The TRAF2 and TRAF6 expression in myomas and myometrium of women in reproduction and perimenopausal age

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Abstract: Uterine myomas represent one of the most common female diseases. Uterine myomas or fibromas are benign, hormone-responding tumours of, respectively, smooth muscles and fibroblasts and their aetiology induces a significant interest. In myomas the presence of aromatase was detected and, in addition, oestrogen was found to be synthesized in myoma cells. The studies were performed on myoma patients of generative age and those in peri-menopausal age. Expression of TRAF2 and TRAF6 proteins was examined using immunohistochemistry and Western blot approach in small and large uterine myomas isolated from women of various age. In addition, the evaluation was conducted at the periphery of every myoma. We indicated that the level of both tested proteins in myomas is higher than in control. TRAF2 level in myometrium was lower than in myomas but higher than in control. In the case of TRAF6 those changes were ambiguous. Age didn't have influence the level of expression in both tested TRAF in studied structures.

Key words: TRAF2, TRAF6, leiomyomas, small and large leiomyoma, women

Introduction

Despite the high frequency of leiomyoma manifestation scanty information is available on their development and growth. On the basis of biochemical and cytogenetic studies every leiomyoma is thought to represent a product of clonal expansion of an individual myocyte [1-3]. Just as it is the case with other tumours, probably several genetically sensitive sites exist, which lead to dysregulation of a smooth muscle cell, resulting in its transformation to the common phenotype of a leiomyoma cell.

Studies performed till now demonstrated that an immune response is linked to activation of NF-κB under effect of IL-1 and TNF-α [4-8]. The two cytokines exert their effect due to activation of a kinase which induces NF-κB and, then, kinase IKK, which directly affects activity of NF-κB transcription factor. The phosphorylated, C-terminal domain of IL-1 receptor-linked kinase interacts with TRAF6 protein [9-11]. Transmission of IL-1 signal may include role of TRAF6 and/or TRAF2 [12-14]. TRAF-6 activates NIK kinase but it should be stressed that NIK is a common mediator of signalling pathways originating from multiple cytokine receptors [15-17].

TRAF (TNF receptor-associated factor) family represents a family of cytoplasmic proteins capable both of negative control of apoptosis and of induction of genes which promote survival. They were shown to serve as adaptor proteins for a broad range of surface receptors, to play an important role not only in apoptosis but also in response to stress. The proteins may directly interact with intracellular domains of surface
receptors, including TNF-α (TRAF2) or IL-1β (TRAF6) receptors.

TRAF proteins may serve for modulation of receptors capacities to mobilize various signalling pathways, which lead to phosphorylation and activation of protein kinases and, then, to activation of transcription factors of Rel and AP-1 family.

In our earlier study we evaluated expression of aromatase in uterine myomas [18]. Expression of the aromatase depends on several factors, including TRAF. Even if neither TRAF2 nor TRAF6 directly affect expression of aromatase, their indirect effect is evident. The two proteins are known to directly stimulate activity of NF-κB. This nuclear factor may bind one of the promoters, such as I4, which mobilizes aromatase synthesis pathway. Expression of aromatase may also be stimulated by PII promoter. While the promoter is activated by PGE2, levels of the prostaglandin are directly related to activity of COX-2, and activity of the cyclooxygenase is stimulated, among others, by NF-κB.

In this aim the attempt will be undertaken to demonstrate whether there exist differences in the amount and distribution of TRAF2 and TRAF6 proteins in myomas of various size, isolated from women of various age, as compared to an unchanged tissue. The evaluation will be made using Western blot technique and quantitative analysis of immunohistochemical tests conducted on tissues of a normal uterus and a myomatous uterus.

**Materials and methods**

**Human material.** Recruitment of patients, clinical studies and hormonal tests will be conducted in the Chair and Department of Gynaecological Endocrinology, Medical University of Silesia in Katowice while enzymatic and protein studies will be executed in the Department of Proteomics.

