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Connexin 43 recruits E-cadherin expression and inhibits the malignant behaviour of lung cancer cells

Hong-Tao Xu, Qing-Chang Li, Yong-Xing Zhang, Yue Zhao, Yang Liu, Zhi-Qiang Yang, En-Hua Wang

Department of Pathology, College of Basic Medical Sciences, China Medical University and Department of Pathology, the First Affiliated Hospital of China Medical University, Shenyang 110001, China.

Abstract: The interaction of connexin 43 and E-cadherin may play an important role in carcinogenesis and malignant behaviour of tumours. In this study, we examined the relationship between connexin 43 and E-cadherin in human non-small cell lung cancers (NSCLC). Expression levels of connexin 43 and E-cadherin were examined in 107 NSCLC specimens by immunohistochemistry. The connexin 43 gene was transfected into lung cancer LH₇ cells. The protein localizations and levels of connexin 43 and E-cadherin were detected using immunofluorescence staining and western blot. Cell cycle and proliferation of lung cancer cells were examined using flow cytometry and MTT. We found that reduced expression of both connexin 43 and E-cadherin significantly correlated to poor differentiation, advanced TNM stage, and lymph note metastasis of NSCLCs. Connexin 43 and E-cadherin expression significantly correlated with each other. Over-expression of connexin 43 significantly induced E-cadherin expression. Moreover, connexin 43-transfected LH₇ cells showed significantly decreased cell proliferation. The percentage of cells in G1 phase increased, while the number of cells in S and G2 phases significantly decreased. We concluded that concurrent reduction of connexin 43 and E-cadherin may contribute to the development of lung cancer. Connexin 43 may induce E-cadherin expression and inhibit cell proliferation and progression of lung cancer.

Key words: Connexin 43 - E-cadherin - Lung Cancer

Introduction

Both gap junctions and adhesion molecules play a critical role in cell contact inhibition, differentiation, proliferation, and homeostasis of various tissues. Recent evidence suggests that gap junctions are not simply pore-forming proteins, but interact with cell adhesion-associated proteins and participate in signalling events [1-3].

Connexin 43 is one of the most common connexins and the major connexin homolog expressed in lung tissue [4-6]. Connexin 43 and E-cadherin play important roles in carcinogenesis and tumour metastasis [7-9]. They are concurrently expressed in many tumours [10,11]. Hernandez-Blazquez et al. indicated that E-cadherin can regulate connexin 43 expression and

Correspondence: Dr. En-Hua Wang, Department of Pathology, College of Basic Medical Sciences, China Medical University and Department of Pathology, the First Affiliated Hospital of China Medical University, Shenyang 110001, China. Tel: (+8624) 23261638, Fax: (+8624) 23261638, e-mail: wangeh@hotmail.com.

function [12]. However, it is still unclear that whether connexin 43 and E-cadherin are concurrently expressed in lung cancers and affect each other or not. In this study, we examined the expression of connexin 43 and E-cadherin in primary lung cancers to clarify the correlation between connexin 43 and E-cadherin, and explore the clinical and pathological implications of expression of these proteins. We also transfected LH_7 cells with connexin 43 gene and investigated the possible effect of connexin 43 on E-cadherin.

Materials and Methods

Patients. A total of 107 samples and 15 corresponding normal lung tissue samples were selected randomly from patients with squamous cell carcinomas (SCC) or adenocarcinomas who underwent surgery in the First Affiliated Hospital of China Medical University between 2001 and 2003. The study was conducted according to institutional review board regulations at China Medical University. There were 69 males and 38 females in our study, with ages ranging from 33 - 76 years, (mean = 57 years). The tumours were diagnosed as SCC (n = 45) or adenocarcinomas (n = 62) [13]. These tumours showed different degrees of differentiation and were classified as well- (n = 38), moderately- (n = 30), or poorly- (n = 39)

