



Transforming growth factor β 1 (TGF β 1) in physiology and pathology

Transformujący czynnik wzrostu β 1 (TGF β 1) w fizjologii i patologii

Dariusz Kajdaniuk¹, Bogdan Marek¹, Halina Borgiel-Marek², Beata Kos-Kudła¹

¹Department of Pathophysiology and Endocrinology, Medical University of Silesia, Zabrze, Katowice, Poland

²Department and Clinic of Maxillofacial Surgery, Medical University of Silesia, Katowice, Poland

Abstract

This review describes precisely the consequence of TGF β 1 prevalence in the organism, and its significant influence on physiological and pathophysiological processes. Organ and tissue distinctiveness hinder unambiguous characterisation of the cytokine. However, there are constant functions of TGF β 1 inducing no controversy: it participates in foetal development, control of cell growth and differentiation, induces fibrosis and scar formation (the process of 'wound healing'), causes the suppression of immune response, is involved in angiogenesis, the development of tumours, and inflammatory processes. Thus, TGF β 1 is a multifunctional cytokine. There are three fundamental directions of its activities: I. TGF β 1 regulates cell proliferation, growth, differentiation and cells movement. II. TGF β 1 has immunomodulatory effects. III. TGF β 1 has profibrogenic effects. TGF β 1 action can be local and systemic. This review describes TGF β 1 in pathology: colitis ulcerosa, Crohn's disease, coeliac disease, diabetic nephropathy, diabetic retinopathy and diabetic foot, pulmonary hypertension, and Alzheimer's disease. TGF β 1 and its receptors are also of interest to endocrinologists. Lack of TGF β 1-dependent growth control may result in oncogenesis: papillary, follicular and anaplastic thyroid cancers, prostate, breast and uterine cervical cancer, oesophagus, gastric, colorectal and liver cancers, NSCLC, and malignant melanoma. Excessive TGF β 1 activity is an integral part of the fibrotic processes occurring in the response to injury. An increased TGF β 1 expression has been observed in patients with pulmonary, kidney, and liver fibrosis. In chronic hepatitis, the prolonged stimulation of hepatic stellate cells being the result of chronic damage to hepatocytes results in the release of profibrogenic abundant factors such as TGF β 1 and leads to the development of liver cirrhosis. The results of experimental procedures and treatment known as anti-TGF β 1 strategy acting against the fibrosis in various tissues leads to hope regarding the use of anti-TGF β 1 strategy in clinical practice. (*Endokrynol Pol* 2013; 64 (5): 384-396)

Key words: TGF β 1, TGF beta 1, TGF β 1, transforming growth factor beta 1, endocrine gland, liver, cancer, neoplasm, fibrosis, angiogenesis, physiology, pathophysiology, pathology

Streszczenie

W artykule poglądowym szczegółowo opisano konsekwencje rozpowszechnienia TGF β 1 w organizmie oraz jego wpływ na szereg procesów fizjologicznych i patofizjologicznych. Istotne odrębności narządowe i tkankowe utrudniają jednoznaczną charakterystykę tej cytokiny. Istnieją jednak stałe funkcje TGF β 1 nie wzbudzające kontrowersji: uczestniczy w rozwoju płodu, regulacji wzrostu i różnicowania komórek, indukuje proces włóknienia i bliznowacenia (proces „gojenia rany”), powoduje hamowanie odpowiedzi immunologicznej, uczestniczy w angiogenezie, w rozwoju nowotworów, w procesach zapalnych - jest więc cytokiną wieloczynnościową. Można wyróżnić trzy fundamentalne kierunki jego działania: I — TGF β 1 reguluje proliferację, wzrost, różnicowanie i przemieszczanie komórek; II — TGF β 1 wykazuje działanie immunomodulujące; III — TGF β 1 wykazuje działanie profibrogenne. Działanie TGF β 1 może mieć charakter miejscowy i systemowy. Opisano udział TGF β 1 w stanach patologicznych: wrzodziejące zapalenie jelita grubego, choroba Crohna, celiakia, cukrzyca (nefropatia, retinopatia, stopa cukrzycowa), nadciśnienie płucne, choroba Alzheimera. TGF β 1 i jego receptory są również przedmiotem zainteresowania endokrynologów. Brak zależnej od TGF β 1 kontroli wzrostu może skutkować onkogenezą: rak brodawkowy, pęcherzykowy i anaplastyczny tarczycy, prostaty, sutka, szyjki macicy, przelyku, żołądka, jelita grubego, wątroby, NSCLC, czerniak złośliwy. Nadmierna aktywność TGF β 1 jest integralną częścią procesów włóknienia zachodzących w odpowiedzi na uszkodzenie. Zwiększoną ekspresję TGF β 1 stwierdzono m.in. u chorych ze zwłóknieniem płuc, nerek i wątroby. U chorych z przewlekłym zapaleniem wątroby długotrwała stymulacja komórek gwiazdzystych będąca wynikiem przewlekłego stanu uszkodzenia hepatocytów skutkuje obfitym uwalnianiem profibrogennych czynników, w tym TGF β 1 prowadząc do rozwoju marskości wątroby. Wyniki eksperymentalnego postępowania i leczenia, określanego jako strategia anti-TGF β 1, przeciwdziałającemu procesowi włóknienia w różnych tkankach stwarzają nadzieję na jego zastosowanie w praktyce klinicznej. (*Endokrynol Pol* 2013; 64 (5): 384-396)

Słowa kluczowe: TGF β 1, TGF beta 1, TGF β 1, transformujący czynnik wzrostu beta 1, gruczoł endokryny, wątroba, rak, nowotwór, włóknienie, angiogeneza, fizjologia, patofizjologia, patologia

This work has used information and materials gathered during the implementation of the grants funded by the State Committee for Scientific Research (KBN; Poland): 3P05B05322, 3P05B03123 and published in the habilitation dissertation by D.Kajdaniuk: ISBN 978-83-7509-108-3, ISSN 1689-6262.