The studies were conducted on 40 patients with myomas, at the reproductive age (below 45th year of age, FSH<30 mIU/ml; Samples were taken in follicular phase of menstrual cycle) and 40 patients with myomas in the perimenopausal age (45-55 years of life, FSH>30 mIU/ml). Inclusion criteria will involve myoma detected by USG, qualification of the patient to hysterectomy, informed consent to the planned studies. The exclusion criteria will include: therapy with any drugs, including hormonal drugs in the minimum of 3 months before inclusion to the studies, neoplastic disease, endometrial hypertrophy, metabolic and systemic disturbances, nicotinism.

Myometrial samples, for use as health controls were taken from 10 young women (<40 years old) undergoing hysterectomies for ovary tumors and 10 perimenopausal age women (>52 years old) undergoing hysterectomies for uterine prolapse. In these studies we used only material from uteruses with one large myoma or one large and few small myomas. The material for studies was excised from uterus taken during surgery, and fixed histopathologically. Samples of leiomyoma 1 × 1 cm and samples of a similar size from health myometrium at the distance of at least 4 cm each were taken. Before isolation of myometrium samples the patients were informed on the aim of the studies. The planned investigative procedures were approved by the Medical Bioethical Commission.

**Immunohistochemistry.** Tissues were fixed in 4% buffered formalin, dehydrated and then embedded in paraffin. Section (5 μm) were mounted on silane-coated slides. Samples were de-waxed, and rehydrated. To unmask the antigen, sections were boiled in 0,01 M citrate buffer, pH 6, in a microwave oven for 10 min at 800 W. Endogenous peroxidase activity was quenched using 1.5% (v/v) solution H2O2 in methanol for 10 min and then washed in PBS-Tween 20 (0,05% v/v) and blocked with 1% BSA for 60 min. The sections were further blocked with avidin-biotin-blocking solution according to the manufacturer's instructions. After this the slides were incubated with rabbit anti-TRAF2 polyclonal antibody (Abcam) or rabbit anti-TRAF6 monoclonal antibody (Abcam) in a humidified chamber for 22 h at 4°C. After washing in PBS-Tween 20 sections were incubated with biotinylated goat anti-rabbit immunoglobulins (Vector Laboratories Inc.) for 30 min, and were incubated with avidin-biotinylated peroxidase complex (Vector) for 30 min. The bound antibodies were visualised with diaminobenzidine (DAB) and H2O2 in PBS, pH 7.5 according to supplier's instructions (Vector). Finally, the tissues were stained with Gill's hematoxylin, dehydrated, and cover-slipped. For negative controls rabbit IgG were used.

In each positively stained cell, the intensity of staining was measured as the optical density of the reaction product, with the program KS 300 VIDAS video image analyzer served by IBAS 2.5 system and a Panasonic digital camera. For each analyzed area, 173 × 130 μm average optical density per unit area was calculated. Finally, the arithmetic mean and standard deviation were calculated.

**Western blotting.** Tissue samples (~500 mg) were homogenized in 0.5 % Triton X-100 lysis buffer containing 20 mM Tris-HCl (pH 7.5), 150 mM NaCl, 10 mM NaF, 2 mM dithiothreitol (DTT), 1 mM sodium orthovanadate, 1 mM phenylmethylsulfonyl fluoride (PMSF) and 20 M aprotinin. Homogenates were centrifuged at 10,000 × g for 10 min. Concentration of protein was determined by Bradford assay. For immunoblotting samples (25 μg of protein) were separated by 12% SDS-polyacrylamide gels, and transferred to nitrocellulose membranes. After blocking with 5% skim milk in PBS-T (PBS with 0.1% Tween 20) for 1h, the membranes were probed with TRAF2 or TRAF6 antibodies (Abcam) overnight. Primary antibodies were detected with

**Characteristics of studied groups**

Group 1. Myometrium of menstruating women, in whom hysterectomy was performed for reasons other than uterine leiomyomas; the 10 patients represented the lesion-free control.

Group 2. Leiomyomas of <3 cm in diameter, sampled from uteri of menstruating women (n=20 women).

