

GA₃ content in young and mature antheridia of *Chara tomentosa* estimated by capillary electrophoresis

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Abstract: The content of gibberellic acid (GA₃) in male sex organs of *Chara tomentosa* L. was estimated using capillary electrophoresis. Young antheridia contained 0.25 µg GA₃ while mature ones 0.48 µg per antheridium. Although there are significant differences in GA₃ content in antheridia between *C. vulgaris* and *C. tomentosa*, these values calculated per one spermatid are 2.4 and 3.3 pg, respectively. The present results compiled with the previous knowledge about regulation of GA₃-dependent development of *Characeae* species allow an implication that the mechanisms controlling antheridia differentiation in both species can be similar.

Key words: Antheridia - Capillary electrophoresis - *Chara tomentosa* - Gibberellic acid

Introduction

Studies on generatively mature thallus of algae *C. vulgaris* revealed the involvement of gibberellic acid in the regulation of its development and spermatogenesis. Exogenous gibberellic acid hastens antheridium development and increases the number of spermatozoids in antheridial filaments [2, 16]. Simultaneously, lengthening of thallus internodes and pleuridium axes becomes restricted [11]. The above data were supported by tests with AMO1618 - an inhibitor of gibberellin synthesis - showing an opposite effect [11]. During spermatogenesis, GA₃ increases endopolyploidy levels in some non-generative cells in antheridium, e.g. DNA content in manubria rises by approximately 20% [16].

It has been repeatedly shown that plasmodesmata, which form a system of intercellular communication, play an important role in differentiation of multicellular plant organisms [10]. Spontaneous or induced break of intercellular connections stops symplasmic communication between *C. vulgaris* antheridia and thallus [7, 17]. During first stages of antheridium development, plasmodesmata are open. Later, due to spontaneous symplasmic isolation, plasmatic connections between shield cells as well as between shield cell, basal cell and manubrial cell break [10]. During antheridium proliferative stage, plasmodesmata between antheridial filament cells

become selectively plugged with an osmophilic substance. This leads to the formation of antheridial filament domains characterized by shifted synchrony of dividing cells.

Spermiogenesis (sperm differentiation) is also preceded by the break of plasmodesmal connections due to spontaneous symplasmic isolation between basal and capitular cells as well as between basal and subbasal cells. Two final mitotic divisions of antheridial filament cells occur in complete symplasmic isolation between antheridium and thallus [7]. Simultaneously, endoreplication in manubria and capitular cells stops [7]. This may be due to lower gibberellin activity inside antheridium, as suggested by the decrease in radioactive GA₃ penetration into antheridium [8]. This result seems to be supported by capillary electrophoresis measurements of GA₃ content [5, 3]. It was observed that *C. vulgaris* antheridia in the proliferative phase (I and II thallus nodes) contained approx. 5.3 times more GA₃/antheridium than those during spermiogenesis [5].

The aim of the present study was to establish whether there was a difference in GA₃ content between young and mature antheridia of another *Characeae* species, i.e. *C. tomentosa*. In this dioecious algae, the number of chromosomes in haploid antheridial filament cells is 14 and DNA content for 1C - 7.4 pg. In *C. vulgaris*, which is a monoecious plant, the number of chromosomes is 28 (1n) and content of 1C DNA is 13.5 pg [19]. During proliferative phase, subsequent mitotic cycles (5-6) yield 64 and 46 cells in antheridial filaments of *C. vulgaris* and *C. tomentosa*, respectively. In *C. vulgaris*,

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Table 1. Comparison of GA₃ content in young and mature antheridia of *Chara vulgaris* and *Chara tomentosa*

GA ₃ content per	<i>C. tomentosa</i>		<i>C. vulgaris</i>	
	Young	Mature	Young	Mature
Gram fresh weight (μg)	137 ± 20	86 ± 9	512 ± 95*	32.5 ± 5*
Antheridium (μg)	0.25 ± 0.056	0.48 ± 0.051	0.09 ± 0.003*	0.017 ± 0.001*
Antheridial filament (ng)	0.08 ± 0.017	0.15 ± 0.016	0.81 ± 0.099	0.15 ± 0.039
Spermatozoid (pg)	–	3.3 ± 0.4	–	2.4 ± 0.014

Means ± SD of three independent experiments. *Data derived from [5].

S+G2+M cell cycle for two-cell antheridial filaments lasts 42 h and decreases to 26 h at 32-cell stage, while in *C. tomentosa* these values are 41.5 h and 21 h, respectively. Budding capitular cells produce about 111 antheridial filaments in *C. vulgaris* and as many as 3217 in *C. tomentosa*. The average number of spermatids in mature antheridia is 7000 and 145700 for *C. vulgaris* and *C. tomentosa*, respectively [18].

Materials and methods

Antheridia from apical part of *Chara tomentosa* L. were investigated. The plants were collected from the pond in Łódź Botanical Garden. The thalli were rinsed with distilled water and then antheridia from proliferative stage (I-II nodes) and spermiogenesis (III-IV nodes) were examined separately.