Dariusz Kajdaniuk MD, Ph.D., Department of Pathophysiology and Endocrinology, Medical University of Silesia, Zabrze, Pl. Traugutta 2, 41-800 Zabrze, Poland, fax: +48 32 271 26 41, e-mail: patofizj@sum.edu.pl

TGFβ1 and its receptors

The name transforming growth factor (TGF) was introduced by Moses et al. [1], who found so called transformation fibroblast stimulating factor to the cancer cell phenotype. Later it turned out that there are factors TGF α and β, and the latter also has isoforms. TGFβ's previously used names were a reflection of the first descriptions of its actions. It was called factor inhibiting differentiation, stimulating cartilage growth, and sarcoma growth factor. Now it is known that TGFβ1 activity is much wider (as described below). In turn, TGFβ2 and TGFβ3 regulate cell proliferation, growth, differentiation and migration. They participate in adipogenesis, chondrogenesis, embryogenesis, tissue remodelling, wound healing, and tumour formation. Transforming growth factor β, and 40 other proteins, including inhibin A and B, and activin A, AB, B, C, E, BMP2-15 (bone morphogenetic proteins) are included in the family of modulators of cell proliferation, differentiation and apoptosis, extracellular matrix (ECM) synthesis. These proteins play an important role in prenatal development, postnatal growth, reconstruction and maintenance of normal organs structure. TGFβ was isolated in 1978 [2–4].

TGFβ is a polypeptide constructed from 112 amino acids, encoded by a gene located on the long arm of chromosome 19. TGFβ exists in five isomeric forms marked with symbols from β1 to β5, homologous in 60–80%. TGFβ1-3 are present in humans, mammals and birds. TGFβ4-5 occur in birds and amphibians. In humans, the predominant isoform is TGFβ1, which is synthesised by almost all cells. Other isoforms are expressed in a limited spectrum of cells and tissues. TGFβ2 is synthesised in large amounts in glioma cells and keratinocytes. TGFβ3 is observed mainly in embryonic heart and lung tissue, and to a negligible extent in the liver, spleen and kidneys [5–7]. TGFβ1 is synthesised primarily by platelets, macrophages/monocytes, lymphocytes, fibroblasts, epithelial cells [8] and dendritic cells [3]. *In vitro*, TGFβ isoforms have a similar biological effect on the tissues, but *in vivo* the effect is varied [9]. *In vivo*, these isoforms show differences in the biological effects mainly conditioned by their different tissue distribution, the degree of target cells differentiation and TGFβ concentration [3]. In the liver, both healthy and with fibrosis, TGFβ1 is the most common isoform [10].

TGFβ1 is a homodimer with a mass 25 kDa. The sequence of amino acids in TGFβ1 proteins from different species are very stable, which leads to the conclusion that in the process of evolution, TGFβ has been only slightly altered, and that both in humans and in animals, its function is similar. This hypothesis is con-

firmed by the properties of TGFβ demonstrated in *in vitro* studies on human and animal cells [3, 5]. TGFβ1 is released from cells as an inactive precursor containing TGFβ1 and propeptide LAP (Latency-Associated Protein) that are connected by non-covalent linkage [11]. In this embodiment, TGFβ1 can be stored in the granules of platelets or on the cell surface. TGFβ1 is connected (through the LAP) by a disulphide bond with LTBP (Latent TGFβ Binding Protein). LTBP1-4 is a component of the ECM, and is necessary both for the synthesis of TGFβ1 and its storage [7]. Changing the conformation of LTBP by the ECM glycoprotein — thrombospondin-1 leads to the release from the complex of an active form of TGFβ [11]. In the blood, TGFβ1 occurs in an inactive form with a half life of 90 minutes. The half-life of the active form reaches only a few minutes. Thus, the LAP and LTBP 'mask' the epitopes of TGFβ1, and the active form is almost undetectable in the blood and tissues (regardless of the method). In the initial phase of TGFβ1 activation, a tissue transglutaminase is involved. TGFβ1 release from the inactive complex occurs during its proteolysis under the influence of plasmin and thrombin [7]. It is believed that certain factors regulate TGFβ1 activity by increasing its synthesis, and others by increasing its bioavailability. TGFβ1 synthesis is stimulated by physical, mechanical, and biochemical factors [12]. Increased TGFβ1 activity has been observed in response to angiotensin II [13–16], LDL, glucose, thromboxane A2. Natural inhibitors of TGFβ1 are follistatin, decorin, and α2-macroglobulin [12]. In the organism, a series of interactions between the ligands TGFβ and their natural inhibitors occur.

TGFβ signalling activity in the cells is possible only when they have specific membrane receptors which occur as dimeric proteins [6]. So far, nine different types of molecules (receptors and proteins) having the ability to bind TGFβ have been identified. The best known receptors are types I, II, and III [11]. The operation of all TGFβ isoforms is done by these three types of receptors for TGFβ — TGFβRI, TGFβRII, and TGFβRIII [17]. TGFβRI is a dimer of molecular weight 53 kDa, dimer TGFβRII weight 75 kDa, and TGFβRIII has a mass 280 kDa [17, 18]. TGFβRI and TGFβRII are the family of transmembrane receptors, their intracellular fragments have domains with serine-threonine kinase activity [19]. The extracellular part of TGFβRII by binding the ligand (TGFβ) activates the intracellular domain of the receptor. The created complex joins TGFβRI that determines specificity of TGFβ recognition [20]. This heteromeric complex is formed by two molecules (dimers) TGFβRI, TGFβRII and TGFβ [18, 21–24]. Activated TGFβRII kinase phosphorylates serine fragments of sequence TTSGSGS in GS domain (domain rich in glycine and serine) of TGFβRI, thus leading to activation of serine-

threonine kinase in the receptor, and thereby starts the signal transduction cascade inside the cell [2, 11, 18, 21–27]. TGF β RII can indeed bind TGF β independently of the presence of the TGF β RI, but without TGF β RI is unable to transduce a biological signal [28]. On the other hand, in the absence of TGF β RII, the cells are insensitive to the action of TGF β [29]. TGF β RII is activated in the process of autophosphorylation, while activation of TGF β RI requires a connection to complex of TGF β 1-TGF β RII, -TGF β RIII [19]. TGF β 1 and TGF β 3 have a high ability to bind with TGF β RII. TGF β 2 exerts its effect only after a previous presentation by TGF β RIII to the other receptors I and II [30, 31]. TGF β RIII is called a betaglycan, which is anchored in the membrane proteoglycan having no enzymatic activity [19], without exerting any intracellular activity. Its role is to present TGF β to the other two receptors. TGF β RIII is the most abundant receptor subtype in most cells [32, 33]. TGF β RIII is located on the cell surface binding the ligand [34] — it has a high affinity for all three isoforms of TGF β . In its absence, the cells do not respond to TGF β 2, whereas the response of cells to the other two isoforms of the cytokine is maintained [27]. The action of TGF β in hepatic stellate cells (HSC) can be modulated by TGF β RIII [34]. In these cells, it is additionally important because the expression of TGF β 2 here is low [35]. TGF β RIII may also be an inhibitor of the signal transduction by prevention of TGF β connections to TGF β RII and TGF β RI in the mechanism independent from ligand binding. In such a situation, it fulfills the function of regulating the ligand binding with the remaining two receptors [36]. TGF β RIII's soluble form, by binding the ligand, neutralises it [34]. The final effect of TGF β 1 depends on its bioavailability, the distribution of receptors in tissues, and the target cell type [12, 37].