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Table 1. Expression	on of conneyir	43 and	F-cadherin	and their r	elation to	a clinico	nathologic charac	eteristics

Characteristics	n		Connexin 43		E-cadherin			
Characteristics		Positive	Negative	P	Positive	Negative	P	
E-cadherin								
Positive	72	51	21	< 0.001				
Negative	35	3	32					
Differentiation								
Well	38	26	12	0.009	30	8	0.002	
Moderate	30	15	15		24	6		
Poor	39	13	26		18	21		
Histological type								
Adenocarcinoma	62	33	29	0.503	42	20	0.907	
Squamous cell carcinoma	45	21	24		30	15		
TNM stage								
1	50	33	17	0.006	39	11	0.001	
П	30	13	17		23	7		
III-IV	27	8	19		10	17		
Lymph node metastasis								
Yes	57	16	41	< 0.001	32	25	0.009	
No	50	38	12		40	10		

differentiated [13]. Fifty-seven cases indicated lymphatic metastasis. All the tumours were classified as stages I, II, and III-IV (n = 50, 30, and 27, respectively) [14,15]. All the resected specimens were fixed with 10% neutral-buffered formalin and embedded in paraffin blocks.

Immunohistochemistry. Tissue blocks were cut into 4-μm sections, deparaffinized, rehydrated, and immunostained with monoclonal anti-connexin 43 antibody (1:50; Santa Cruz Biotechnology, USA) and monoclonal anti-E-cadherin antibody (1:100; Santa Cruz Biotechnology, USA). Detection was performed using the streptavidin-peroxidase method. Negative control slides were stained in the absence of primary antibodies.

Based on the scale of Nemeth et al. [16], the immunostaining grade of connexin 43 and E-cadherin was as follows: specimens in which none, $\leq 25\%$, > 25% - < 75%, and $\geq 75\%$ of tumour cells showed positive staining were defined as (-), (+), (++), and (+++), respectively. Samples scoring (-) - (+) were considered negative, and those that scored (++) - (+++) were considered positive.

Cell culture and transfection. The LH_7 cell line is derived from the PG cell line, established from a human pulmonary giant cell carcinoma. LH_7 cells were a gift from Dr. Jie Zheng, College of Medicine, Beijing University, China. LH_7 cells are highly metastatic and express low levels of connexin 43 [17, 18]. The LH_7 cells were grown in RPMI 1640 medium (GiBCO, USA) with 10% fetal calf serum (GiBCO, USA), at 37°C in a humidified atmosphere (5% CO₂ 95% air).

The connexin 43 expression vectors pcDNA3.1(+)-Cx43 were obtained from Professor YW Zhang (Tokyo Medical and Dental

University, Japan) as a gift [2]. LH $_7$ cells were transfected with pcDNA3.1(+)-Cx43 or pcDNA3.1(+) as control using FuGENE 6 transfection reagent according to the manufacturer's instructions (Roche, USA). After 36 h, the culture medium was changed and G418 (Roche, USA) was added at a concentration of 350 μ g/ml. Culture medium was changed every three days. After four weeks of selection, six neomycin resistant clones were selected and kept in culture with the same concentration of G418 during every following experiment.

Western blot. LH $_7$ cells transfected with pcDNA3.1(+)-Cx43 (called LH $_7$ -Gja1 cells) or pcDNA3.1(+) [called LH $_7$ (-) cells] were lysed and separated by 10% SDS-PAGE, and were transferred to a polyvinylidene fluoride membrane. The transferred samples were further incubated with anti-connexin 43 antibody (1:300) and anti-E-cadherin antibody (1:300) overnight. The proteins were visualized with an automatic electrophoresis analytic system (Chemilmager 5500, AlPha InnCh, USA). β-actin was used as internal control

Immunofluorescence staining. Cells were fixed, permeablised and incubated with anti-connexin 43 (1:300) and anti-E-cadherin antibodies (1:300) overnight, and labelled with a FITC-conjugated secondary antibody for 1 h, followed by staining with 0.1% DAPI for 3 min. Cells examined using an immunofluorescence microscope (BX60, OLYMPUS, Japan).