Group 3. Myometrium of menstruating women, sampled at the distance of not less than 4 cm from margins of a small leiomyoma (n=20 women; autologous control).

Group 4. Leiomyomas of >5 cm in diameter, sampled from uteri of menstruating women (n=20 women).

Group 5. Myometrium of menstruating women, sampled at the distance of not less than 4 cm from margins of a large leiomyoma (n=20 women; autologous control).

Group 6. Myometrium of non-menstruating women, in whom hysterectomy was performed for reasons different than uterine leiomyomas (n=10 women; lesion-free control).

Group 7. Leiomyomas of <3 cm in diameter, sampled from uteri of non-menstruating women (n=20 women).

Group 8. Myometrium of non-menstruating women, sampled at the distance of not less than 4 cm from margins of a small leiomyoma (n=20; autologous control).

Group 9. Leiomyomas of >5 cm in diameter, sampled from uteri of non-menstruating women, (n= 20 women).

Group 10. Myometrium of non-menstruating women, sampled at the distance of not less than 4 cm from margins of a large leiomyoma (n=20 women, autologous control).
Results

Immunohistochemical studies

Optical density of cells with expression of an evaluated protein reflects concentration of the immunocytochemical reaction product in evaluated uterine structures.

TRAF2

Young women

Evaluating levels of TRAF2 in studied structures (Fig. 1), optical density of the immucytochemical reaction products in small myomas of young women was found to be higher, amounting to around 185% of the control level (Fig. 2). The same analysis targeted at vicinity of the myomas demonstrated also higher expression of the protein, which in this case reached 152% of the control level. Analysis of the data showed that expression of TRAF2 at the periphery of myomas amounted to 82% of the value noted in the myoma.

Comparing optical density in large myomas manifesting TRAF2 expression with the control level the demonstrated expression of the protein was found to be equal to 196% of the control level. Similarly to small myomas, TRAF2 level measured at the periphery was slightly lower than that in the myomas, amounting to 182% of the control level. This corresponded to 93% of TRAF2 expression level noted in large myomas.

In women of reproductive age, TRAF2 expression level in large uterine myomas resembled the value noted in small myomas. Evaluation of the protein expression at the direct periphery of the large myomas demonstrated a similar expression level to that noted at the periphery of small myomas.

Women at perimenopausal age

Evaluation of TRAF2 level in studied structures (Fig. 1) showed that in women of perimenopausal age optical density of the reaction product in small myomas was increased to almost 165% of the control level (Fig. 2). Analysis of myoma periphery demonstrated the level higher than in the control (150% of the control level).

Comparing optical density of large myomas with TRAF2 expression as compared to the control, the former was found to manifest higher expression of the protein, amounting to almost 175% of the control level. In contrast to small myomas, activity of TRAF2 measured at the periphery was lower than inside the myoma and amounted to around 147% of the control level.

Analysis of TRAF2 expression in uterine myomas of women of post-reproductive age disclosed that in large myomas expression of TRAF2 was comparable to the expression documented in small myomas. At the direct vicinity of large myomas the expression resembled that observed around small myomas.

Effect of age

Evaluating content of TRAF2 reaction product, a level of the protein in control samples obtained from young women was slightly lower than in women of perimenopausal age women. A similar observation was made upon quantitation of the protein expression in small myomas and at their periphery in women of the two age groups.

Analysis of the quantitative data related to large myomas demonstrated that in women of perimenopausal age TRAF2 expression level was lower and amounted to 89% of the level detected in young women. Also at the periphery of large myomas the level was distinct in the two age groups: in young women it amounted to 124% of the value observed in the older group.

TRAF6

Young women

Upon evaluation of TRAF6 levels in the studied structures (Fig. 3), in young women optical density of the immunochemistry reaction product was increased in small myomas, reaching around 235% of the control level (Fig. 4). The same analysis targeted at the myoma periphery also in this case demonstrated expression of the protein higher than in the control, reaching the level of 185%. Analysis of the data demonstrated that expression of TRAF6 at the periphery of myoma amounted to 78% of the value noted in the myoma.