After binding TGF β with receptors, signal transduction occurs to the nucleus with the participation of proteins Smad present in the cytoplasm, which ultimately results in the influencing of the processes of transcription and translation of TGF β -dependent genes i.e. showing typical for TGF β activity [18]. System of intracellular transmitters Smad proteins is thus a specific link between receptors and the cellular nucleus [2, 38–40]. After activating serine-threonine kinase in TGF β RI, it comes to further signal transduction by phosphorylation of cytoplasmic proteins Smad2 [23] and Smad3 [23, 41] binding to receptors. Similar in the structure Smad2 and Smad3 are classified as a group called R-Smad (receptor regulated Smads). Phosphorylated R-Smad can be separated from the connection to the receptor and from protein SARA [23]. Auxiliary protein SARA (Smad Anchor for Receptor Activation) is a cytoplasmic protein anchored in the cell membrane that binds both the R-Smad and TGF β /

/TGF β RII/TGF β RI. SARA participates in the early stages of signal transduction — recognising the non phosphorylated R-Smad joins it to the newly formed receptor complex, and is dissociated itself [42]. Phosphorylated R-Smad forms a complex with the co-Smad (common partner Smad) namely Smad4, and so the newly formed complex is transported to the nucleus [26, 42], where it regulates the transcription of TGF β -dependent genes [43]. Thus, phosphorylated Smad act as transcription factors, which by binding to specific DNA sequences are responsible for the transcription of specific genes [2, 11, 19, 41]. Smad4 co-operates with other transcription factors in the regulation of TGF β -dependent gene expression — therefore it is a grasping point for these factors. Signal transduction from the receptor to the nucleus can be inhibited by a protein Smad6 and Smad7 called I-Smads (Smads inhibitors), which despite the absence of motif in its structure which can be phosphorylated by receptor kinases, are able to interact with membrane receptors, thus impairing their interaction with the R-Smad [26]. Smad7 expression is induced by TGF β , leading to inhibition of the cellular response to this cytokine. It is therefore an autoregulation in the negative feedback mechanism [26, 42]. Smad7 forms a stable complex with activated TGF β RI and thus impairs the phosphorylation of Smad and inhibits signalling cascade [38, 43, 44]. There are known also proteins Smad 1, 5, 8, which transmit impulses from other ligands related to TGF β , including BMPs [23]. Overall in the Smad system, ten different proteins carrying a signal have been identified which were activated by the TGF β -receptor complex RI-III [2]. Under disease conditions, Smads also interact with other signalling pathways, such as the mitogen-activated protein kinase and nuclear factor- κ B pathways [45].

TGF β 1 in physiology and pathophysiology

The consequence of TGF β 1 prevalence in the organism is its significant influence on a number of physiological and pathophysiological processes. As a result of this, interrelations with other cytokines and biologically active substances as well as organ and tissue distinctiveness hinder unambiguous characterisation of TGF β 1. However, there are constant functions of the cytokine inducing no controversy: it participates in foetal development, control of cell growth and differentiation, induces fibrosis and scar formation (the process of 'wound healing'), causes the suppression of immune response, is involved in angiogenesis, the development of tumours, and inflammatory processes [6, 11, 42, 46–48]. Thus, TGF β 1 is a multifunctional cytokine.

There are three fundamental directions of its activities:

I. TGFβ1 regulates cell proliferation, growth, differentiation and cells movement. Growth factors/cytokines may affect stimulating or inhibiting on the cell proliferation and differentiation by linking to specific receptors, which triggers a cascade of signals leading to the activation or repression of various genes. Growth factors acting on cells in the resting state or in the G₀ phase introduce them into the cell cycle. Growth factors called 'competence factors' bring the cells into the G₁ phase and conduct them through this phase, while under the influence of growth factors termed 'progressive factors' DNA synthesis occurs. Passing through the G₁ phase requires stimulation by growth factors throughout its duration (a few hours). If this stimulating signal is broken, the cell returns to the G₀ phase. In G₁ phase, there is a critical point at which the simultaneous operation of 'competence' and 'progressive' factors is necessary. Then, only the presence of the latter is required [47–49]. It has been shown that TGFβ1 inhibits the cell cycle in the G₁ phase, which explains the fact that this cytokine is a potent inhibitor of cell proliferation [11]. In addition to TGFβ1, interferon and tumour necrosis factor (TNF) can antagonise the pro-proliferative effect of growth factors. In the case of TGFβ1, such an effect is noticeable even in the late G₁ phase [49]. The regulatory impact of TGFβ1 on quantitative cellular changes is the result of, on the one hand, the possibilities of inhibition of cells proliferation, and on the other hand, the induction of apoptosis. The action of TGFβ1 results in decline of Fas expression and in increased expression of Bcl-2 [12]. Thus, TGFβ1 is an endogenous factor controlling apoptosis in normal and pathological tissues, and thereby is a factor controlling the balance between replication and cell death [50]. In certain cell types, the absence of growth factors alone induces their apoptosis [49]. TGFβ1 plays a role in regeneration (repair — 'wound healing') of damaged tissues and organs, bone remodelling, malignant processes (tumour formation, metastasis), and the generation of histopathological lesions like fibrosis [4, 7, 11]. It is involved in organogenesis during intrauterine life (embryogenesis), hematopoiesis, and adipogenesis. It promotes chondrogenesis and angiogenesis [11]. It is involved in skin formation. TGFβ1 operates differently on the target cells, and this effect depends on the target cell type [51, 52]: TGFβ1 usually stimulates the proliferation of certain cell types of mesenchymal origin such as hepatic stellate cells, fibroblasts and osteoblasts both *in vitro* and *in vivo* [4, 51, 53]. TGFβ1 (and TGFβ2) intensifies the Schwann cells proliferation [53]. During bone resorption, osteoclasts release and activate TGFβ1 from the bone matrix; thus, elevated bone resorption