Cell cycle and proliferation. The cells were grown in a 96-well plate at a density of 1.5×10^6 cells/ml for 48 h, and stained with 1 mg/ml propidium iodide (Sigma, USA). The percentages of LH₇-Gja1, LH₇,

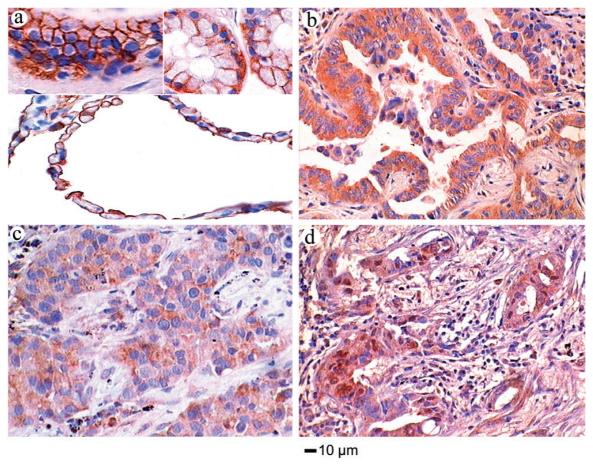


Fig. 1. Expression of connexin 43 and E-cadherin in lung cancers and normal lung tissues. (a) upper left: membrane expression of E-cadherin in normal epithelial cells of tunica mucosa bronchiorum. Upper right and below: membrane expression of connexin 43 in normal bronchial submucosal glands and alveolar epithelial cells. (b) E-cadherin expressed in cytoplasm of well-differentiated adenocarcinoma cells. (c) Connexin 43 expressed in cytoplasm of poorly-differentiated squamous cell carcinoma cells. (d) Connexin 43 expressed in cytoplasm and nucleus of moderately-differentiated adenocarcinoma cells.

or LH₇(-) cells in G1 phase, G2 phase and S phase were determined by flow cytometry (FACSCalibur, BD Biosciences, USA).

The CellTiter 96 AQueous cell proliferation assay (Promega, USA) was performed to study alterations in cell proliferation, according to the manufacturer's instructions. The absorbance at 490 nm, which is directly proportional to the number of living cells in culture, was detected in LH₇, LH₇(-) and LH₇-Gja1 cells each day for 5 d after transfection.

Statistical analyses. The χ^2 test was used to clarify the relationships among connexin 43, E-cadherin and clinicopathologic characteristics. The Mann and Whitney U test was used to analyze the results of the western blot, cell growth rate, and cell cycle experiments. P values less than 0.05 were considered statistically significant.

Results

Reduced expression of both connexin 43 and E-cadherin is associated with poor differentiation, advanced TNM stages and lymph node metastasis in lung cancers

Connexin 43 and E-cadherin were typically expressed at the cell membrane in all 15 normal lung

tissues (Fig. 1a). However, in lung cancer tissues, expression of connexin 43 was positive in only 50.5% (54/107) of samples. E-cadherin expression was positive in only 67.3% (72/107) of lung cancer samples. Expression of connexin 43 and E-cadherin in the samples was primarily cytoplasmic, and membrane expression was dramatically reduced (Figs. 1b and 1c). Several samples showed simultaneous nuclear and cytoplasmic expression (Fig. 1d). Correlations between the clinicopathologic characteristics of the samples and expression patterns of connexin 43 and E-cadherin are summarized in Table 1. Significant reduction of connexin 43 and E-cadherin expression was associated with patient samples that are poorly-differentiated, had advanced TNM stages and lymph node metastases. Reduced connexin 43 and E-cadherin levels were independent of tumour histological types. Interestingly, expression of connexin 43 and E-cadherin were significantly correlated with each other (P < 0.001, Contingency coefficient = 0.504; Table 1).

