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Growth hormone/insulin-like growth factor-1 axis, calciotropic hormones and bone mineral density in young patients with chronic viral hepatitis

Czynność osi GH/IGF-I, stężenie hormonów kalciotropowych we krwi oraz gęstość mineralna kości u młodych osób z przewlekłym wirusowym zapaleniem wątroby

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Abstract

Introduction: Chronic liver disease caused by HBV and HCV infections, due to its great prevalence and serious medical consequences, is at the present time a significant clinical problem. An impaired liver function can provoke severe disturbances in calcium and phosphorus homeostasis, and consequently in the bone metabolism resulting in hepatic osteodystrophy. The aim of this study was to determine whether there are significant differences in bone mineral density (BMD) and/or circadian levels of hormones connected with bone metabolism and bone turnover markers in patients with chronic viral hepatitis.

Material and methods: Circadian levels (AUC, area under the curve) of GH, IGF-I, IGFBP-3, osteocalcin (BGLAP), C-terminal telopeptide of type I collagen (ICTP), PTH, 25(OH)D, total calcium and total phosporus were measured in the blood of members of the study group (n = 80). BMD was assessed using the dual-energy X-ray absorptiometry method of the L2-L4 lumbar spine. Data was compared to that of healthy individuals (n = 40).

Results: BMD (1.05 g/cm³ vs. 1.20 g/cm³), total calcium concentration (2.20 mmol/L vs. 2.45 mmol/L), total phosphorus concentration (1.06 mmol/L vs. 1.33 mmol/L), IGF-I (AUC 3,982.32 ng/mL vs. 5,167.61 ng/mL), IGFBP-3 (AUC 725.09 ng/L vs. 944.35 ng/L), 25(OH)D (AUC 356.35 ng/mL vs. 767.53 ng/mL) and BGLAP (AUC 161.39 ng/L vs. 298 ng/L) were lower in the study group. GH (AUC 88.3 ng/mL vs. 48.04 ng/mL), iPTH (AUC 1,201.94 pg/mL vs. 711.73 pg/mL) and ICTP (AUC 104.30 μ g/L vs. 54.49 μ g/L) were higher in patients with hepatitis. Positive correlations were noted between bone mineral density and IGF-I, IGFBP-3, and BGLAP levels.

Conclusions: Chronic viral hepatitis causes a decrease in bone mineral density. Impaired liver function disrupts homeostasis of the calcium–vitamin D–parathyroid hormone axis and provokes secondary hyperparathyroidism. Chronic viral hepatitis induces a decrease in the synthesis of IGF-I and IGFBP-3 and an increase in GH secretion. Hepatic osteodystrophy is probably caused by both changes in calciotropic hormones as well as in the somatotropin hormone axis. (Endokrynol Pol 2015; 66 (1): 22–29)

Key words: GH; IGF-I; PTH; osteocalcin; vitamin D; BMD; chronic hepatitis; bone; liver

Streszczenie

Wstęp: Przewlekłe zakażenia HBV i HCV są obecnie znaczącym problemem klinicznym. W wyniku zaburzeń czynności wątroby może dochodzić do zaburzeń w homeostazie wapnia i fosforu oraz w metabolizmie kostnym prowadzących do osteodystrofii wątrobowej. Celem badania była ocena gęstości mineralnej kości (BMD), okołodobowych stężeń hormonów związanych z metabolizmem kości oraz markerów obrotu kostnego u chorych na przewlekłe wirusowe zapalenie wątroby.

Materiał i metody: W grupie badanej (n = 80) oznaczano we krwi okołodobowe stężenia (AUC, area under the curve [pole pod krzywą]) GH, IGF-I, IGFBP-3, osteokalcyny (BGLAP), C-terminalnego telopeptydu kolagenu typu I (ICTP), PTH, 25(OH)D, całkowitego wapnia oraz fosforu. BMD (L2-L4) oceniono z użyciem DEXA. Dane porównano ze zdrową grupą kontrolną (n = 40).

Wyniki: BMD (1,05 g/cm³ vs. 1,20 g/cm³), stężenia wapnia (2,20 mmol/l vs. 2,45 mmol/l) i fosforu (1,06 mmol/l vs. 1,33 mmol/l), IGF-I (AUC 3982,32 ng/ml vs. 5167,61 ng/ml), IGFBP-3 (AUC 725,09 ng/l vs. 944,35 ng/l), 25(OH)D (AUC 356,35 ng/ml vs. 767,53 ng/ml), BGLAP (AUC 161,39 ng/l vs. 298 ng/l) okazały się niższe w grupie badanej niż w grupie kontrolnej, zaś stężenia GH (AUC 88,3 ng/ml vs. 48,04 ng/ml),

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PTH (AUC 1201,94 pg/ml vs. 711,73 pg/ml) i ICTP (AUC 104,30 μ g/l vs. 54,49 μ g/l) były większe u osób z zapaleniem wątroby. Stwierdzono dodatnią korelację między BMD a stężeniami IGF-I, IGFBP-3 oraz BGLAP.

Wnioski: Przewlekłe wirusowe zapalenie wątroby prowadzi do zmniejszenia gęstości mineralnej kości. Upośledzona funkcja wątroby zakłóca homeostazę wapnia, witaminy D, PTH, prowadzi do wtórnej nadczynności przytarczyc. Dochodzi do zmniejszenia syntezy IGF-I i IGFBP-3 oraz do zwiększenia wydzielania GH. Osteodystrofia wątrobowa jest prawdopodobnie spowodowana zarówno poprzez zmiany stężenia hormonów kalciotropowych, jak i zaburzenia funkcjonowania osi somatotropinowej. (Endokrynol Pol 2015; 66 (1): 23–29)

Słowa kluczowe: GH; IGF-I; osteokalcyna; witamina D; gęstość mineralna kości; przewlekłe zapalenie wątroby

Introduction

Chronic liver disease caused by HBV and HCV infections is at the present time a significant clinical problem, given the numerous complications of liver dysfunction as a consequence of a chronic inflammatory state. Moreover, the prevalence of HBV and HCV infections is estimated at respectively 0.1-7% and 0.4-20% of the general population in some European regions [1].