increases the level of active TGFβ in the local environment during ageing [4]. TGFβ1 inhibits the growth of epithelial cells (including the intestine), endothelial, hematopoietic (including megakaryocytes and erythrocyte precursors), keratinocytes [7, 11, 51, 54, 55]. TGFβ1 is a potent inhibitor of the hepatocytes proliferation *in vitro* and *in vivo* - in humans and animals [6, 42, 55, 56]. The degree of inhibition of cell proliferation by TGFβ1 depends among others on the cells types, this cytokine concentration, and its interactions with biologically active substances. For example, the most sensitive to the inhibitory effect of TGFβ1 are lung epithelial cells and keratinocytes. TGFβ1 in a concentration of 1–2 fg/cell inhibits the growth of smooth muscle cells, fibroblasts, and chondrocytes; in higher concentrations, it has the opposite effect - it stimulates the growth of these cell types [53]. TGFβ1 plays a role in tissue regeneration, on the one hand by promoting cell differentiation [57], and on the other by inhibiting excessive cell proliferation [58, 59]. TGFβ1 is an important factor inducing apoptosis of liver cells *in vitro* and *in vivo* [42, 60]. TGFβ1 inhibits liver regeneration in rats, in contrast to hepatocyte growth factor (HGF) [61] and transforming growth factor α (TGFα) [61, 62]. TGFβ1 inhibits hepatocyte growth even in the healthy liver and thus may contribute to the maintenance of a constant mass of the organ. That has been demonstrated by introducing TGFβRII into the liver cells of healthy rats (adenoviruses were the vectors) which led to the inhibition of DNA synthesis in the liver [63]. In certain tissues and organs, TGFβ1 promotes repair processes caused by trauma or pathological lesions. In the repair processes TGFβ1 stimulates fibroblast migration to sites of injury and stimulates the synthesis of ECM proteins [57] as fibronectin, collagen, and proteoglycans. These proteins are the ligands for integrins i.e. surface receptors of the cells involved in the response to tissue injury which means that they can move to the place of their action [7, 54]. Under physiological conditions, TGFβ1 protects the intestinal mucosa from damage [7]. The intestinal mucosa revealed the presence of TGFβ1-3 and their receptors I–III [64]. *In vitro* TGFβ1 inhibits the proliferation of intestinal epithelial cells and promotes their differentiation [57, 59]. TGFβ1 synthesised by enterocytes and cells of the lamina propria colonic mucosa [64] plays here a role in regulation of proliferation, differentiation and apoptosis [2]. Despite the properties of inhibiting the proliferation of epithelial cells, in the case of damage of the intestinal mucosa TGFβ1 promotes the healing process. It is also possible due to the ability to induce and encourage migration of mature, functionally efficient epithelial cells from the environment which leads to cover the wound within a few hours [7, 54]. TGFβ1 also acts as an inhibitor of atherosclerosis — inhibits the prolifera-

tion of smooth muscle and endothelial cells. Reduction of TGF β RII expression and its mutations have been shown in populations of cells from atherosclerotic lesions [3]. TGF $\beta 1$ and TGF $\beta 2$ promotes repair of experimentally caused skin injuries in rats which occurs with excessive exposure of fibrous tissue [65]. Under physiological conditions TGF $\beta 1$ and other growth factors contained in the tear film are responsible for the normal state of the cornea [66].

II. TGF $\beta 1$ has immunomodulatory effects. It is involved in the suppression of the immune response [6, 42, 67]. It inhibits the proliferation, differentiation and activity of cells involved in humoral and cellular responses, reduces the expression of MHC molecules, the cellular toxicity, the production of antibodies, and inhibits the secretion of cytokines [68]. On the surface of B cells are located receptors for TGF $\beta 1$ — their stimulation leads to inhibition of B cell proliferation and production of immunoglobulin G and M (IgM, IgG) and to increased IgA secretion [53, 69, 70]. TGF $\beta 1$ inhibits T cell proliferation [53] in response to polyclonal mitogens [71]. On the other hand, in the inflamed tissues an increase of activation of T helper cells CD4 + which produce TGF $\beta 1$ is seen [70, 72]. This cytokine exerts an inhibitory effect on some of the NK cells functions [69] e.g. on their cytotoxic activity [53]. In addition, it inhibits macrophages' maturation and activity [53, 67, 71]. Environmental conditions can determine the reactivity of the cells to TGF $\beta 1$ e.g. activated T cells do not react with TGF $\beta 1$ in the absence of interleukin (IL) -10 [73]. TGF $\beta 1$ is required for the maintenance of immune homeostasis. TGF $\beta 1$ by reducing apoptosis of B cells leads to prolongation of their life and elongation of long-term immunological memory [74]. The immunosuppressive and anti-inflammatory effects of TGF $\beta 1$ [42] have been confirmed *in vitro* [11, 67, 75] and *in vivo* [11, 75]. In mice, lack of TGF $\beta 1$ gene causes multiple organ inflammatory response, and lethal cachexia develops within two weeks [76]. In transgenic mice lacking the TGF $\beta 1$ gene (knock out), inflammation of the small intestine and colon develops [59]. In transgenic mice with Smad7 overexpression glomerulonephritis develops caused by antibodies against the basement membrane [77] and inflammation of the respiratory tract [38]. In transgenic mice lacking Smad3 gene, a massive inflammation of the gastrointestinal tract was detected [44]. An excessive suppressive effect of TGF $\beta 1$ on the the immune system increases susceptibility to infection [53]. Mouse model studies have demonstrated that TGF $\beta 1$ is involved in the pathogenesis of autoimmune diseases — systemic administration of TGF $\beta 1$ suppress autoimmune disease while the anti-TGF $\beta 1$ antibodies caused its progression. It has been demonstrated that mutations in the TGF $\beta 1$ gene result in the development of phenotype with characteristics typical of autoimmune diseases [3].