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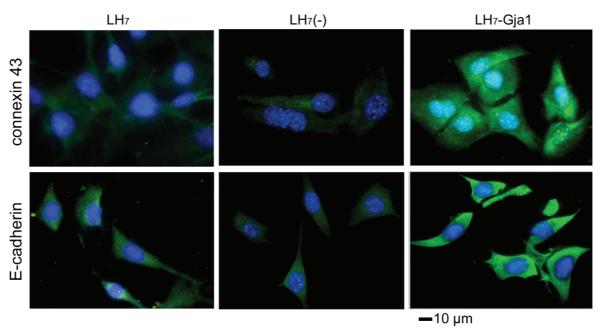


Fig. 2. Immunofluorescence of connexin 43 and E-cadherin in LH_7 , $LH_7(-)$ and LH_7 -Gja1 cells. The expression of connexin 43 and E-cadherin in LH_7 or $LH_7(-)$ cells is very low. After transfection, the expression of connexin 43 and E-cadherin is enhanced in LH_7 -Gja1 cells.

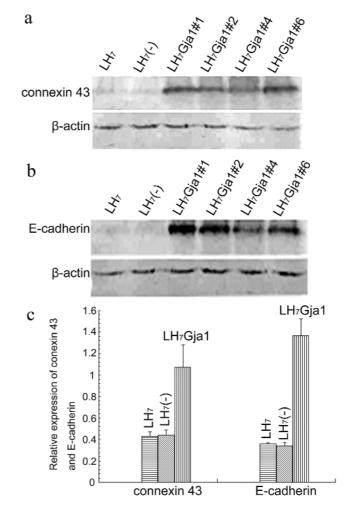
E-cadherin expression is recruited by connexin 43 transfection

After 4 weeks of G418 selection, stable LH_7 -Gja1 cell clones were obtained. These clones were named LH_7 Gja1#1, LH_7 Gja1#2, LH_7 Gja1#4, and LH_7 Gja1#6. As shown in Fig. 2, the immunofluorescence detection of connexin 43 and E-cadherin expression in the LH_7 -Gja#1 clone is obviously stronger than in LH_7 or LH_7 (-) cells. However, localization of both proteins was cytoplasmic rather than at the cell surface. The western blot results show that E-cadherin expression is significantly recruited in LH_7 -Gja1 cells than in LH_7 or LH_7 (-) cells (P < 0.001, Fig. 3).

Overexpression of connexin 43 inhibits cell cycle transition and proliferation of LH_7 -Gja1 cells

The percentage of G1 phase cells in all the LH_7 -Gja1 cell clones was significantly higher than that of LH_7 or LH_7 (-) cells, while the percentages of LH_7 -Gja1 cells

Fig. 3. Connexion 43 and E-cadherin expression in LH₇, LH₇(-) and LH₇-Gja1 cell clones. (a) After connexion 43 transfection, expression was enhanced in LH₇-Gja1 cell clones, LH₇Gja1#1, LH₇Gja1#2, LH₇Gja1#4, and LH₇Gja1#6, compared with LH₇ and LH₇(-) cells. Expression of E-cadherin (b) was greatly enhanced in these cell clones, but not in LH₇ or LH₇(-) cells. β-actin served as internal control. (c) The relative expression was quantified by the ratios of connexion 43 or E-cadherin and β-actin. Expression of connexin 43 and E-cadherin was significantly higher in LH₇-Gja1 cells than in LH₇ or LH₇(-) cells (P < 0.001). The results are the average of three independent experiments.