The liver, due to its complex metabolic and secretory functions, has long been classified as a functional part of the endocrine system [2–5]. Among the many aspects of the interaction with the endocrine system, an interesting one is the effect on bone metabolism. This influence is manifested by the interaction with the somatotropic axis, the regulation of the calciotropic hormones — PTH and vitamin D, or even bone turnover markers such as osteocalcin, or C-terminal telopeptide of type I collagen [6–10].

According to previous studies, osteopenia, osteoporosis or osteomalacia may affect as many as 40–50% of patients suffering from impaired liver function. These changes appear in the literature under the name of hepatic osteodystrophy [11–14]. Primary osteoporosis is characterised by specific features: it applies to younger patients, requires a primary disease, and appears as the result of a combination of different pathogenetic factors [15–17]. To date, the pathogenesis and mechanism of hepatic osteodystrophy has not been well explored. It is still not possible to consistently and unambiguously present the factors contributing to this phenomenon [5, 11–13, 18–21].

The aim of this study was to evaluate some potential pathogenetic factors influencing bone status in patients with chronic active viral hepatitis, and to determine their importance in hepatic osteodystrophy development. For this purpose, we decided to assess the bone mineral density (BMD) and hormones and markers associated with bone metabolism. In evaluating these parameters we took into account the existence of diurnal chronobiological cycles, by making multiple measurements throughout the day. Among the chosen parameters were: somatotropic axis hormones — growth hormone (GH), insulin-like growth factor-I (IGF-I), insulin-like growth factor binding protein 3 (IGFBP-3), and calciotropic hormones — parathyroid

hormone (PTH) and calcidiol [25(OH)D] reflecting the level of vitamin D. Of bone turnover markers, we chose osteocalcin (BGLAP; bone gamma-carboxyglutamate protein) as an indicator of osteogenesis, and C-terminal telopeptide of type I collagen (ICTP) as an indicator of bone loss.

Material and methods

Eighty patients with chronic viral hepatitis (B or/and C) were included in the study. The study group consisted of 45 men with a mean age of 37 years (SD = 6.3 years) and 35 women with a mean age of 34 years (SD = 5.2years). The inclusion criterion was a diagnosis of chronic B or C hepatitis. This diagnosis had been based on the clinical picture, laboratory tests results — ALT, AST, GGTP, alkaline phosphatase, total bilirubin, INR, proteinogram, serological tests results — HBsAg, HBcAg, HBeAg, anti-HBs, anti-HBc, anti-HBe, molecular tests results — in B-type hepatitis the presence of HBV DNA and DNA-polymerase activity, in C-type hepatitis the presence of HCV RNA and liver biopsy. Hepatitis group candidates were excluded if they suffered from chronic diseases of the endocrine system, cardiovascular system, respiratory system, or excretory system. Also candidates who used calcium supplements, vitamin D, corticosteroids or other drugs influencing bone metabolism were excluded. Additional exclusion criteria were a history of alcohol abuse or a family history of osteoporosis. The control group consisted of 40 healthy volunteers. This group included 20 men and 20 women with a mean age of 36 years (SD = 6.7 years) who did not meet exclusion criteria. All participants underwent a full clinical examination including calculation of BMI and routine biochemical blood tests. Permission to conduct the study was obtained from the Local Ethics Committee for Scientific Research of the Medical University of Silesia.

Blood sample analyses

All participants had a peripheral venous catheter established into their antecubital vein one hour before the start of the study. Blood samples, in the amount of 8 ml each, were collected through this catheter at 08.00, 12.00, 16.00, 20.00, 24.00 and 04.00. The samples were centrifuged to obtain blood serum, which was frozen

Table I. Characteristics of patients with chronic viral hepatitis and control group

Tabela I. Charakterystyka chorych z przewlektym wirusowym zapaleniem wątroby oraz grupy kontrolnej

	Chronic hepatitis (N = 80)	Control group (N = 40)	p value
Age [years]	34 (SD ± 0.82)	36 (SD ± 1.50)	NS
BMI [kg/m²]	22.3 (SD ± 0.50)	22.8 (SD ± 0.89)	NS
ALT [IU/L]	120.3 (SD ± 12.74)	19.2 (SD ± 1.65)	P < 0.001
ALP [IU/L]	180.43 (SD ± 7.51)	182.30 (SD ± 7.02)	NS
Total bilirubin [µmol/L]	19.4 (SD ± 2.57)	13.2 (SD ± 1.58)	NS
Creatinine [µmol/L]	89.6 (SD ± 3.38)	84.7 (SD ± 3.89)	NS

BMI — body mass index; ALT — alanine transaminase; ALP — alkaline phosphatase; NS — statistically non-significant

at -70°C for subsequent analysis. In every serum sample, the radioimmunological assay method was used to assess the concentration of selected parameters: Growth hormone (GH-RIA kit, OPiD POLATOM, Poland), insulin-like growth factor-I (IGF-I RIA-kit, Biosource, Belgium), insulin-like growth factor binding protein-3 (IGFBP-3 IRMA-kit, Diagnostic Systems Laboratories, TX, USA), osteocalcin (BGLAP IRMA-kit, Biosource, Belgium), C-terminal telopeptide of type I collagen (ICTP RIA-kit, Orion Diagnostica, Finland), parathyroid hormone (iPTH IRMA-kit, Biosource, Belgium), and calcidiol (25(OH)D3 IRMA-kit, Biosource, Belgium). Total calcium serum concentration was assessed by the colorimetric method (Calcium Liquicolor, Humen, Germany), as was total inorganic phosphorus concentration (Phosphorous Liquirapid, Humen, Germany).