III. TGF $\beta 1$ has profibrogenic effects. This activity is carried out through the ability for stimulating the ECM synthesis which has fundamental importance for the processes of scarring and tissue reconstruction [6]. TGF β through increasing the production of ECM proteins and their receptors [6, 26] becomes the strongest inducer of the synthesis of collagen type I, II, III, V, VI, X and fibronectin, osteopontin, osteonectin, thrombospondin, proteoglycans and alkaline phosphatase [26]. Some of them, such as fibronectin and tenascin, are absent at physiological conditions. TGF $\beta 1$ has profibrogenic action via stimulation of mesenchymal cells and fibroblasts to synthesise ECM proteins [68]. TGF $\beta 1$ stimulates the proliferation and/or synthesis of ECM components in cultures of fibroblasts obtained from skin [8], lung [78] and stimulates collagen synthesis in pancreatic [8] and liver [79] fibroblasts. It simultaneously blocks the destruction of newly synthesised ECM by inhibiting the synthesis of matrix-metalloproteinases (MMPs) — enzymes involved in the degradation of ECM proteins - and by increasing the expression of genes responsible for the production of tissue inhibitor of metalloproteinases (TIMPs) — TIMP-1 [80] and plasminogen activator inhibitor-1 (PAI-1). TGF $\beta 1$ exacerbating the expression of PAI-1 reduces the conversion of plasminogen to plasmin — protease that directly degrades ECM proteins and activates MMPs [26, 41]. TGF $\beta 1$ mitogenic effect on fibroblasts is mediated by connective tissue growth factor (CTGF) — a cytokine that promotes fibrosis in the skin, lungs, kidneys, and liver [81, 82, 83]. Furthermore, TGF $\beta 1$ stimulates the production and release of vasoactive factors such as endothelin I, nitric oxide (NO), C natriuretic peptide, PGE2 which may also affect the ECM production and the proliferation of ECM-producing cells. Finally, TGF $\beta 1$ potentiates its own production and thus biological activity [68]. TGF $\beta 1$ stimulates the secretion of cytokines such as vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF), platelet-derived growth factor (PDGF), and IL-6. After repairing the damaged tissue, the release and biological activity of TGF $\beta 1$ are ruptured - this occurs in the acute process. In chronic processes, the excessive and persistent production of TGF $\beta 1$ occurs which leads to progressive fibrosis [7, 47, 48, 68]. Synthesis and profibrogenic action of TGF $\beta 1$ are intensified by angiotensin II which is therefore another mediator of ECM production in the liver [13, 14–16]. TGF $\beta 1$ is one of the key mediators of fibrogenesis.

TGF $\beta 1$'s action can be local and systemic. TGF $\beta 1$, released locally as a result of injury or the immune response, has pro-inflammatory properties (e.g. it stimulates granulocyte macrophage chemotaxis and the release of pro-inflammatory cytokines (TNE, IL-1, IL-6) [7, 47, 48] and profibrogenic properties

[7, 54]. Among the systemic properties of TGF β 1, an immunosuppressive effect seems to be the most important [7, 47, 48].

TGF β 1 in pathology

Elevated TGF β 1 plasma levels and a positive correlation of levels with the degree of damage to the intestinal mucosa have been shown in patients with colitis ulcerosa [84]. These values were increased slightly during treatment; after successful treatment confirmed by clinical and endoscopic activity, index returned to the normal range [59]. Although the quoted results suggest TGF β 1 participation in the processes of repair intestinal tissues that were previously destroyed in autoimmune mechanism, it cannot be excluded that TGF β 1 is involved in the initiation of the disease process [84]. Crohn's disease, in which the inflammatory process covers the entire wall of the intestine, and repair processes can lead to the formation of solid wall stenosis, higher levels of TGF β 1 expression in intestine miofibroblasts were found significantly in patients with a form of the disease with strictures extending [85]. There was a positive correlation between the TGF β 1 mRNA expression in the intestinal mucosa and the severity of the inflammatory response [5]. In children with coeliac disease, low TGF β 1 expression in intestinal epithelium was found [86]. Antibodies against endomysium (EmA) are routinely determined in patients with coeliac disease; in fact, there are anti-transglutaminase antibodies which change the complex of TGF β 1 inactive to an active one. It is known that circulating antibodies against transglutaminase may lead to a deficiency of TGF β 1 and hence to the development of autoimmunity. This may be the explanation for the tendency of patients with coeliac disease to develop autoimmune diseases [7]. In 79% of children with allergic enteropathy, a decreased TGF β 1 mRNA expression in intestine epithelium was shown [87]. Reduced TGF β 1 levels in the inflamed duodenum [88] and accelerated healing of gastric ulcers after injection of TGF β 1 in the ulcer area [89] have been shown in the immunohistochemical assessment. TGF β 1 is involved in the development of kidney diseases such as fibrosis, diabetic nephropathy, and glomerulosclerosis [68]. Serum concentrations of TGF β 1 may be an additional parameter in predicting the occurrence of diabetic retinopathy in juvenile patients with type 1 diabetes mellitus [90]. In 95% of children with atopic dermatitis, specific TGF β 1 gene polymorphism coexisted with decreased TGF β 1 production [91]. In patients with diabetic foot and leg ulcers due to chronic venous insufficiency, locally reduced levels of TGF β 1 and its receptors have been shown [92]. In patients with resistant-to-treatment leg ulcers, a reduced TGF β RII expression in fibroblasts has been demonstrated [93]. In asthma, reduced

TGF β RII expression has been demonstrated which may be one of the mechanisms leading to thickening of the bronchial basal lamina [94, 95]. Severe pulmonary inflammation induced by elevated levels of cytokines (therein TGF β 1), combined with increased viral replication due to decreased interferon- γ levels, may contribute to worsening respiratory symptoms in patients with bronchial asthma and A (H1N1) 2009 infection [96]. Mutations in the genes for TGF β receptors are the causative agent of Rendu-Osler-Weber disease, vascular dysplasia with autosomal dominant inheritance [3]. Discontinuation of TGF β 1 activity is also in hereditary chondrodysplasia, and pulmonary hypertension [23]. Decreased serum levels of the angiogenic factors VEGF and TGF β 1 in Alzheimer's disease and amnesic mild cognitive impairment. These observations suggest that angiogenesis might be involved in the onset process of Alzheimer's disease and the decrease of angiogenic factors might be related to the severity of cognitive impairment [97].