Comparing optical density of large myomas with TRAF6 expression with the control level, the protein expression was found to represent 288% of the control value. Similarly to small myomas, TRAF6 expression level measured at the periphery was slightly lower than that in myomas and corresponded to 166% of the con-
control level or 58% of the expression of TRAF6 noted in large myomas.

Quantitative analysis of TRAF6 expression level in uterine myomas of women in reproductive age, in large myomas the expression level was higher than that observed in small myomas, amounting to 123% of the protein expression level in small myomas. In direct vicinity of large myomas the expression level was lower than the level noted around small myomas, corresponding to 90% of the latter level.

**Women of perimenopausal age**

Evaluation of TRAF6 expression level in the studied structures (Fig. 3) disclosed that in women of perimenopausal age optical density of the reaction product increased in small myomas to almost 210% of the control level (Fig. 4). At periphery of the myomas expression of the protein also was higher than in the control, reaching the level equivalent to 195% of the control. Analysis of the data showed that TRAF6 expression at periphery of small myomas reached 95% of the value observed in the myomas.

Evaluation of optical density of large myomas with TRAF6 expression showed that the expression was higher, corresponding to 240% of the control level. Similarly to small myomas TRAF6 activity measured at periphery was slightly lower than that in the myomas, amounting to around 222% of the control value.

Analysis of TRAF6 expression in uterine myomas of women in post-reproductive age demonstrated that in large myomas TRAF6 expression was higher as compared to the expression observed in small myomas. In a direct vicinity of large myomas it was found to be slightly higher than the level noted in vicinity of small myomas.

**Effect of age**

Comparison of the product content in the reaction for TRAF6 showed levels of the protein in control samples originating from young women was slightly higher than the level observed in perimenopausal women. A similar observation was made during quantitative evaluation of the protein expression in small myomas. Periphery of small myomas demonstrated similar values of TRAF6 expression in the two age groups of women.

Analysis the quantitative data related to large myomas demonstrated that TRAF6 expression level in women of perimenopausal age was lower, amounting to 83% of the level demonstrated in young women. The reaction product measured in vicinity of large myomas was also distinct in the two age groups: in young women it amounted to 75% of the value observed in the older group.

**Western blot analysis**

**TRAF2**

Upon comparison of TRAF2 expression in myometrium of women of various age the samples originating from women in perimenopausal age were found to be slightly higher, although the difference failed to reach statistical significance.

For a better and a more legible evaluation of changes in TRAF2 expression (and the remaining proteins evaluated by Western blot analysis) a quantitative analysis was performed exclusively in the same age groups, in every case accepting the expression value noted in a pure control as 100%.

Expression of TRAF2 measured at the periphery of myomas originating from uteri of young females was higher than that in the control, amounting to 115% and 120% (Fig. 5; Table 1) in, respectively small and large myomas although the differences proved insignificant. The TRAF2 expression level detected in small myomas isolated from uteri of young women was clearly higher than in the control and amounted to 135%, and the difference was significant. Even higher was TRAF2 expression level detected in large myomas, in which the expression level reached 140% of the control level.

Upon analysis of differences in TRAF2 expression between myomas and their direct vicinity expression of the protein in small myomas was significantly higher than that in the surrounding myometrium. A similar difference in the level of TRAF2 expression was detected in cases of large myomas.

Analysis of differences in TRAF2 expression in uteri of perimenopausal women demonstrated similar but slightly more accentuated differences: expression of the protein measured at the periphery of myomas was also higher than that in the control. The levels amounted to 125% and 120% in small and large myomas, respectively (Fig. 5; Table 1), but also in this case the differences proved to be insignificant.

Expression level of the evaluated TRAF2 protein in small myomas isolated from uteri of perimenopausal women was higher than that in the control and reached 145%, and the difference was significant. In large myomas of the age group TRAF2 expression level was even higher, amounting to 160% of the control level.