Bone densitometry

All participants underwent assessment of bone mineral density (BMD) with the use of the dual-energy X-ray absorptiometry method (DEXA) of the L2-L4 lumbar spine. A Lunar DPX densitometer (General Electric, USA) was used to make measurements. Measurement results were presented as either absolute density values [g/cm³] or T-score and Z-score values.

Statistics

All acquired data was implemented to create a database using Microsoft Excel forming a part of Microsoft Office 2000 (Microsoft, USA). For each quantitative parameter, basic statistical characteristics were made — mean, median, standard deviation (SD), and standard error of the mean (SEM). Area under curve (AUC) values

Table II. Bone mineral density, total calcium and inorganic phosphorus parameters of patients with chronic hepatitis and control group

Tabela II. Gęstość mineralna kości, stężenie wapnia oraz fosforu we krwi u chorych z przewlekłym wirusowym zapaleniem wątroby oraz w grupie kontrolnej

	Chronic hepatitis	Control s	p value
BMD [g/cm ³]	1.05 (SD ± 0.029)	1.20 (SD ± 0.032)	p = 0.0011
BMD Z-score	-0.36	0.48	p < 0.05
BMD T-score	-0.52	0.18	p < 0.05
Total calcium [mmol/L]	2.20 (SD ± 0.031)	2.45 (SD ± 0.054)	p < 0.001
Inorganic phosphorus [mmol/L]	1.06 (SD ± 0.029)	1.33 (SD ± 0.036)	p < 0.001

BMD — bone mineral density

of the parameters was determined by the trapezoidal method. Shapiro-Wilk test was performed for the evaluation of a normal distribution parameter. Student's t-test was used for the comparison of unpaired groups of parameters that follow normal distribution. In the use of unrelated variables, Fisher's exact test was used to check the homogeneity of variance. Pearson's correlation coefficient was used to investigate multivariate data correlation. A p value < 0.05 was accepted as statistically significant.

Results

We found a significant difference in the ALT serum concentration. It was greatly elevated (120.3 IU/L vs. 19.2 IU/L, p < 0.001) in patients compared to controls (Table I and II).

We observed a difference in BMD in the L2-L4 vertebrae densitometry (DEXA) between study participants and the control group, taking into account both the absolute BMD values (1.05 g/cm³ vs.~1.20 g/cm³, p = 0.0011) and the indications of Z-score (-0.36 vs.~0.48, p < 0.05) and T-score (-0.52 vs.~0.18, p < 0.05). We noted, however, that despite much lower values compared to the controls, the results of the study group members were within the reference values. In the group of patients with chronic hepatitis, we found in blood lower total calcium levels (2.20 mmol/L vs.~2.45 mmol/L, p < 0.001) and inorganic phosphorus levels (1.06 mmol/L vs.~1.33 mmol/L, p < 0.001) (Table III).

We observed significantly increased total daily release of GH in the study group compared to controls (88.3 ng/mL vs. 48.04 ng/mL, p < 0.001). However, we

Table III. Diurnal oscillations of GH, IGF-I and IGFIBP-3 concentrations in blood of patients with chronic viral hepatitis and control group

Tabela III. Okołodobowe stężenia GH, IGF-I, IGFBP-3 we krwi chorych z przewlekłym wirusowym zapaleniem wątroby oraz w grupie kontrolnej

Sampling GH [ng/mL]				IGF-I [ng/mL]			IGFBP-3 [ng/L]		
time	Study group	Control group	p value	Study group	Control group	p value	Study group	Control group	p value
08.00		1.21 (SD ± 0.034)	•	183.93 (SD ± 8.29)	263.03 (SD ± 9.25)	•	39.97 (SD ± 1.27)	50.16 (SD ± 2.37)	p < 0.001
12.00		1.23 (SD ± 0.034)			251.07 (SD ± 9.59)	•	38.62 (SD ± 1.19)		p < 0.001
16.00		2.52 (SD ± 0.123)	•		252.94 (SD ± 8.62)	•	38.34 (SD ± 1.44)	46.70 (SD ± 2.41)	p = 0.0036
20.00		4.56 (SD ± 0.119)	•		274.87 (SD ± 9.07)	•	35.84 (SD ± 1.37)	48.79 (SD ± 2.20)	p < 0.001
24.00		3.56 (SD ± 0.106)		233.76 (SD ± 8.03)	282.19 (SD ± 9.29)	p < 0.001	34.13 (SD ± 1.17)	48.29 (SD ± 2.22)	p < 0.001
04.00		1.32 (SD ± 0.141)	•		240.39 (SD ± 8.37)	p < 0.001	31.43 (SD ± 1,32)	42.98 (SD ± 2.39)	p < 0.001
AUC			•	3,982.32 (SD ± 133.012)		•	725.09 (SD ± 21.83)		p < 0.001

GH — growth hormone concentration; IGF-I — insulin-like growth factor 1; IGFBP-3 — insulin-like growth factor binding protein 3; AUC — area under curve

did not notice significant changes in the daily oscillation profile of GH ejection. The study group was characterised by lower daily concentrations of IGF-I (3,982.32 ng/mL vs. 5,167.61 ng/mL, p < 0.001) compared to controls. IGFBP-3 showed similar changes (725.09 ng/L vs. 944.35 ng/L, p < 0.001). We did not observe changes in the daily oscillations of these parameters (Table IV).

Daily secretion of calcidiol was lower in patients with chronic viral hepatitis compared to control group (356.35 ng/mL vs. 767.53 ng/mL, p < 0.001). Moreover, we noted a flattening of the profile of daily 25(OH)D oscillation in patients with hepatitis, manifested as the disappearance of the peak of the highest concentration which occurred about 12.00 in the healthy control group. An inverse relationship concerned the diurnal concentration of intact parathyroid hormone (iPTH), which was higher in the study group (1,201.94 pg/mL vs. 711.73 pg/mL, p < 0.001) (Table V).