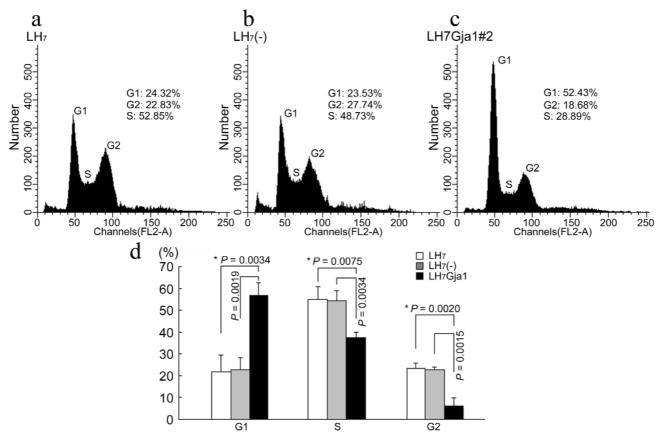


Fig. 4. (a-c) Cell cycle distribution of LH_7 , $LH_7(-)$ and LH_7 -Gja1 clone (LH_7 Gja1#2) assayed by flow cytometry after PI staining. (d) Histogram of LH_7 , $LH_7(-)$ and LH_7 -Gja1 cells. The bars represent the percentages of G1, S or G2 phase cells in LH_7 , $LH_7(-)$ or LH_7 -Gja1 cells. The results are the average of three independent experiments.

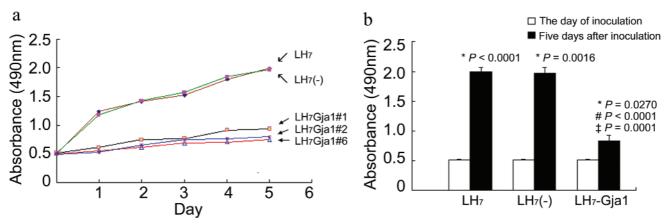


Fig. 5. (a) Growth curves of LH_7 , $LH_7(-)$ and LH_7 -Gja1 clones. The absorbance at 490 nm represents viable cells. (b) Comparison of viable cells among LH_7 , $LH_7(-)$ and LH_7 -Gja1 cells on the day of inoculation and at five days after inoculation. * vs. the day of inoculation, # vs. LH_7 cells and ‡ vs. $LH_7(-)$ cells. The results are the average of three independent experiments.

in S and G2 phases were significantly lower than those of LH_7 or LH_7 (-) cells (P < 0.01, Fig. 4). Further experiments showed that the growth rates of LH_7 -Gja1 cells were also significantly lower than LH_7 or LH_7 (-) cells over one-week period in culture (P < 0.001, n = 3) (Fig. 5).

Discussion

The reduction of connexin 43 or E-cadherin has been reported in several human cancers [7-9]. Our study demonstrates that the concurrent reduction of connexin 43 and E-cadherin is significantly related to differ-

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entiation and progression in 107 lung cancer samples. Furthermore, overexpression of connexin 43 not only restores the level of E-cadherin but also hampers proliferation of human lung cancer cells.

Connexin 43 and E-cadherin expression and their clinical/pathological correlations have not been well documented in human lung cancers [8,18,19]. In a previous study of 24 cases, Jinn et al. reported that connexin 43 was significantly decreased in poorly-differentiated lung cancer. Immunostaining for E-cadherin shows a heterogeneous decrease in expression [19]. Our study not only confirms this early report, but also demonstrates that concurrent reduction of connexin 43 and E-cadherin is significantly related to poor differentiation, advanced TNM staging and lymph node metastasis. These results indicate that concurrent reduction of connexin 43 and E-cadherin may reflect cellular activity in cancer cells and progression of lung cancer.