TGF β 1 in endocrinology and oncology

TGF β 1 (and other growth factors) and its receptors are also of interest to endocrinologists [47, 48, 72, 98–102]

TGF β isoforms are present in the front lobe of the pituitary, where they modulate growth and secretory function of cells. In the pituitary cell lines, TGF β 1 stimulates VEGF production in a dose-dependent manner and dependent on the type of target cells [103]. Among the pituitary cells, lactotroph cells produce the most TGF β 1 and contain too the most TGF β RII. TGF β 1 inhibits lactotroph cells proliferation and PRL secretion. Synthesis of TGF β 1 in lactotroph cells is reduced during the oestrogen administration in ovariectomised rats. During the oestrogen administration, decreased mRNA TGF β RII expression in oestrogen-sensitive tissues is found which additionally influences the lactotroph cells growth and prolactin secretion. The development of pituitary tumours in mice has been associated with resistance to TGF β [104]. TGF β 1 could be a potential serum marker for invasiveness of prolactinomas — the simultaneous determination of TGF β 1 and PRL levels could improve the noninvasive assessment of prolactinoma behaviour [105]. TGF β 1 is thought to have important roles in several stages of folliculogenesis [106]. Maurya et al.'s [107] study demonstrates the importance of the liberation of biologically active TGF β 1 from its latent complex during embryo implantation period and its regulation by oestradiol [107]. TGF β 1 and its receptor soluble endoglin are altered in polycystic ovary syndrome during controlled ovarian stimulation [108].

Lack of TGF $\beta 1$ -dependent growth control may result in oncogenesis [6, 11, 109]

The changed expression of growth factors and their receptors is an element of neoplastic transformation and tumour progression [110]. In normal conditions, TGF $\beta 1$ is a potent inhibitor of the growth of many cell types, including neoplastic [11]. In the early stages of cancer development, its cells respond to antimitotic effect of TGF $\beta 1$ [18]. However, at the entry of tumour cells into the phase of uncontrollable growth, most of them lose sensitivity to the inhibitory effect of TGF $\beta 1$. It is surprising that this occurs despite the presence on the tumour cell surface of the receptors for TGF $\beta 1$. What's more, these cancer cells begin to secrete TGF $\beta 1$ themselves [11]. The TGF $\beta 1$ -dependent immunosuppressive activity, stimulating angiogenesis [11, 18, 47, 48], increasing the affinity of cancer cells to cell adhesion molecules [11] creates a microenvironment favourable to tumour growth and its metastasis — increases cancer cells invasiveness [18]. Additionally, TGF $\beta 1$ induces the death of the surrounding healthy cells and thus eliminates their effect designed to inhibit tumour growth [50]. It appears that cancer cells to receive anti-mitotic stimulus carried by TGF $\beta 1$ need higher TGF $\beta 1$ concentrations than normal cells. On the other hand, higher TGF $\beta 1$ concentrations have more potent anti-mitotic and pro-apoptotic effects on tumour stromal cells and have a more immunosuppressive effect on the environment and strongly stimulate angiogenesis in the environment. The results of clinical and experimental studies indicate that the molecular reasons for the lack of cell response to TGF $\beta 1$ during malignant transformation are caused by mutation in the TGF β RII receptor [111] and/or within the intracellular proteins Smad2, Smad3, Smad4 [39] responsible for providing information to the nucleus or Smad7, which is an inhibitor of TGF $\beta 1$ activation [38, 44].

In the tissues of papillary thyroid carcinoma (PTC) follicular and anaplastic thyroid cancers, zero or reduced TGF β RII expression has been found compared to benign tumours and normal tissues (in immunohistochemical studies with anti-TGF β RII) [112]. In the differentiated and undifferentiated tissues of the thyroid cancers, resistance to TGF $\beta 1$ coexisted with a reduction in mRNA and protein TGF β RII expression [113]. TGF $\beta 1$ mRNA expression in papillary thyroid cancer cells was higher compared to the surrounding tissues, while the TGF β RII was lower. An inverse correlation between TGF β RII and tumour size was found, and there was no such correlation with respect to TGF $\beta 1$, which suggests that primarily TGF β RII plays a role in the pathogenesis of papillary thyroid cancer [114]. Metastatic thyroid cancer can also be characterised by a decreased sensitivity to the action of TGF $\beta 1$ [115]. In

human PTC, higher expression levels of TGF $\beta 1$ were closely related with lymph node metastasis, whereas for α -smooth muscle actin (α -SMA) and Smad3 expression increased significantly with advanced tumour stages. Moreover, a significant correlation was found between higher TGF $\beta 1$ expression in PTC cells and increased α -SMA levels in the fibroblasts surrounding the cancer cells. These findings suggest that the activation of TGF β /Smad3 pathways in cancer cells influence tumour growth [116].

Prostate cancer is characterised by a loss of expression of receptors for TGF β and is resistant to anti-proliferative and pro-apoptotic activity of TGF $\beta 1$. In humans, high expression of TGF $\beta 1$ and the lack of expression of its receptors in prostate cancer tissues are associated with a particularly poor prognosis [117], and TGF β RII overexpression restores sensitivity to TGF $\beta 1$ and transmission of signals in the cancer cells. TGF $\beta 1$ *in vitro* is, however, an inhibitor of breast cancer cells proliferation. Progression of tumour *in vivo* can be taken from the fact that TGF $\beta 1$ secreted by the tumour may lead to suppression of immune response and to an enhancement of angiogenesis in the tumour environment [55]. The polymorphisms in the genes related to angiogenesis - PAI-1 (675 4G/5G) and TGF $\beta 1$ (G-800A), segregated solely or combined, might contribute to the increased susceptibility to uterine cervical cancer [118].

TGF β RII loss of gene expression occurs in primary oesophagus cancer. Mutations being the microsatellite instability within the TGF β RII in gastric cancer tissues have been found [119] and they coexisted with mutations of the p53 gene [120]. In colorectal cancer tissue, the presence of mutations in the TGF β RII (> 10–20% cases) [2], microsatellite instability within the receptor (60–90% cases) [111] and mutations in the Smad (10% cases) have been revealed [40]. *In vitro* colon cancer cells exhibit increased synthesis of TGF $\beta 1$ concomitant with uncontrolled proliferation and constant growth of the tumour mass [11]. In colon cancer, abnormal TGF β RI expression has been also described [3]. There is evidence that TGF $\beta 1$ -induced disruption of apoptosis as well as growth inhibition is an integral part of a multi-step process of developing HCC (primary liver cancer, hepatocellular carcinoma) [121]. TGF $\beta 1$ induces apoptosis of normal liver cells [42, 122] including hepatocytes through the autocrine way. However, HCC cells are resistant to TGF $\beta 1$ which generates through the paracrine way TGF $\beta 1$ -mediated apoptosis of cells neighbouring the tumour, which facilitates its expansion *in situ* [122]. TGF β RII mutation is the cause of the lack of HCC cells response to TGF $\beta 1$ [123]. Immunohistochemical analysis revealed increased expression of TGF $\beta 1$ [124, 125] and lower TGF β RII expression in HCC tumour tissues compared to their surrounding tissues,