We observed that the daily concentration of osteocalcin was lower in patients with hepatitis than in the control group (161.39 ng/L vs. 298.00 ng/L, p < 0.001). In the case of ICTP, we noticed an increase in its release among participants of the study group (104.30 μ g/L vs. 54.49 μ g/L, p < 0.001).

Our results revealed a statistically significant positive correlation between BMD values and diurnal serum concentrations of IGF-I, IGFBP-3 and BGLAP in blood. In addition, we found a positive link between the concentrations of IGFBP-3 and BGLAP. We also observed a significant negative bond between BMD values

and the daily GH secretion. Additionally, we found a negative correlation linking the daily levels of IGF-I and ICTP or between IGF-I and iPTH.

Discussion

The material analysed in this paper is highly homogeneous. Among the participants of the study group, we did not identify the features of cirrhosis of the liver, severe cholestasis, or other conditions that may have an impact on the calcium-phosphate metabolism. The general characteristics differentiating feature of research subjects for the members of the control group was a significantly elevated level of alanine aminotransferase (ALT; 120.3 IU/L vs. 19.2 IU/L, p < 0.001), reflecting active inflammation of the liver. Moreover, individuals included in our study were characterised by a relatively low age (mean age 34 years, SD = 5.2years), which enabled us to exclude changes in bone metabolism associated with senility [3, 14, 21-26]. In our study, we implemented repeated measurements over the course of 24 hours, allowing us to reflect diurnal level variations with respect to chronobiological cycles [3, 4, 8, 27–29].

In our study, we found a statistically significant difference in BMD expressed either as absolute values, or as a T-score and Z-score occurring between patients with hepatitis and the controls (BMD $vs.~1.05~g/cm^3$. $1.20~g/cm^3$; p=0.0011, T-score -0.52~vs.~0.18, p<0.05, Z-score -0.36~vs.~0.48, p<0.05). The results, despite

Table IV. Diurnal oscillations of 25(OH)D and iPTH concentrations in blood of patients with chronic viral hepatitis and in control group

Tabela IV. Okołodobowe stężenia 25(OH)D i iPTH we krwi chorych z przewlekłym wirusowym zapaleniem wątroby oraz w grupie kontrolnej

Sampling time	25(OH)D [ng/mL]			iPTH [pg/mL]	iPTH [pg/mL]			
	Study group	Control group	P value	Study group	Control group	p value		
08.00	18.35 (SD ± 1.35)	34.90 (SD ± 3.82)	p < 0.001	56.46 (SD ± 8.97)	35.72 (SD ± 6.53)	p = 0.0384		
12.00	18.20 (SD ± 1.43)	48.10 (SD ± 6.32)	p < 0.001	53.11 (SD ± 5.36)	33.25 (SD ± 4.78)	p = 0.0061		
16.00	17.49 (SD ± 1.69)	37.12 (SD ± 4.37)	p < 0.001	64.98 (SD ± 10.71)	38.33 (SD ± 8.09)	p = 0.0107		
20.00	17,72 (SD ± 1,60)	38.90 (SD ± 7.13)	p = 0.0062	55.21 (SD ± 6.68)	33.04 (SD ± 5.06)	p = 0.0056		
24.00	17.25 (SD ± 1.39)	35.84 (SD ± 3.86)	p < 0.001	57.05 (SD ± 6.50)	32.12 (SD ± 4.43)	p = 0.0013		
04.00	17.85 (SD ± 1.82)	32.38 (SD ± 4.23)	p = 0.0055	70.44 (SD ± 10.95)	39.38 (SD ± 9.51)	p = 0.0033		
AUC	356.35 (SD ± 28.74)	767.53 (SD ± 356.71)	p < 0.001	1,201.94 (SD ± 114.62)	711.73 (SD ± 121.68)	p < 0.001		

25(OH)D — calcidiol; iPTH — intact parathyroid hormone; AUC — area under curve

Table V. Diurnal oscillations of BGLAP and ICTP concentrations in blood of patients with chronic viral hepatitis and in control group

Tabela V. Okołodobowe stężenia BGLAP i ICTP we krwi chorych z przewlekłym wirusowym zapaleniem wątroby oraz w grupie kontrolnej

Sampling time	BGLAP [ng/L]			ICTP [μ g/L]	ICTP [µg/L]		
	Study group	Control group	p value	Study group	Control group	p value	
08.00	8.26 (SD ± 0.71)	13.36 (SD ± 1.04)	p < 0.001	5.72 (SD ± 0.46)	2.77 (SD ± 0.29)	p < 0.001	
12.00	7.58 (SD ± 0.52)	14.61 (SD ± 1.28)	p < 0.001	4.87 (SD ± 0.36)	2.69 (SD ± 0.35)	p < 0.001	
16.00	8.32 (SD ± 0.91)	13.44 (SD ± 1.09)	p = 0.0014	4.77 (SD ± 0.40)	2.46 (SD ± 0.37)	p < 0.001	
20.00	7.72 (SD ± 0.69)	14.52 (SD ± 1.22)	p < 0.001	4.70 (SD ± 0.34)	2.52 (SD ± 0.26)	p < 0.001	
24.00	7.96 (SD ± 0.84)	15.30 (SD ± 1.27)	p < 0.001	5.40 (SD ± 0.384)	2.67 (SD ± 0.24)	p < 0.001	
04.00	8.66 (SD ± 1.08)	17.60 (SD ± 1.59)	p < 0.001	6.18 (SD ± 0.49)	3.23 (SD ± 0.26)	p < 0.001	
AUC	161.39 (SD ± 13.40)	298.0 (SD ± 21.44)	p < 0.001	104.30 (SD ± 6.90)	54.49 (SD ± 4.84)	p < 0.001	

