Alkaline phosphatase, cytokeratin 7, cytokeratin 8 in the diagnosis of primary lung adenocarcinoma from 148 pleura fluids specimens

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Abstract: Adenocarcinomas are the most common cause of malignancy in pleura fluids. Usual primary sites include the lung, breast, gastrointestinal tract, and genitourinary tracts. Predicting the site of origin of an adenocarcinoma can be difficult due to overlapping morphologic characteristics. We investigated the use of alkaline phosphatase (AP), Cytokeratin7 (CK7) Cytokeratin8 (CK8) to distinguish adenocarcinomas of lung in 148 body cavity fluid samples. Overall results for primary lung adenocarcinomas, demonstrated CK8 reactivity in 106 (72%) of 148 cases. 95 primary lung carcinoma samples (65%) were positive for CK7. AP was expressed in 81% of primary lung adenocarcinomas. Positive immunoreactivity for AP was characterized by a red, diffusely apical cytoplasmic staining in tumor cells that occurred singly or in groups. There was a significant difference between AP, CK 7 and CK 8 expressions in primary lung adenocarcinomas (P=0.02; Chi-squared test). The sensitivity of AP, CK8, CK7 as a marker for primary lung adenocarcinomas were 82%, 72%, 64%, respectively. Thus the AP positive staining largely confirmed the cytologic diagnosis of lung adenocarcinoma.

Key words: lung adenocarcinoma, alkaline phosphatase, cytokeratin 7, cytokeratin 8, immunocytochemistry, body cavity fluids

Introduction

The incidence of lung cancer in the United States has been steadily increasing in recent years, and lung cancer now ranks as the primary cause of death from cancer in American men [1]. Non-small cell carcinomas account for about 85% of all lung cancers. About 60% of lung cancers are adenocarcinomas, and about 26% are squamous cell carcinomas [2]. The proportion of adenocarcinomas has been increasing recently, and the percentage of woman with adenocarcinoma has also been rising [3].

Adenocarcinomas are the most common epithelial malignancies found in body cavity fluids. Common primary sites include the lung, breast, gastrointestinal tract, and genitourinary tract. Identification of the specific site of origin may carry important prognostic as well as therapeutic implications. However, determination of the primary site of adenocarcinoma based on cytomorphology alone can be a challenging task. Tissue specific immunohistochemical markers offer an attractive means for confirming metastatic disease in patients with known primary tumors and may assist in identifying the primary sites in patients with adenocarcinomas of unknown origin.

Blobel et al. [4] showed that alveolar cells of human lung contained cytokeratin (CK) polypeptides typical of simple epithelia (CK7, 8, 18, and 19). Basal cells of the bronchial epithelium, on the other hand, contained CK5 and small amounts of CK6. Consequently, it has been also found "simple-epithelium-type" cytokeratins in all adenocarcinomas and later also in squamous cell carcinomas [5]. CK8 is the most widely expressed cytokeratin in various epithelial cells and cancer cells among at least 21 related cytokeratins [6]. One study has demonstrated that CK8 is found in the serum of a subgroup of patients with non-small cell
lung carcinomas [7]. CK7, a neutral-basic type II cytokeratin found in adenocarcinomas of breast and lung, among others [8–10].

Alkaline phosphatase (AP) isoenzymes are widely distributed in various organs, and activity and/or AP identification in bronchial fluid is a marker of lung carcinoma and metastases [11–13]. Human APs are reportedly encoded by four different genes: a gene for the liver/bone/kidney isozyme (tissue-nonspecific type, TNAP), a gene for the adult intestinal isozyme (IAP) a gene for the placental (PLAP), a gene for the germ cell isozyme (placental-like AP or GCAP [14]. TNAP is the predominant AP isozyme in the lung, where it is largely produced by type II pneumocytes [15], but trace amounts of PLAP are also expressed by type I pneumocytes [16].

Muessngh et al. [17] have found that AP is elevated in 23% of 286 patients suffering from various neoplasms. AP was elevated most commonly in cases of colorectal cancer (54%), ovarian cancer (44%) and lung cancer (40%). It seems that expression of certain cytokeratins (CK7, CK8) and AP may be useful in the diagnosis of primary lung adenocarcinomas. This study was undertaken to evaluate the clinical utility of CK7, CK8 and AP in the diagnosis of primary lung adenocarcinoma in body cavity fluids specimens.

Materials and methods

Cases. The cytological files of the Hannover Cytopathology Institute were reviewed between 6 January 2005 and 21 February 2007. The 148 pleural fluids were collected from 148 patients who were diagnosed with adenocarcinoma. Medical records were reviewed to verify that the cases were of primary lung origin and did not represent metastatic disease from primary tumors at other sites. The selection of patients was restricted to include those in whom the primary site of origin for the carcinoma was confirmed based on clinical findings (n=148). Slides that were stained previously by routine Giemsa methods were retrieved for each patient. The diagnosis of adenocarcinoma based on cytomorphology was confirmed. Classic cytologic features of malignancy included three-dimensional aggregates comprised of cells with increased nuclear-to-cytoplasmic ratios, irregular nuclear membranes, course chromatin, large irregular nucleoli and finely vacuolated cytoplasm. The sensitivity of CK7, CK8 and AP in body cavity fluid samples for primary lung adenocarcinomas was calculated.