Earlier reports indicated that potential connections exist between connexins and cadherins, and suggest that cadherins might be a prerequisite for gap junction formation [20,21]. Other reports suggest that E-cadherin regulates gap junction intercellular communication (GJIC) and involves posttranslational regulation of connexin 43 [22,23]. Our results show that connexin 43 and E-cadherin are concurrently expressed in 77.6% of human lung cancer samples (including positive and negative expression of both proteins). Furthermore, our study demonstrates that transfection with connexin 43 gene in LH₇ lung cancer cells increases the expression of E-cadherin. The results revealed that E-cadherin not only regulates expression and function of connexin 43, but it is also induced by connexin 43. The correlation and coordination between E-cadherin and connexin 43 may have important implications for regulating progression of lung cancer.

Connexin 43 suppresses proliferation of cancer cells by inhibiting cell cycle progression [24,25]. Zhang et al. [2,3,26] demonstrated that after transfection of connexin 43, the cell cycle transition from G1 to S phase of tumour cells was inhibited, whether GJIC was present or not. These results further show that connexin 43 increases the synthesis and decreases the degradation of p27, and inhibits the expression of Sphase kinase-associated protein 2 (Skp2). Consistent with these findings, our study shows that although gap junctions and cell adhesions were not clearly observed, the proportion of cells in G1 is significantly increased and the cells in S and G2 phases are significantly decreased in LH₇-Gja1 cells. These results indicate that increased expression of cytoplasmic connexin 43 could still regulate cell cycle and inhibit proliferation of lung cancer cells. Shima et al. [27] also observed similar results in the cytoplasm of connexin 43-transfected basaloid squamous cell carcinoma cells, suggesting that connexin 43 may play a role as a tumour suppressor.

In summary, we have shown that Concurrent reduction of connexin 43 and E-cadherin may reflect cellular activity in cancer cells and progression of lung cancer. Connexin 43 may induce E-cadherin expression and inhibit cell proliferation and progression of lung cancer.

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References

- [1] McLachlan E, Shao Q, Wang HL, Langlois S, Laird DW. Connexins act as tumor suppressors in three-dimensional mammary cell organoids by regulating differentiation and angiogenesis. *Cancer Res.* 2006;66:9886-9894.
- [2] Zhang YW, Kaneda M, Morita I. The gap junction-independent tumor-suppressing effect of connexin 43. *J Biol Chem.* 2003;278:44852-44856.
- [3] Zhang YW, Nakayama K, Nakayama K, Morita I. A novel route for connexin 43 to inhibit cell proliferation: negative regulation of S-phase kinase-associated protein (Skp 2). *Cancer Res.* 2003;63:1623-1630.
- [4] Willecke K, Eiberger J, Degen J, et al. Structural and functional diversity of connexin genes in the mouse and human genome. *Biol Chem.* 2002;383:725-737.
- [5] Okuma A, Kuraoka A, Iida H, Inai T, Wasano K, Shibata Y. Colocalization of connexin 43 and connexin 45 but absence of connexin 40 in granulosa cell gap junctions of rat ovary. J Reprod Fertil. 1996;107:255-264.
- [6] Risley MS, Tan IP, Roy C, Saez JC. Cell-, age- and stage-dependent distribution of connexin43 gap junctions in testes. J Cell Sci. 1992;103:81-96.
- [7] Kato Y, Hirano T, Yoshida K, et al. Frequent loss of E-cadherin and/or catenins in intrabronchial lesions during carcinogenesis of the bronchial epithelium. *Lung Cancer*. 2005;48: 323-330.
- [8] Chen JT, Cheng YW, Chou MC, et al. The correlation between aberrant connexin 43 mRNA expression induced by promoter methylation and nodal micrometastasis in non-small cell lung cancer. Clin Cancer Res. 2003;9:4200-4204.
- [9] Brehm R, Rüttinger C, Fischer P, et al. Transition from preinvasive carcinoma in situ to seminoma is accompanied by a reduction of connexin 43 expression in Sertoli cells and germ cells. *Neoplasia*. 2006;8:499-509.
- [10] Torres LN, Matera JM, Vasconcellos CH, Avanzo JL, Hernandez-Blazquez FJ, Dagli ML. Expression of connexins 26 and 43 in canine hyperplastic and neoplastic mammary glands. *Vet Pathol.* 2006;42:633-641.
- [11] Sun WH, Liu HM, Li YJ, Ji XR, Liang DP. A study of the relationship between the expression of connexin43, E-cadherin and biological behaviors of human laryngeal cancer. *Zhonghua Er Bi Yan Hou Ke Za Zhi.* 2004;39:293-297.
- [12] Hernandez-Blazquez FJ, Joazeiro PP, Omori Y, Yamasaki H. Control of intracellular movement of connexins by E-cadherin in murine skin papilloma cells. *Exp Cell Res*. 2001;270:235-247.
- [13] Travis WD, Brambilla E, Muller-Hermelink HK, Harris CC (eds). Pathology and genetics of tumors of the lung, pleura, thymus and heart. World Health Organization Classification of Tumors. Lyon: IARC Press; 2004.