 $BGLAP - bone \ gamma-carboxyglutamate \ protein, \ osteocalcin; \ ICTP - carboxy-terminal \ telopeptide \ of \ type \ I \ collagen; \ AUC - area \ under \ curve$

a decrease in BMD values, do not meet the criteria for the diagnosis of osteopenia or osteoporosis [15, 16]. Our results comply with those of previous reports [30-35], but the loss of bone mass among our subjects was significantly lower than has been found by some other authors. We believe that this is due to the decreased severity of hepatic impairment presented by our patients. Pelazas-Gonzalez et al. [36] observed

that chronic HCV infection in well-nourished patients with preserved liver function does not cause osteoporosis. The selection of our study group, excluding cholestasis, helped us to eliminate the influence of abnormal intestinal absorption of calcium caused by bile obstruction [37] and cholestasis itself, which is considered to be an independent risk factor for osteoporosis [38].

Among patients with hepatitis, we noticed a statistically significant reduction in total calcium concentration in blood (2.20 mmol/L vs. 2.45 mmol/L, p < 0.001) as well as the level of inorganic phosphorus (1.06 mmol/L vs. 1.33 mmol/L, p < 0.001) compared to the value achieved by the control group. We believe that our results are derived from abnormalities in the metabolism of vitamin D and PTH [11, 15].

The common complication described in chronic liver diseases, and cirrhosis in particular, is hypogonadism [4, 26, 39], which also stands as an important risk factor for osteoporosis [10, 15, 40, 41]. This disorder manifests in laboratory tests as hyperoestrogenism [4, 9, 42–44], and a decrease in free testosterone serum level [43, 44]. It is true that our study did not have marked levels of sex hormones, but by carefully collecting medical history and physical examination we were most probably able to exclude the influence of gonadal dysfunction among study participants. None of the study participants showed any signs of alcohol abuse. The consumption of excessive amounts of alcohol is a risk factor for osteoporosis, independent of hypogonadism and liver dysfunction [26, 45].

In the course of our study, we observed that in patients with chronic hepatitis daily IGF-I level (3,982.32 ng/mL $vs.\,5$,167.61 ng/mL, p < 0.001) and IGFBP-3 level (725.09 ng/mL $vs.\,944.35$ ng/mL, p < 0.001) were reduced compared to the control group. Moreover, these phenomena were accompanied by increased daily secretion of GH in the study group (88.3 ng/mL $vs.\,48.04$ ng/mL, p < 0.001). This trend has been observed by different authors [33, 46, 47]. We noted a positive correlation linking BMD and IGF-I and IGFBP-3.

These observations confirm the importance of growth hormone axis function and, consequently, the concentrations of IGF-I and IGFBP-3, for bone metabolism [48–50]. Another feature is the appearance of a positive relationship between the values of IGF-I and BGLAP and negative between IGF-I and ICTP and PTH. These relationships indicate a direct effect of liver function on bone anabolism and catabolism respectively.

In our opinion, the cause of the increase in the daily GH secretion in patients with chronic hepatitis disorder feedback was due to impaired production of IGF-I in the liver [8, 51]. However, there are reports which undermine the aforementioned hypothesis. In animal studies, the elimination of hepatic IGF-I synthesis caused its level to decrease by 75%, although that action did not have any significant influence on the BMD [52, 53]. Furthermore, some scientists deny the existence of a link between IGF-I and BMD in men [54–56]. Papers dealing with osteoporosis in liver disease suggest the existence of a link between reduced BMI and decreased IGF-I and BMD [57-61]. However, our observation did not confirm such a relationship.

Our results show a significant reduction in blood levels of calcidiol in study subjects compared to the control group (712.71 ng/mL vs. 1,535 ng/mL, p < 0.001). Vitamin D deficiency may lead to hypocalcaemia, the development of secondary hyperparathyroidism, and hypophosphatemia [62]. Our research revealed a similar tendency, but it should be mentioned that, despite significant differences in the concentrations of calcium and phosphorus in relation to controls, they stayed within the range of reference values for a healthy population. Results demonstrating reduced levels of (25(OH)D) in the serum of patients with impaired liver function, including cirrhosis, have appeared in the past [5, 30, 63, 64]. However, some reports remain contradictory. Diamond et al. [65] and Tsunoka et al. [14] observed that the levels of (25(OH)D) and (1,25(OH)2D) show significant differences between the groups of patients with liver cirrhosis and chronic hepatitis. They point to the lower values of these parameters in patients with cirrhosis, suggesting a relationship between both calcidiol or calcitriol levels and liver efficiency. But there are also other opinions: Duarte et al. [66] did not show any differences in this area between people with liver cirrhosis and a healthy control group. It is worth noting that we found a change in the circadian oscillation profile of serum (25(OH)D); in the patient group, it showed a much narrower range of diurnal changes in addition to an overall lower level compared to healthy subjects. We believe this is the premise suggesting liver dysfunction in the regulation of vitamin D, far in advance of the development of cirrhosis.

Our study showed significantly elevated levels of PTH in the hepatitis group members compared to the control group (1,201.94 pg/mL vs. 711.73 pg/mL, p < 0.001). An increase in PTH levels in patients with chronic hepatitis or cirrhosis is usually the result of secondary hyperparathyroidism [32, 66, 67]. As previously mentioned, many authors have suggested that the reason for hyperparathyroidism in such patients is the decrease of vitamin D level [67, 68]. Previous studies have shown that the lowest normal calcidiol levels, at which PTH secretion is minimal, is on average 40 ng/mL [62, 69]. Among participants in our study, in all the samples taken during the day, the concentrations of (25(OH)D) levels were below the designated threshold. This seems to confirm the hypothesis of hyperparathyroidism being dependent on vitamin D levels in patients with chronic active hepatitis. The effect of PTH on bone metabolism has been the subject of numerous considerations. In the course of hyperparathyroidism, the intensification of bone resorption with an increase in the number of osteoclasts, severe alteration of bone structure and a reduction of BMD has been demonstrated [25, 70], which is consistent with our results.