including normal [125]. Recently, in an experimental study, HCC cells were divided into those in which the expression of TGF β 1 comes late, which was associated with their increased invasiveness and shorter survival of mice, and those in which the expression of TGF β 1 appeared early, and where the prognosis was better [126]. In relation to infection with hepatitis B virus (HBV) and hepatitis C virus (HCV), the TGF β 1 participation in malignant transformation and HCC progression were highlighted [124, 127]. TGF β 1 expression in HCC tissues was correlated with the degree of HBV replication and did not correlate with sizes and number of the tumours. Patients with HCC in the course of HBV infection demonstrated higher TGF β 1 blood levels compared to controls (non-cancerous liver diseases) [124]. Higher TGF β 1 serum concentrations in HCC patients were found compared to patients with chronic hepatitis C. In HCC patients, there was no difference in the TGF β 1 levels between one and other genotypes of HCV [128]. The TGF β 1 concentration in blood [129, 130] and expression of its mRNA in the liver [130] were significantly higher in patients with HCC compared to patients with chronic hepatitis, liver cirrhosis and healthy subjects. The plasma TGF β 1 levels after treatment resulting in clinical improvement, although decreased, however did not correlate with α -fetoprotein [129]. It has even been suggested that in patients with HCC and low production of α -fetoprotein, a finding of increased TGF β 1 concentration in urine may be useful in the diagnosis [131]. In addition, the way of Smad3 phosphorylation, resulting in the creation of two possible isoforms, alters the final effect of TGF β 1. It has been demonstrated that in patients with chronic hepatitis C comes to the formation such Smad3 isoform which is able to change the way of TGF β 1 signal transduction from one that inhibits the tumour development to one that amplifies the fibrogenesis and increases the risk of developing HCC [41].

The expression of TGF β 1 and TGF β RI-III were twice higher in well-differentiated non-Hodgkin's lymphoma compared to low-differentiated [71]. In some forms of leukemias, the malignant cells have no receptors for TGF β RII and TGF β RI that causes, that they are not sensitive to the inhibitory effects of TGF β 1 and resulting from this the excess of TGF β 1 leads to inhibition of proliferation of normal cells. In addition, the overproduction of TGF β 1 by leukemia cells, monocytes and megakaryocytes is the cause of bone marrow fibrosis. TGF β RII gene mutations or lack of the gene expression in some T-cell leukemias and disorders in TGF β RI gene expression in chronic lymphocytic B-cell leukemia and in cutaneous form of T-cell lymphoma have been described. TGF β RII gene mutations or lack of the gene expression have also been described in lung cancer

tissues [3], and reduced TGF β RII expression (but not TGF β RI) may participate in urethane-induced carcinogenesis in the lungs of mice [132]. TGF β 1 genotypes polymorphisms (rs1800469, rs1982073) could be useful for predicting distant metastasis-free survival in patients with inoperable non-small cell lung cancer (NSCLC) treated with definitive radiation therapy [133]. TGF β -induced expression of IGFBP-3 regulates IGF-I receptors signalling in human osteosarcoma cells [134]. Bronchioloalveolar invasion in NSCLC is associated with tumours expression of TGF β 1 assessed with immunohistochemical staining using anti-TGF β 1 antibody [135]. It was also found a higher concentration of TGF β 1 (4.2 x) and TGF β 2 (1.5 x) (but not TGF β 3) in the blood of patients with metastatic malignant melanoma in contrast to the initial development phase which indicates a systemic immunosuppressive TGF β 1 activity in the terminal stage of the disease [136].

TGF β 1 and fibrosis

The relationship between TGF β 1 and fibrosis of various tissues and organs is increasingly being described. Although TGF β 1 plays a critical role in tissue repair, overproduction of the cytokine can lead to an excessive, uncontrolled depositing fibrous tissue [65]. Each of the processes participating in the 'wound healing' (hemostasis platelet, influx of inflammatory cells and fibroblasts, formation of ECM and blood vessel) is associated with TGF β 1 that not only stimulates the formation of scar tissue but also reduces the production of ECM-degrading enzymes. Moreover, the cells involved in 'wound healing' are stimulated to produce TGF β 1 which multiplexing effect of its actions. Overexpression of TGF β 1 caused by chronic and repetitive injuries of tissues or dysfunction of regulation the expression of this cytokine is a major factor in the pathogenesis of organ fibrosis [3, 47, 48]. An increased TGF β 1 expression was observed among others in patients with pulmonary, kidney, and liver fibrosis [11] and with scleroderma [3]. In patients with pulmonary fibrosis, the coexistence of the TGF β 1 gene polymorphism with an increased synthesis of this cytokine and intensification of ECM formation has been shown [137]. Induction of renal fibrotic genes by TGF β 1 requires epidermal growth factor receptor (EGFR) activation, p53 and reactive oxygen species [138]. In patients with scleroderma, the enhanced response to TGF β 1 caused by Smad7 mutations also results in intensified synthesis of fibrous tissue [38]. In patients with chronic hepatitis, the prolonged stimulation of hepatic stellate cells being the result of chronic damage to hepatocytes results in the release of profibrogenic abundant factors as TGF β 1 and leads to the development of liver cirrhosis. TGF β 1 leads to the ECM accumulation in the

mechanism: 1) directly increasing the synthesis of ECM components as procollagen 1 α (I), 2) inhibition of tissue collagenases expression, 3) increasing synthesis of ECM-degrading enzyme inhibitors (as PAI-1, TIMPs) [41, 139, 140]. In chronic hepatitis, hepatocyte proliferation inhibitory effect of TGF $\beta 1$ is attached to the above-mentioned mechanisms [141]. TGF $\beta 1$ is a link between hepatic inflammatory and fibrosis processes [41, 47, 48]. Benzoubir et al. [142] presented a paradigm where HCV may be related to liver pathogenesis through its ability to induce a local, intrahepatic TGF β activation. They argue for a dual impact of HCV core on liver fibrosis and liver carcinogenesis: HCV core could act both as autocrine and paracrine factors modulating TGF β responses within hepatocytes and in stromal environment through TGF β activation [142].