Evaluating TRAF2 expression level in myomas and in the surrounding myometrium it was shown that in the older age group expression of TRAF2 in small myomas was significantly higher than that in the surrounding myometrium. An even higher difference, of 135% order was disclosed during analysis of differences in TRAF-2 expression levels in large myomas.
Fig. 1. Immunohistochemical staining of uterini samples from reproductive (A-E) and perimenopausal age women (F-J) with rabbit anti-TRAF2 polyclonal antibodies. A and F – healthy myometrium, B and G – small myomas, C and H – periphery of small myomas, D and I – large myomas, E and J – periphery of large myomas (original magnification ×200).

Fig. 2. Quantitative evaluation of TRAF2 expression in immunohistochemical staining. Date shown represent mean ± SD. Differences in TRAF2 expression: * – in reproductive age women and perimenopausal age women; a – in small myomas and healthy myometrium; b – in periphery of small myomas and healthy myometrium; c – in large myomas and healthy myometrium; d – in periphery of large myomas and healthy myometrium; e – in small myomas and its periphery; f – in small myomas and its periphery, are statistically significant.
Fig. 3. Immunohistochemical staining of uterini samples from reproductive (A-E) and perimenopausal age women (F-J) with rabbit anti-TRAF6 polyclonal antibodies. A and F – healthy myometrium, B and G – small myomas, C and H – periphery of small myomas, D and I – large myomas, E and J – periphery of large myomas (original magnification ×200).

Fig. 4. Quantitative evaluation of TRAF6 expression in immunohistochemical staining. Date shown represent mean ± SD. Differences in TRAF6 expression: * – in reproductive age women and perimenopausal age women; a – in small myomas and healthy myometrium; b – in periphery of small myomas and healthy myometrium; c – in large myomas and healthy myometrium; d – in periphery of large myomas and healthy myometrium; e – in small myomas and its periphery; f – in small myomas and its periphery, are statistically significant.
Comparison of TRAF6 expression levels in myometrium of women of various age groups showed that in samples originating from perimenopausal women it was even higher, reaching 120% of the value noted in the younger group. However, the difference proved to be insignificant.

Expression of TRAF6 measured at the periphery of myomas originating from uteri of young women resembled that in the control, amounting to, at most, 110% of the control value. TRAF6 expression level in small myomas isolated from uteri of perimenopausal women was higher than that in the control and amounted to 125%, but the difference was insignificant. Even higher level of TRAF6 expression was seen in large myomas, in which the level of the discussed TRAF protein reached 135% of the control level (Fig. 6; Table 1).

In analysis of TRAF6 levels in myomas and in the surrounding myometrium it was demonstrated that in the group of young women expression of TRAF6 in small myomas was slightly, but insignificantly higher than in the surrounding myometrium. A more pronounced difference, of 125% order, was detected during analysis differences in TRAF6 expression levels in large myomas, as compared to those in their direct neighbourhood.

Analysis of TRAF6 expression in uteri of perimenopausal women demonstrated more pronounced differences: expression of the protein measured at the periphery of myomas was higher than that in the control, amounting to 135% and 125% in small and large myomas, respectively (Fig. 6; Table 1). The difference was significant for periphery of small myomas.

The detected expression level of the evaluated TRAF6 protein in small myomas isolated from uteri of perimenopausal women was higher than that in the control, amounting to 135%, and the difference was significant. In large myomas of the age group the detected TRAF6 expression level was even slightly higher, reaching 145% of the control value.

Evaluation of TRAF6 expression level in myomas and the surrounding myometrium demonstrated that in the older age group expression of TRAF6 in small myomas was the same as in the surrounding myometrium. On the other hand, a difference of 115% order was noted upon analysis of TRAF6 expression in large myomas. However, the difference was insignificant.

**Discussion**

Uterine leiomyomas are the most frequent tumours of female genital tract. More than 70% of preparations, resulting from hysterectomy performed for various reasons, demonstrated presence of uterine leiomyomas [19-21].