Immunocytochemistry procedure. Cytologic smears were prepared by standard cytologic method. The slides were hydrated in decreasing ethanol solutions. Endogenous peroxidase was blocked with hydrogen peroxide for 3 minutes. The slides were rinsed in water, were incubated with the biotin blocking system (Dako) prior to application of the primary antibody. The CK7 and CK8 monoclonal antibodies were used at a dilution of 1:100 and incubated with the samples for 1 hour. The slides were then rinsed in buffer and incubated for 25 minutes with the linking solution (LSAB+ kit; Dako; biotinylated antimouse, antirabbit, and antigoat). This was followed by a rinse in buffer and incubation with streptavidin peroxidase for 25 minutes. After rinsing in buffer, the slides were submerged in AEC for 5 minutes. The slides were counterstained with Mayer’s hematoxilen. The slides were then dehydrated through gradient alcohols, cleared in xylene, and coverslipped. Only cytoplasmic staining was regarded as a positive result. The intensity of staining was graded on a 0 (absent), 1+ (weak), 2+ (moderate), and 3+ (strong). All material was evaluated blindly by two observers. Statistical analysis was performed using the chi-squared test and the orthogonal test. The chi-square test was used to determine statistical significance.

Enzyme cytochemistry. For detection of AP activity, 17.5 mg naptihyle phosphate sodium salt monohydrate were dissolved in 35 ml 0.1 mole veronalnatrium buffer (pH=9.4) and mixed with 35 mg varianmine blue B salt. The mixture was freshly prepared before use. We incubated cells for 60 minutes at 4°C and counterstained with Mayer’s hemalution solution for 4 min. Then the slides were mounted with glycerol gelatin. Cytoplasmic staining of tumor cell was considered positive.

Results

The results of AP, CK7, CK8 immunostaining of lung adenocarcinomas in 148 pleura fluids are shown in Table 1. AP was expressed in 81% of primary lung adenocarcinomas. Positive immunoreactivity for AP was characterized by a red diffusely cytoplasmic staining in tumor cells that occurred singly or in groups. AP showed cytoplasmic staining in adenocarcinoma cells (Fig. 1). 27 of 148 fluids from adenocarcinoma of lung origin were negative for AP. 95 (65%) primary lung carcinoma samples were positive for CK7 (Fig.2a, b).

Overall results for primary lung adenocarcinomas demonstrated CK8 reactivity in 106 (72%) of 148 cases (Fig.3). Staining intensity was moderate in most primary (1+, 31 cases; 2+, 52 cases; 3+, 23 cases) lung adenocarcinomas.

Table 1 has showed the staining patterns of primary lung adenocarcinomas from pleural effusions for AP, CK7 and CK8. For all primary lung adenocarcinomas studied, 121 (81%) of cases exhibited AP, 95 (65%) of cases exhibited CK7, 106 (72%) of cases exhibited CK8 immunostaining.

There was a significant difference between AP, CK7 and CK8 expressions in primary lung adenocarcinomas (P=0.02; Chi-squared test). The statistical values are given in Table 2. The sensitivity of AP, CK7 and CK8 as a marker for primary lung adenocarcinoma were 82%, 64%, and 72% respectively. AP had the highest sensitivity of the primary lung adenocarcinoma studied, with a sensitivity of 82%.

Discussion

Adenocarcinoma is the most common cause of malignancy in body cavity fluids. Usual primary sites include the lung, breast, gastrointestinal tract, and genitourinary tracts. Predicting the site of origin of an adenocarcinoma can be difficult due to overlapping morphologic characteristics. Lung is a common primary
We investigated the use of AP, CK7, CK8 to distinguish adenocarcinomas of lung in 148 pleura fluids. Organ specific markers using immunohistochemistry offer an attractive means for subclassifying adenocarcinomas according to primary site. CK7 and CK8 are tissue specific protein expressed selectively in the epithelial cells of the lung [21].