However, there have also been reports that short-pulse increases in PTH have anabolic effects on bone mass and increases their mechanical strength [24, 71]. There is a suggestion that this effect results from the extension of the life of osteoblasts by PTH-dependent inhibition of apoptosis [72]. In patients with chronic active hepatitis with secondary hyperparathyroidism, one would expect an increased bone turnover; however, this occurs only in about 50% of patients [30, 54, 73], and in the remainder a dynamic bone disease has been described [73].

Our study showed significantly lower serum BGLAP concentration in patients with hepatitis (161.39 ng/L vs. 298 ng/L, p < 0.001), and significantly higher levels of ICTP (104.30 μ g/L vs. 54.49 μ g/L, p < 0.001) than in the control group. In addition, we noted a statistically significant positive correlation joining BMD with BGLAP concentration. Our findings are similar to those from other authors [74, 75], while Gallego-Rojo et al. [30] showed a reduction in the concentration of BGLAP only in patients with end-stage liver cirrhosis. Previous studies have also shown increased activity of bone resorption including ICTP [30, 76]. Most research leads us to the hypothesis that the severity of the resorption process is the predominant cause of hepatic osteodystrophy.

Conclusions

In our study, we have shown that the impairment of liver function in chronic hepatitis due to HBV or HCV infection leads in young patients to significant disorders of bone metabolism. These changes manifest themselves as a decrease in bone mineral density which may reflect in an increased risk of pathological bone fractures.

In our research we observed that chronic liver inflammation leads to impaired function of the calcium-vitamin D-parathyroid hormone axis, causes secondary hyperparathyroidism and, consequently, an imbalance in bone turnover processes. Furthermore, we have shown that in chronic active hepatitis, somatotropin axis adjustment is disturbed; this manifests as a decrease in the synthesis of IGF-I and IBFBP-3 and, coupled with these, an increase in growth hormone secretion.

These observations allow us to conclude that an important pathogenetic role underlying hepatic osteodystrophy is played by both changes in calciotropic hormones and the hormone axis. We believe that there may be some significance in a change in the metabolism of bone turnover markers — BGLAP and ICTP.

We believe that the issue we have raised in this study requires further research.

References

- European Centre for Disease Prevention and Control. Hepatitis B and C in the EU neighbourhood: prevalence, burden of disease and screening policies. Stockholm: ECDC; 2010.
- Kuemmerle JF. Insulin-like growth factors in the gastrointestinal tract and liver. Endocrinol Metab Clin North Am 2012; 4: 409–423.
- Marek B, Buntner B, Kos-Kudła B et al. Liver and endocrine system. Part I: pituitary-thyroid axis activity disorders. Wiad Lek 1998; 51: 271–276.
- Marek B, Buntner B, Kos-Kudła B et al. Liver and endocrine system. Part II: pituitary-gonadal axis and adrenal gland activity disturbances. Wiad Lek 1998; 51: 352–359.
- Diamond T, Stiel D, Lunzer M et al. Osteoporosis and skeletal fractures in chronic liver disease. Gut 1990; 31: 82–87.
- Johansson AG, Eriksen EF, Lindh E et al. Reduced serum levels of the growth hormone-dependent insulin-like growth factor binding protein and a negative bone balance at the level of individual remodeling units in idiopathic osteoporosis in men. J Clin Endocrinol Metab 1997; 82: 2795–2798.
- Johansson AG, Burman P, Westermark K et al. The bone mineral density in acquired growth hormone deficiency correlates with circulating levels of insulin-like growth factor I. J Intern Med 1992; 232: 447–452.
- Marek B, Kułakowska E, Rzytki P et al. Function of the growth hormone axis-insulin-like growth factors-insulin-like growth factor binding proteins in patients with chronic liver diseases. Pol Merkur Lekarski 2001; 10: 185–190.
- Herrera A, Lobo-Escolar A, Mateo J et al. Male osteoporosis: A review. World J Orthop 2012; 12: 223–234.
- Drake MT, Murad MH, Mauck KF et al. Risk factors for low bone massrelated fractures in men: a systematic review and meta-analysis. J Clin Endocrinol Metab 2012; 97: 1861–1870.
- 11. Goel V, Kar P. Hepatic osteodystrophy. Trop Gastroenterol 2010; 31: 82-86.
- Gasser RW. Cholestasis and metabolic bone disease a clinical review. Wien Med Wochenschr 2008; 158: 553–557.
- Rouillard S, Lane NE. Hepatic osteodystrophy. Hepatology 2001; 33: 301–307.
- Tsuneoka K, Tameda Y, Takase K et al. Osteodystrophy in patients with chronic hepatitis and liver cirrhosis. J Gastroenterol 1996; 31: 669–678.
- Walker-Bone K. Recognizing and treating secondary osteoporosis. Nat Rev Rheumatol 2012; 8: 480–492.
- Compston JE. Bone densitometry and clinical decision making. J Clin Densitom 1999: 2: 5–9.
- 17. Johnell O, Odén A, De Laet C et al. Biochemical indices of bone turnover and the assessment of fracture probability. Osteoporos Int 2002; 13: 523–526.
- 18. Maurel DB, Boisseau N, Benhamou CL et al. Alcohol and bone: review of dose effects and mechanisms. Osteoporos Int 2012; 23: 1–16.
- Caregaro L, Alberino F, Amodio P et al. Nutritional and prognostic significance of insulin-like growth factor 1 in patients with liver cirrhosis. Nutrition 1997; 13: 185–190.
- Johansson AG, Forslund A, Hambraeus L et al. Growth hormone-dependent insulin-like growth factor binding protein is a major determinant of bone mineral density in healthy men. J Bone Miner Res 1994; 9: 915–921.
- Santolaria F, González-Reimers E, Pérez-Manzano JL et al. Osteopenia assessed by body composition analysis is related to malnutrition in alcoholic patients. Alcohol 2000; 22: 147–157.
- Demontiero O, Vidal C, Duque G. Aging and bone loss: new insights for the clinician. Ther Adv Musculoskelet Dis 2012; 4: 61–76.
- Ohashi S, Tanaka S. Etiology of osteoporosis decrease of bone mineral density and deterioration of bone quality. Clin Calcium 2012; 22: 805–811.
- Hock JM, Gera I. Effects of continuous and intermittent administration and inhibition of resorption on the anabolic response of bone to parathyroid hormone. J Bone Miner Res 1992; 7: 65–72.
- Parisien M, Silverberg SJ, Shane E et al. The histomorphometry of bone in primary hyperparathyroidism: preservation of cancellous bone structure. J Clin Endocrinol Metab 1990; 70: 930–938.
- Diamond T, Stiel D, Posen S. Effects of testosterone and venesection on spinal and peripheral bone mineral in six hypogonadal men with hemochromatosis. J Bone Miner Res 1991; 6: 39–43.
- Morris CJ, Aeschbach D, Scheer FA. Circadian system, sleep and endocrinology. Mol Cell Endocrinol 2012; 349: 91–104.
- 28. Szulc P, Kaufman JM, Delmas PD. Biochemical assessment of bone turnover and bone fragility in men. Osteoporos Int 2007; 18: 1451–1461.
- Frystyk J, Freda P, Clemmons DR. The current status of IGF-I assays
 — a 2009 update. Growth Horm IGF Res 2010; 20: 8–18.
- Gallego-Rojo FJ, Gonzalez-Calvin JL, Munoz-Torres M et al. Bone mineral density, serum insulin-like growth factor I, and bone turnover markers in viral cirrhosis. Hepatology 1998; 28: 695–699.
- Guanabens N, Pares A, Marinoso L et al. Factors influencing the development of metabolic bone disease in primary biliary cirrhosis. Am J Gastroenterol 1990; 85: 1356–1362.