Certain traits of TRAF proteins suggest that they act as cytoplasmic adapters, which may support transmission of an intra-cellular signal due to their capacity to bind receptors, including their self-recruitment to the signalling complex. It is known that TRAF pro-

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Table 1. Western blot analysis of TRAF2 protein in uterus tissue samples from myometrium and leiomyomas from young and perimenopausal age women. Quantitative analysis indicated, that TRAF2 level in myometrium perimenopausal age women is 107% those in young women.

<table>
<thead>
<tr>
<th>Control</th>
<th>Small myomas</th>
<th>Large myomas</th>
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<tr>
<td></td>
<td>Myometrium</td>
<td>Myometrium</td>
</tr>
<tr>
<td>Reproductive age women</td>
<td>135±7*</td>
<td>140±8*</td>
</tr>
<tr>
<td>Perimenopausal women</td>
<td>145±4*</td>
<td>120±6*</td>
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Differences in TRAF2 expression: * – in reproductive age women and perimenopausal age women; a in small myomas and healthy myometrium; b in periphery of small myomas and healthy myometrium; c in large myomas and healthy myometrium; d in periphery of large myomas and healthy myometrium; e in small myomas and its periphery, f in large myomas and its periphery, are statistically significant for p=0.05.

Table 2. Western blot analysis of TRAF6 protein in uterus tissue samples from myometrium and leiomyomas from young and perimenopausal age women. Quantitative analysis indicated, that TRAF6 level in myometrium perimenopausal age women is 113% those in young women.

<table>
<thead>
<tr>
<th>Control</th>
<th>Small myomas</th>
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<tr>
<td></td>
<td>Myometrium</td>
<td>Myometrium</td>
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<tr>
<td>Reproductive age women</td>
<td>125±6*</td>
<td>135±8*</td>
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<tr>
<td>Perimenopausal women</td>
<td>135±7*</td>
<td>125±5*</td>
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Differences in TRAF6 expression: * – in reproductive age women and perimenopausal age women; a in small myomas and healthy myometrium; b in periphery of small myomas and healthy myometrium; c in large myomas and healthy myometrium; d in periphery of large myomas and healthy myometrium; e in small myomas and its periphery, f in large myomas and its periphery, are statistically significant for p=0.05.
proteins may serve in modulation of receptor’s capacity to induce various signalling pathways [22], which lead to phosphorylation and activation of protein kinases and, then, to activate transcription factors of, among others, AP-1 family.

In our studies we have demonstrated that myometria of myoma-free women manifest evident expressions of TRAF2 and TRAF6 proteins. Their levels were well marked but similar in samples originating from young women and women of perimenopausal age (Fig. 2 and 4; Table 1 and 2). This indicates that absence of diagnosis of myoma presence not necessarily is equivocal with an absence of neoplastic risk in a patient. A stable level of TRAF2 and TRAF6 expression represents a stimulating factor in the NF-κB activation pathway, which, as generally known, directly affects proliferative processes. Even if neither TRAF2 nor TRAF6 affect directly expression of aromatase, their indirect effect is evident. As generally known, the two proteins directly stimulate activity of NF-κB. This nuclear factor may associate with one of promoters, such as I.4, which mobilizes aromatase synthesis pathway.

