontal section of the spinal cord in an embryo at stage 15. Haematoxylin and eosin, $\times 100$; A — anterior horn, B — ventral midline cells, C — spinal ganglion.

Sturrock [19] in mouse, Yoshioka and Tanaka in rats [23] and Gamble [3] and Malinsky and Brichova [13] in humans found that the floor plate ependymal cells differentiate during an earlier stage of embryonic development and are closely associated with the growth of decussating nerve fibres in the marginal layer, as was observed in our study. Midline-associated glial cells appear to be a source of contact-dependent or diffusible guidance cues for commissural axons [6].

REFERENCES

1. Briscoe J, Ericson J (2001) Specification of neuronal fates in the ventral neural tube. *Curr Opin Neurobiol*, 11: 43–49.
2. Colamarino SA, Tessier-Lavigne M (1995) The role of the floor plate in axon guidance. *Annu Rev Neurosci*, 18: 497–529.
3. Gamble HJ (1969) Electron microscope observations on the human fetal and embryonic spinal cord. *J Anat*, 104: 435–453.
4. Götz K, Briscoe J, Ruther U (2005) Homozygous *Ft* embryos are affected in floor plate maintenance and ventral neural tube patterning. *Dev Dyn*, 233: 623–630.
5. His W (1888) Zur Geschichte des Gehirns, sowie der zentralen und peripherischen Nervenbahnen beim menschlichen Embryo. *Abh Math Phys Kl Konigl Sachsischen Gesellschf Wiss*, 8: 341–392.
6. Kadison SR, Murakami F, Matise MP, Kaprielian Z (2006) The role of floor plate contact in the elaboration of contralateral commissural projections within the embryonic mouse spinal cord. *Dev Biol*, 296: 499–513.
7. Kaprielian Z, Imondi R, Runko E (2000) Axon guidance at the midline of the developing CNS. *Anat Rec (New Anat)*, 261: 176–197.
8. Kaprielian Z, Runko E, Imondi R (2001) Axon guidance at the midline choice point. *Dev Dyn*, 221: 154–181.
9. Kingsbury BF (1920) The extent of the floor plate of His and its significance. *J Comp Neurol*, 32: 113–135.
10. Kingsbury BF (1930) The developmental significance of the floor-plate of the brain and spinal cord. *J Comp Neurol*, 50: 177–207.
11. Klar A, Baldassare M, Jessel TM (1992) F-spondin: a gene expressed at high levels in the floor plate encodes a secreted protein that promotes neural cell adhesion and neural expression. *Cell*, 69: 95–100.
12. Maghadam KS, Chen A, Heathcote R (2003) Establishment of a ventral cell fate in the spinal cord. *Dev Dyn*, 227: 552–562.
13. Malinsky J, Brichova H (1967) Fine structures of ependyma in spinal cord of human embryos. *Folia morph (Prague)*, 15: 68–78.
14. Mambetisaeva ET, Andrews W, Cumurri L, Annan A, Sundaresan U (2005) Robo family of proteins exhibit differential expression in mouse spinal cord and Robo-Slit interaction is required for midline crossing in vertebrate spinal cord. *Dev Dyn*, 233: 41–51.
15. Mitchell KJ, Doyle JL, Serafini T, Kennedy TE, Tessier-Lavigne M, Goodman CS, Dicsion BJ (1996) Genetic analysis of Netrin genes in *Drosophila*: Netrins guide CNS commissural axons and peripheral motor axons. *Neuron*, 17: 203–215.
16. O’Rahilly R, Müller F (2006) The embryonic human brain: an atlas of developmental stages. Wiley-Liss, Hoboken, NJ.
17. Placzek M (1995) The role of the notochord and floor plate in inductive interactions. *Curr Opin Genet Dev*, 5: 499–506.
18. Placzek M, Jessel TM, Dodd J (1993) Induction of floor plate differentiation by contact-dependent, homogenetic signals. *Development*, 117: 205–218.
19. Sturrock RR (1981) An electron microscopic study of the development of the ependyma of the central canal of the mouse spinal cord. *J Anat*, 132: 119–136.
20. Tanaka O, Yoshioka T, Shinohara H (1988) Secretory activity in the floor plate neuroepithelium of the developing human spinal cord: morphological evidence. *Anat Rec*, 222: 185–190.
21. Tessier-Lavigne M, Goodman CS (1996) The molecular biology of axon guidance. *Science*, 274: 1123–1133.
22. Yamada T, Placzek M, Tanaka H, Dodd J, Jessel TM (1991) Control of cell pattern in the developing nervous system: polarizing activity in the floor plate and notochord. *Cell*, 64: 635–647.
23. Yoshioka T, Tanaka O (1989) Ultrastructural and cytochemical characterization of the floor plate ependyma of the developing rat spinal cord. *J Anat*, 165: 87–100.
24. Zou Y, Stoeckli E, Chen H, Tessier-Lavigne M (2000) Squeezing axons out of the gray matter: a role for slit and semaphorin proteins from midline and ventral spinal cord. *Cell*, 102: 363–375.