Isolation and purification of GA₃ were carried out according to [1]. The identification of GA₃ was done using the Capillary Electrophoresis System - BioFocus 3000 (BioRad) using the CZE (capillary zone electrophoresis): in 30 mmol dm⁻³ sodium borate buffer, pH 8.5; capillary: 24×25 μm, silica coated; detection: 258 nm; injection: high pressure, 20 psi; temperature: ambient; voltage: 10 kV. Electropherograms were recorded and analysed using BioFocus Integration software 5.0, based on Microsoft Windows, at a detector rise time setting of 1 s and data sampling rate of 5 Hz. The UV spectrum of GA₃ was measured by BioFocus 3000 UV scanning detector. The retention times of standard and measured GA₃ are shown in Figure 1.

Results and discussion

Gibberellins, due to their involvement in induction and regulation of sex organs in algae [13], homosporous ferns [21] and higher plants [20, 22], are called plant sex hormones. High content of GA₃ in young nodes and several-fold less content of GA₃ in older nodes in male *C. tomentosa* thalli was revealed by capillary electrophoresis and biotest. The average GA₃ content in the male thallus was 4-fold higher than in the female one [6]. High GA₃ content was also observed in young nodes of *C. vulgaris* thalli [4].

Electrophoretic measurements showed that 1 g fresh weight (FW) of *C. tomentosa* young antheridia contained about 137 μg GA₃ while mature ones 87 μg which gives 0.48 μg and 0.25 μg of GA₃ per antheridium, respectively (Tab.1). Comparing the present results with data from Maszewski's paper [18] it was estimated that

the amount of GA₃ per one mature antheridial filament of both *C. vulgaris* and *C. tomentosa* was 0.15 μg while in young antheridia GA₃ content was 10-fold higher in *C. vulgaris* than in *C. tomentosa*. Finally, however, the amount of GA₃ per one spermatid in mature antheridium of *C. vulgaris* and *C. tomentosa* was similar: 2.4 and 3.3 pg, respectively.

The present findings indicate that in *C. tomentosa*, young antheridia contain 60% more GA₃ per 1 g FW than mature ones. Similar changes in GA₃ content related to the developmental stage were observed in *C. vulgaris* [5]. It is worth noticing that a similar amount of GA₃ corresponds to one spermatid in mature antheridia of both species (Tab. 1).

The results obtained on the basis of endogenous and exogenous GA₃ measurements clearly indicate that proliferation phase of antheridial filament development requires a high level of GA₃ both for division of antheridial filament cells and for endoreplication of nongenerative antheridial cells. Low concentration of GA₃ in mature antheridia may reflect no need for GA₃ during spermiogenesis, when its level is limited due to spontaneous symplasmic isolation observed during development of *C. vulgaris* antheridia [17]. Low GA₃ level may be required for normal spermiogenesis, when spermatid growth is blocked as high chromatin condensation and starch accumulation in plastids occur [14]. Low GA₃ level facilitates starch accumulation in mature nodes of thallus [4]. Stimulation of internode elongation and maintenance of thallus apical domination also requires lower GA₃ level [2].

The presented data suggest that regardless of differences between *C. vulgaris* and *C. tomentosa* developmental cycles, GA₃ may play a similar role in the regulation of sex organ development. In both species, high and low GA₃ levels are associated with antheridial filament cell proliferation and spermatid differentiation, respectively. This similarity is especially visible when GA₃ content is expressed per one spermatid. It may mean that differentiation mechanisms of *C. tomentosa* antheridia are similar to those of *C. vulgaris* where intercellular communication plays a crucial role.

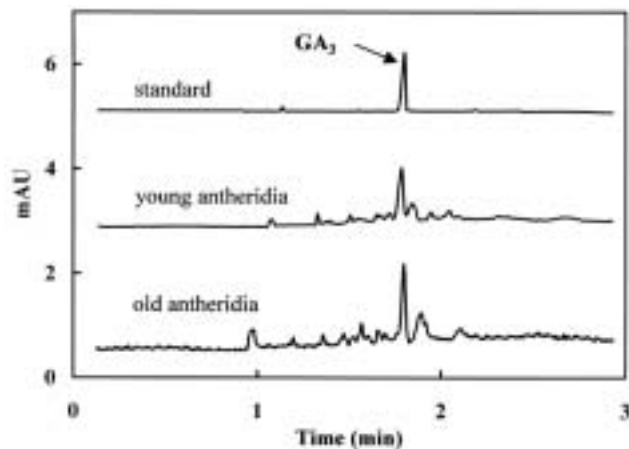


Fig. 1. Capillary zone electrophoresis electropherogram of GA₃ isolated from young and mature antheridia of *Chara tomentosa* and electropherogram of standard gibberellic acid (GA₃).