TRAF proteins also seem to play a significant role in modulation of an early stage in the receptor-induced activation of NF-κB [9,23,24]. As has been demonstrated, NF-κB-induced kinase (NIK) represents a part of the signalling complex, formed by multimerization of TRAF proteins following recruitment to cell surface receptors [15,25-27]. An evidence for the ability of TRAF proteins to affect activation of NF-κB with receptor mediation has originated from the demonstration that TRAF2 may inhibit activation of NF-κB in response to oligomerization of numerous TNFR-linked molecules, including TNFRII, CD40 and other [28]. In our studies we have shown that expression of TNF-α has been increasing in all studied experimental groups (unpublished data). The cytokine directly activates, among others, TRAF2. Subsequently we have shown that also IL-1 reached high levels, particularly in cases of small and large myomas (unpublished data). In concert with TNF-α the cytokine stimulates TRAF6 [29]. The two studied heretofore proteins exert a direct stimulatory effect on NF-κB activity. This may be synonymous to an increased proliferation in the pathologically altered tissue. However, it is suggested that engagement of the proteins may activate NF-κB in more than a single pathway. Although certain TRAF, and TRAF2 in particular, activate NF-κB and AP-1, the differential recruitment of TRAF-activating or TRAF-inhibiting molecules seems to involve just one of several control levels in receptor-induced signal transduction. The fact that TRAF-binding sites in receptors significantly differ in sequence provides an argument for differences in affinity of TRAF molecules binding to various receptor proteins. Moreover, the potential for homo- and/or heterodimerization of TRAF molecules, which leads to formation of higher order complexes might add here another level of control [12]. It is probable that differences in levels of TRAF expression and/or depletion of various TRAF molecules following recruitment to the receptor and the nature of multimerized, TRAF-recruiting receptor result in formation of various poly-protein aggregates and these either allow cells to survive or induce apoptosis [28,30-32].

A few experiments provided the proof for action of TRAF proteins as mediators of life or death and represented subsequent steps in understanding the role of TRAF as one of the key controllers of cell reaction to stress. The clearly higher levels of TRAF2 and TRAF6 expression, documented in both types of myomas and at their periphery may corroborate the hypothesis. As described above, TRAF2 and TRAF6 seem to act as inducers of positive signals for cell growth and proliferation, with mediation of kinase cascades and induction of NF-κB-controlled genes [33]. When the molecules are available in significant quantities, activation of TNFR seems to promote stress response. In contrast, in cells in which the factors are depleted or blocked by inhibitors, the apoptotic response is favoured [28,30-32]. Thus, availability of TRAF proteins and their activity may regulate the key point of cell survival, involving on one hand stress response and, on the other, a programmed cell death.

The data provide proof for important role of TRAF2 in activation of JNK pathway through TNFR. Even if experiments in tissue culture systems indicate that TRAF2 may control activation of NF-κB, existence of TRAF-independent pathways has been
demonstrated. Evidently this might have been expected since, as we have shown, TRAF2 level in myomas was lower than the level of TRAF6.

The experiments suggest that TRAF2 may exert also an anti-apoptotic activity. Studies on cells with deficiency of TRAF2 confirmed the anti-apoptotic activity of TRAF2 and its significance for TNF-induced activation of JNK [34-36]. Thus, even if TRAF2 manifests an evident anti-apoptotic activity, it also can bind members of TNFR family to pro-apoptotic pathways.

Literature references related to the subject are quite scanty. In the few studies on local expression of TRAF2 and TRAF6 in uterine myomas an attempt was made to find out if proteins manifested in myoma cells play a significant role in their growth and if myomas synthesize sufficient amounts of oestrogens for promotion of myoma growth. Myoma cells are thought to be able to synthesize appropriate amounts of oestrogens and, moreover, the local aromatization in myoma tissue is thought to be responsible for the cell growth-promoting effect.

In our paper we were defining level of TRAF expression with two techniques. We indicated that profiles of quality changes are the same but quantity changes are different. One can notice that results received in immunohistochemical stains are visibly higher than that in myomas and clearly higher than the one received in Western blot method. Lower values obtained in Western blot method are is probably caused by the fact that staining in this method is applied to whole mass of sample in the distinction of immunohistochemical staining where its apply exclusively to cells.

In discussions on TRAF proteins, an important observation should be taken into account: in our studies we have noted that in myometria originating from women with myoma presence, expression of TRAF proteins has been evident. Its level has been even higher than that in myomas and clearly higher than the level seen in a healthy myometrium. It should be borne in mind that the samples were isolated outside of the so called surgical safety margin and, thus, they originated from the region of uterus, in which no relapse of myoma should occur. In the region our studies have documented TRAF protein expression sufficiently high to warrant in appropriate conditions the development of de novo myoma. It remains to be established how far from the tumorous lesion the level of TRAF protein expression is sufficient for such a complication. This requires further studies.

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