- [14] Mountain CF. The international system for staging lung cancer. Semin Surg Oncol. 2000;18:106-115.
- [15] Watanabe Y. TNM classification for lung cancer. Ann Thorac Cardiovasc Surg. 2003;9:343-350.
- [16] Nemeth L, Rolle U, Puri P. Altered cytoskeleton in smooth muscle of aganglionic bowel. Arch Pathol Lab Med. 2002; 126:692-696.
- [17] Zhu W, Zheng J, Fang W. Isolation and characterization of human lung cancer cell subline with different metastatic potential. Zhonghua Bing Li Xue Za Zhi. 1995;24:136-138.
- [18] Zhang ZQ, Zhang W, Wang NQ, Bani-Yaghoub M, Lin ZX, Naus CC. Suppression of tumorigenicity of human lung carcinoma cells after transfection with connexin43. *Carcinogen*esis. 1998;19:1889-1894.
- [19] Jinn Y, Ichioka M, Marumo F. Expression of connexin32 and connexin43 gap junction proteins and E-cadherin in human lung cancer. *Cancer Lett.* 1998;127:161-169.
- [20] Kostin S, Hein S, Bauer EP, Schaper J. Spatiotemporal development and distribution of intercellular junctions in adult rat cardiomyocytes in culture. Circ Res. 1999;85:154-167.
- [21] Ko K, Arora P, Lee W, McCulloch C. Biochemical and functional characterization of intercellular adhesion and gap junctions in fibroblasts. *Am J Physiol Cell Physiol*. 2000;279: C147-157.

- [22] Yano T, Yamasaki H. Regulation of cellular invasion and matrix metalloproteinase activity in HepG2 cell by connexin 26 transfection. *Mol Carcinog*. 2001;31:101-109.
- [23] Jongen WM, Fitzgerald DJ, Asamoto M, et al. Regulation of connexin 43-mediated gap junctional intercellular communication by Ca²⁺ in mouse epidermal cells is controlled by E-cadherin. *J Cell Biol*. 1991;114:545-555.
- [24] Huang RP, Fan Y, Hossain MZ, Peng A, Zeng ZL, Boynton AL. Reversion of the neoplastic phenotype of human glioblastoma cells by connexin 43 (cx43). *Cancer Res.* 1998; 58:5089-5096.
- [25] Lee HJ, Lee IK, Seul KH, Rhee SK. Growth inhibition by connexin26 expression in cultured rodent tumor cells. *Mol Cells*. 2002;14:136-142.
- [26] Zhang YW, Morita I, Ikeda M, Ma KW, Murota S. Connexin43 suppresses proliferation of osteosarcoma U2OS cells through post-transcriptional regulation of p27. *Oncogene*. 2001;20:4138-4149.
- [27] Shima K, Muramatsu T, Abiko Y, Yamaoka Y, Sasaki H, Shimono M. Connexin 43 transfection in basaloid squamous cell carcinoma cells. *Oncol Rep.* 2006;16:285-288.

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