- Schiefke I, Fach A, Wiedmann M et al. Reduced bone mineral density and altered bone turnover markers in patients with non-cirrhotic chronic hepatitis B or C infection. World J Gastroenterol 2005; 11: 1843–1847.
- George J, Ganesh HK, Acharya S et al. Bone mineral density and disorders of mineral metabolism in chronic liver disease. World J Gastroenterol 2009; 15: 3516–3522.
- Cheong JY, Cho SW, Hwang IL et al. Association between chronic hepatitis B virus infection and interleukin-10, tumor necrosis factor-alpha gene promoter polymorphisms. J Gastroenterol Hepatol 2006; 21: 1163–1169.
- Redondo-Cerezo E, Casado-Caballero F, Martin-Rodriguez JL et al. Bone mineral density and bone turnover in non-cirrhotic patients with chronic hepatitis C and sustained virological response to antiviral therapy with peginterferon-alfa and ribavirin. Osteoporos Int 2014; 25: 1709–1715.
- Pelazas-González R, González-Reimers E, Alemán-Valls MR et al. Bone alterations in hepatitis C virus infected patients. Eur J Intern Med 2013; 24: 92–96.
- Gurlek A, Gedik O. Endogenous sex steroid, GH and IGF-I levels in normal elderly men: relationships with bone mineral density and markers of bone turnover. J Endocrinol Invest 2001; 24: 408–414.
- Lindor KD, Janes CH, Crippin JS et al. Bone disease In primary biliary cirrhosis: does ursodeoxycholic acid make a difference? Hepatology 1995; 21: 389–392.
- George J, Ganesh HK, Acharya S at al. Bone mineral density and disorders of mineral metabolism in chronic liver disease. World J Gastroenterol 2009; 15: 3516–3522.
- Krysiak R, Okopień B. Pathogenesis and clinical presentation of andropause. Pol Merkur Lekarski 2012; 187: 70–73.
- Dandona P, Rosenberg MT. A practical guide to male hypogonadism in the primary care setting. Int J Clin Pract 2010; 64: 682–696.
- Gavaler JS, Van Thiel DH. The association between moderate alcoholic beverage consumption and serum estradiol and testosterone levels in normal postmenopausal women: relationship to the literature. Alcohol Clin Exp Res 1992; 16: 87–92.
- Longcope C, Pratt JH, Schneider S et al. Estrogen and androgen dynamics in liver disease. J Endocrinol Invest 1984; 7: 629–634.
- Wang YJ, Wu JC, Lee SD et al. Gonadal dysfunction and changes in sex hormones in postnecrotic cirrhotic men: a matched study with alcoholic cirrhotic men. He patogastroenterology 1991; 38: 531–534.
- 45. Poor G, Atkinson EJ, O'Fallon WM et al. Predictors of hip fractures in elderly men. J Bone Miner Res 1995; 10: 1900–1907.
- Colakoğlu O, Taşkiran B, Colakoğlu G et al. Serum insulin like growth factor-1 (IGF-1) and insulin like growth factor binding protein-3 (IG-FBP-3) levels in liver cirrhosis. Turk J Gastroenterol 2007; 18: 245–249.
- 47. Raslan HM, Elhosary Y, Ezzat WM et al. The potential role of insulin-like growth factor 1, insulin-like growth factor binding protein 3 and bone mineral density in patients with chronic hepatitis C virus in Cairo, Egypt. Trans R Soc Trop Med Hyg 2010; 104: 429–432.
- Perrini S, Laviola L, Carreira MC et al. The GH/IGF1 axis and signaling pathways in the muscle and bone: mechanisms underlying agerelated skeletal muscle wasting and osteoporosis. J Endocrinol 2010; 205: 201–210.
- Giustina A, Mazziotti G, Canalis E. Growth hormone, insulin-like growth factors, and the skeleton. Endocr Rev 2008;29: 535–559.
- 50. Doga M, Bonadonna S, Gola M et al. GH deficiency in the adult and bone. J Endocrinol Invest 2005; 28(8 Suppl.): 18–23.
- 51. Moller S, Becker U. Insulin-like growth factor 1 and growth hormone in chronic liver disease. Dig Dis 1992; 10: 239–248.
- Sjögren K, Liu JL, Blad K et al. Liver-derived insulin-like growth factor I (IGF-I) is the principal source of IGF-I in blood but is not required for postnatal body growth in mice. Proc Natl Acad Sci USA 1999; 96: 7088 7002
- Sjögren K, Jansson JO, Isaksson OG et al. A model for tissue specific inducible insulin-like growth factor-I (IGF-I) inactivation to determine the physiological role of liver-derived IGF-I. Endocrine 2002; 19: 249–256.
- Compston JE. Hepatic osteodystrophy: vitamin D metabolism in patients with liver disease. Gut 1986; 27: 1073–1090.