Cytokeratins, belonging to the intermediate filament (IF) protein family, are particularly useful tools in oncology diagnostics. At present, more than 20 different cytokeratins have been identified, of which

<table>
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<th>Tumor Markers</th>
<th>n</th>
<th>Negative n (%)</th>
<th>Positive n (%)</th>
<th>1+ (%)</th>
<th>2+ (%)</th>
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<tr>
<td>Alkaline Phosphatase</td>
<td>148</td>
<td>27 (18)</td>
<td>121 (81)</td>
<td>31 (21)</td>
<td>58 (39)</td>
<td>32 (21)</td>
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<tr>
<td>Cytokeratin 7</td>
<td>148</td>
<td>53 (35)</td>
<td>95 (65)</td>
<td>26 (18)</td>
<td>36 (24)</td>
<td>33 (22)</td>
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<tr>
<td>Cytokeratin 8</td>
<td>148</td>
<td>42 (28)</td>
<td>106 (72)</td>
<td>31 (20)</td>
<td>52 (35)</td>
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Figure 1. Cytochemical staining of primary lung adenocarcinoma with alkaline phosphatase. Couterstained with hematoxylin (pleura, original magnification x 600)

Figures 2a,b. Two primary lung adenocarcinoma samples show intense cytoplasmic staining with cytokeratin 7 ((a) Pleura, immunoperoxidase-hematoxylin, x200) and (b) Pleura, immunoperoxidase-hematoxylin, x400)
cytokeratins 8, 18, and 19 are the most abundant in simple epithelial cells. Upon release from proliferating or apoptotic cells, cytokeratins provide useful markers for epithelial malignancies. Previous examination has demonstrated CK 7 and CK 8 positivity in the vast majority of lung adenocarcinomas studied [7,22]. Using commercially available monoclonal antibody for CK8, we demonstrated a sensitivity of 72% for lung adenocarcinomas. 95 primary lung carcinoma samples (65%) were positive for CK7. The sensitivity of CK7 as a marker for primary lung adenocarcinomas was 65%.

Human alkaline phosphatase isoenzymes have been classified electrophoretically and immunohistochemically into four major groups: hepatic, intestinal, bone and placental [23]. Placental alkaline phosphatase (PLAP) levels are elevated during pregnancy and in cigarette smokers. However, in non-pregnant women and non-smokers, PLAP activity is very low, compromising less than 1% of the total alkaline phosphatase activity [17,24].

Following the discovery by Fishman et al. [14] that PLAP elevation is associated with bronchogenic carcinoma, many other studies have confirmed that this isoenzyme is ectopically produced by various neoplasms [25-27]. Muensh et al. [17] have found, that PLAP is elevated in 23% of 286 patients suffering from various neoplasms. PLAP was elevated most commonly in cases of colorectal cancer (54%), ovarian cancer (44%) and lung cancer (40%). Other studies have confirmed raised serum concentrations of this alkaline phosphatase isoenzyme in 15-64% of patients with ovarian cancer [28-31]. One study showed that almost all ovarian cancer tissues contained PLAP to some extend, however there was a low frequency of detectable serum levels of PLAP, especially in the early stages of the disease [29]. In this study, AP was expressed in 81% of primary lung adenocarcinomas. Positive immunoreactivity for AP was characterized by a red diffusely cytoplasmic staining in tumor cells that occurred singly or in groups. The specificity and positive predictive values of the AP antibody for the adenocarcinomas of lung origin in our series were both 81%.

Staining for AP activity is a rapid and simple test, which does not require any special equipment. When used carefully and in conjunction with additional testing such as cytology, detection of AP activity appears to be a useful tool in the diagnosis of primary lung adenocarcinoma. Generally, the cytologic and clinical features of the tumors can assist in diagnosis. In addition, detection of AP activity only improves the ability of the clinician and pathologist to appropriately diagnose primary adenocarcinoma of lung.

This application of the AP marker onto cytological slides of lung adenocarcinoma can be very helpful in daily clinical cytology practice. The staining technique is simple, and the AP yields highly sensitive results. A positive reaction for AP may serve to confirm the spread of a known primary lung tumor, establish the recurrence of a previously treated adenocarcinoma of the lung, or identify the lung as a possible source of malignancy in a patient with an unknown primary site. In this study, AP was expressed in 81% of primary lung adenocarcinomas. Positive immunoreactivity for AP was characterized by a red diffusely cytoplasmic staining in tumor cells that occurred singly or in groups. The specificity and positive predictive values of the AP antibody for the adenocarcinomas of lung origin in our series were both 81%. 27 of 148 fluids from adenocarcinoma of lung origin were negative for AP.

The histologic distinction between primary pulmonary adenocarcinomas and adenocarcinomas metastases to the lung can be a difficult task, with significant clinical ramifications. In this regard, immunohistochemical evaluation of CK7 and CK8 and AP expression patterns has been a useful method of accurately classifying these entities [32-36].

Our findings have confirmed the results of other studies with regard to the high frequency and intensity of CK7, CK8 and AP expressions in lung adenocarcin-
nomas [37-40]. The sensitivity of AP, CK8, CK7 as a marker for primary lung adenocarcinomas were 81%, 72%, 65%, respectively. Thus the AP positive staining largely confirmed the cytologic diagnosis of lung adenocarcinoma.

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Submitted: 5 May 2008
Accepted after reviews: 9 November 2008