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References

- [1] Fujioka S, Yamane H, Spray CR, Gaskin P, MacMillan J, Phinney BO, Takahashi N (1988) Qualitative and quantitative analyses of gibberellins in vegetative shoots of normal, dwarf-1, dwarf-2, dwarf-3, and dwarf-5 seedlings of *Zea mays* L. *Plant Physiol* 88: 1367-1372
- [2] Godlewski M, Kwiatkowska M (1980) Effect of gibberellic acid on the formation and development of antheridia and oogonia in *Chara vulgaris* L. *Acta Soc Bot Pol* 49: 459-469
- [3] Kaźmierczak A (1999) Determination of GA₃ in *Chara vulgaris* by capillary electrophoresis system. *Acta Physiol Plant* 21: 344-348
- [4] Kaźmierczak A (2001) The relationship between fertility and contents of gibberellic acid, sugars and dry mass in apical parts of *Chara vulgaris* thalli. *Biol Plant* 44: 439-441
- [5] Kaźmierczak A, Kwiatkowska M, Popłońska K (1999) GA₃ content in antheridia of *Chara vulgaris* at the proliferative stage and in spermiogenesis estimated by capillary electrophoresis. *Folia Histochem Cytobiol* 37: 49-52.
- [6] Kaźmierczak A, Rosiak M (2000) Content of gibberellic acid in apical parts of male and female thalli of *Chara tomentosa* in relation to the content of sugars and dry mass. *Biol Plant* 43: 369-372
- [7] Kwiatkowska M (1988) Symplasmic isolation of *Chara vulgaris* antheridium and mechanisms regulating the process of spermatogenesis. *Protoplasma* 142: 137-146
- [8] Kwiatkowska M (1991) Autoradiographic studies on the role of plasmodesmata in the transport of gibberellin. *Planta* 189: 294-305
- [9] Kwiatkowska M (1995) Effect of symplasmic isolation and antigibberellin treatment on morphogenesis in *Chara*. *Folia Histochem Cytobiol* 33: 133-137
- [10] Kwiatkowska M (2003) Plasmodesmal changes are related to different developmental stage of antheridia of *Chara* species. *Protoplasma* 222: 1-11
- [11] Kwiatkowska M, Godlewski M (1980) Effect of gibberellic acid and AMO-1618 on the development of vegetative systems in generatively matured thalli of *Chara vulgaris* L. *Acta Soc Bot Pol* 49: 445-458
- [12] Kwiatkowska M, Godlewski M (1988) Studies on the role of gibberellins in the regulation of spermatogenesis in *Chara vulgaris* L. *Acta Soc Bot Pol* 57: 547-553.
- [13] Kwiatkowska M, Gosek A, Godlewski M (1991) Effect of GA₃, IAA and their mixtures on the formation and development of cell systems in the vegetative and generative thallus of *Chara vulgaris* L. *Acta Soc Bot Pol* 60: 313-326
- [14] Kwiatkowska M, Popłońska K (2002) Further ultrastructural research of *Chara vulgaris* spermiogenesis: endoplasmic reticulum, structure of chromatin, ³H-lysine and ³H-arginine incorporation. *Folia Histochem Cytobiol* 40: 85-97
- [15] Kwiatkowska M, Popłońska K, Żylińska K (1990) Biological role of endoreplication in the process of spermatogenesis in *Chara vulgaris* L. *Protoplasma* 155: 176-187
- [16] Kwiatkowska M, Wojtczak A, Popłońska K (1998) Effect of GA₃ treatment on the number of spermatozooids and endopolyploidy levels of non-generative cells in antheridia of *Chara vulgaris* L. *Plant Cell Physiol* 39: 1388-1390
- [17] Kwiatkowska M, Wojtczak A, Popłońska K (2002) Ultrastructural and autoradiographic study of *Chara vulgaris* L. manubria. *Acta Biol Cracov Ser Bot* 44: 99-105
- [18] Maszewski J (1991) Endopolyploidization patterns in non-generative antheridial cells in mono- and dioecious *Chara* spp. (*Characeae*) with different DNA C-values. *Plant Syst Evol* 177: 39-52
- [19] Maszewski J, Kołodziejczyk P (1991) Cell cycle duration in antheridial filaments of *Chara* spp. (*Characeae*) with different genome size and heterochromatin content. *Plant Syst Evol* 175: 23-38
- [20] Metzger JM (1995) Hormones and reproductive development. In: *Plant Hormones*, Davis PJ [Ed]. Kluwer Academic Publishers, Dordrecht, Netherlands, pp 617-648
- [21] Yamane H (1998) Ferns antheridiogens. *Int Rev Cytol* 184: 1-31
- [22] Yamasaki S, Fujii N, Takahashi H (2000) The ethylene-regulated expression of *CS-ETR2* and *CS-ERS* genes in cucumber plants and their possible involvement with sex expression in flowers. *Plant Cell Physiol* 41: 608-616

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