- Lloyd ME, Hart DJ, Nandra D et al. Relation between insulin-like growth factor-I concentrations, osteoarthritis, bone density, and fractures in the general population: the Chingford study. Ann Rheum Dis 1996; 55: 870–874.
- Ormarsdottir S, Ljunggren O, Mallmin H et al. Circulating levels of insulin-like growth factors and their binding proteins in patients with chronic liver disease: lack of correlation with bone mineral density. Liver 2001; 21: 123–128.
- Clemmons DR, Underwood LE, Dickerson RN et al. Use of plasma somatomedin-C/insulin-like growth factor I measurements to monitor the response to nutritional repletion in malnourished patients. Am J Clin Nutr 1985; 41: 191–198.
- Ormarsdottir S, Ljunggren O, Mallmin H et al. Low body mass index and use of corticosteroids, but not cholestasis, are risk factors for osteoporosis in patients with chronic liver disease. J Hepatol 1999; 31: 84–90.
- Holecki M, Zahorska-Markiewicz B, Więcek A et al. Obesity and bone metabolism. Endokrynol Pol 2008; 59: 218–223.
- Ostrowska Z, Kobielski A, Kos- Kudla B et al. Obesity and the relationship between somatotrophic axis and bone tissue. Endokrynol Pol 2009; 60: 302–309.
- Czerwińska E, Marcinowska-Suchowierska E, Walicka M et al. The influence of bariatric surgery on calcium homeostasis and biochemical markers of bone turnover in patients with morbid obesity Endokrynol Pol 2007; 58: 130–139.
- Marcinowska-Suchowierska E. Vitamin D: contemporary status of knowledge. Using of vitamin D in the prevention and treatment of osteoporosis. Pol Arch Med Wewn 2002; 107: 111–119.
- Crawford BA, Labio ED, Strasser SI et al. Vitamin D replacement for cirrhosis-related bone disease. Nat Clin Pract Gastroenterol Hepatol 2006; 3: 689–699.
- Lim LY, Chalasani N. Vitamin d deficiency in patients with chronic liver disease and cirrhosis. Curr Gastroenterol Rep 2012;14: 67–73.
- Diamond T, Stiel D, Lunzer M et al. Ethanol reduces bone formation and may cause osteoporosis. Am J Med 1989; 86: 282–288.
- Duarte MP, Farias ML, Coelho HS et al. Calcium-parathyroid hormonevitamin D axis and metabolic bone disease in chronic viral liver disease. J Gastroenterol Hepatol 2001; 16: 1022–1027.
- Fisher L, Fisher A. Vitamin D and parathyroid hormone in outpatients with noncholestatic chronic liver disease. Clin Gastroenterol Hepatol 2007; 5: 513–520.
- Cunningham J, Locatelli F, Rodriguez M. Secondary hyperparathyroidism: pathogenesis, disease progression, and therapeutic options. Clin J Am Soc Nephrol 2011; 6: 913–921.
- L ng KVQ, Nguy n LTH. Theoretical basis of a beneficial role for vitamin D in viral hepatitis. World J Gastroenterol 2012; 18: 5338–5350.
- Dobnig H, Turner RT. The effects of programmed administration of human parathyroid hormone fragment (1–34) on bone histomorphometry and serum chemistry in rats. Endocrinology 1997; 138: 4607–4612.
- Weir EC, Terwilliger G, Sartori L et al. Synthetic parathyroid hormonelike protein (1–74) is anabolic for bone in vivo. Calcif Tissue Int 1992; 51: 30–34.
- Jilka RL, Weinstein RS, Bellido T et al. Increased bone formation by prevention of osteoblast apoptosis with parathyroid hormone. J Clin Invest 1999; 104: 439–446.
- Monier-Faugere MC, Geng Z, Mawad H et al. Improved assessment of bone turnover by the PTH-(1–84)/Large C-PTH fragments ratio in ESRD patients. Kidney Int 2001; 60: 1460–1468.
- Crosbie OM, Freaney R, McKenna MJ et al. Bone density, vitamin D status, and disordered bone remodeling in end-stage chronic liver disease. Calcif Tissue Int 1999; 64: 295–300.
- Looker AC, Bauer DC, Chesnut CH 3rd et al. Clinical use of biochemical markers of bone remodeling: current status and future directions. Osteoporos Int 2000; 11: 467–480.
- Eriksen EF, Brixen K, Charles P. New markers of bone metabolism: clinical use in metabolic bone disease. Eur J Endocrinol 1995; 132: 251–263.