Progress in understanding the phenomena being arranged in a network of regulatory processes associated with organ fibrosis facilitates the use of currently known drugs as the search for new drugs targeting various stages of fibrogenesis. These drugs, used regardless of the aetiology of disease, could lead to arrest or even regression of fibrosis. Excessive TGF $\beta 1$ activity is an integral part of the fibrotic processes occurring in the response to injury. The results of experimental procedures and treatment known as anti-TGF $\beta 1$ strategy acting against the fibrosis in various tissues including liver, lung and kidney create hope regarding the use of anti-TGF $\beta 1$ strategy in clinical practice [38, 143]. Since in patients with chronic hepatitis B and C the activation of TGF $\beta 1$ -receptors system is observed, the goal of therapy of anti-TGF $\beta 1$ could be any of the system components. For this, antibodies against TGF $\beta 1$, TGF $\beta 1$ soluble receptors, substances blocking TGF $\beta 1$ receptors and binding TGF $\beta 1$ have been used experimentally. These substances used *in vivo* often do not achieve a sufficient concentration in the target cells or produce extrahepatic side effects. The specific drug carrier which would give it to hepatic stellate cells is still being sought [144]. The introduction of TGF β RII (using adenovirus) that does not transmit signals to the liver in rats has significantly reduced the extent of experimentally induced fibrosis [145, 146]. Similarly, liver fibrosis caused by biliary damage was significantly inhibited as a result of inducing the formation of complexes Fc domain of immunoglobulin G and extracellular part of TGF β RII which gives the effect of binding the receptor [147]. Promising results have been obtained on the same path in experimentally induced glomerulonephritis concomitant with renal fibrosis [148]. The Ling et al. [149] in rat model demonstrates that murine neutralising TGF β monoclonal antibody 1D11 can reverse pre-existing hepatic fibrosis induced by extended dosing of thioacetamide. The regression of fibrosis (evaluated using hepatic TGF $\beta 1$ mRNA, tissue hydroxyproline,

plasminogen activator inhibitor 1: PAI-1) was accompanied by a marked reduction in concomitantly developed cholangiocarcinomas. This suggests that therapeutic dosing of a TGF β antagonist can diminish and potentially reverse hepatic fibrosis and also reduce the number and size of attendant cholangiocarcinomas. Growing knowledge about the routes of intracellular signalling TGF $\beta 1$ [23, 24] leads to the development of further ways of interrupting the signals cascade in hepatic stellate cells, and the results cited above by Dooley et al. [43] indicate that Smad7 analogues could be transposed into anti-fibrosis treatment. In contrast to the pathogenic role of active TGF $\beta 1$, latent TGF $\beta 1$ plays a protective role in renal fibrosis and inflammation. TGF β /Smad signalling plays a regulating role in microRNA-mediated renal injury. Thus, targeting TGF β signalling by gene transfer of either Smad7 or microRNAs into diseased kidneys has been shown to retard progressive renal injury in a number of experimental models [45]. *In vivo* Smad7 has the same effect as the soluble receptors TGF β RII used in experimental models of liver fibrosis [147, 150]. The application of TGF β RII acting as false transmitters in rats [145, 150] and soluble TGF β RII in mice [151] with liver fibrosis were associated with improvements in outcome aspartate aminotransferase, alanine aminotransferase and bilirubin (which was not observed in relation to the applied Smad7 [43]). However, we do not know what the long-term consequences of Smad7 overexpression in hepatocytes are. The Smad7 overexpression can lead to neoplastic transformation. There are hopes that such risk might be overcome by using suitably modified adenoviruses. On the other hand, antagonists of TGF β signals might find application only in chronic liver injury because in acute failure TGF $\beta 1$ triggers a cascade of signals Smad2-4 simultaneously with the activation of Smad7 which in negative feedback mechanism inhibits the transmission of signals. This results in a transient production of ECM involved in tissue reparation. In chronic liver injury, cascade of signals Smad is not expired because Smad7 activity is very low that is manifested by liver fibrosis [139]. *In vitro* it has been demonstrated that cyclosporin analogue NIM811 which was deprived of immunosuppressive activity is able to reduce collagen production by hepatic stellate cells and increase activity of MMP-1 (matrix-metalloproteinase-1) through inhibition of TGF β signalling (inhibits phosphorylation of Smad2 and Smad3, enhances phosphorylation of Smad7) [152]. Resistin-induced TGF $\beta 1$ from Kupffer cells enhanced HSC collagen I expression. Resistin directly and indirectly modulates HSC behaviour towards a more pro-fibrogenic phenotype [153]. It has been shown that blockade of thrombospondin 1 (TGF $\beta 1$ activator) protects rats from liver damage and fibrosis induced by dimethylnitrosamine [154] and the inhibition of

TGF β 1 by providing anti-TGF β 1, decorin (binds TGF β 1), antisense oligonucleotides reduces the accumulation of ECM in glomerulonephritis [68]. There are also interesting observations concerning the activity of mineralocorticoids and approved in the treatment of autoimmune liver diseases glucocorticosteroids. In hepatic stellate cells and in miofibroblasts, the presence of receptors for these above hormones has been detected. Up to eight hours after glucocorticosteroids administration, the TGF β RIII expression was increasing in a dose dependent manner and time of their administration, while TGF β RI and TGF β RII expression did not change. Glucocorticosteroids through modulation of mRNA TGF β RIII expression can influence the final effect of TGF β 1. Since TGF β 1 and glucocorticosteroids have immunosuppressive activity and after glucocorticosteroids administration tissue sensitivity to the TGF β 1 is increased, it can be assumed that they act synergistically. Thus, the beneficial effects of glucocorticosteroids in autoimmune hepatitis may be a result of such synergistic mechanism of intensifying immunosuppression. Due to the profibrogenic TGF β 1 action, in this situation an immunosuppressive effect seems to be clinically more important and the above explanation is not justified in the case of chronic hepatitis other than an autoimmune aetiology. It has been found that the degree of induction of TGF β RIII expression is dependent on the nature of stimulating hormone — dexamethasone, hydrocortisone, aldosterone and their doses. Glucocorticosteroids through modulation of expression of mRNA TGF β RIII affect the final effect of TGF β 1. On the other hand, the increase of mRNA TGF β RIII expression in hepatic stellate cells by aldosterone is appeared through mineralocorticosteroids receptors present in the cells [37], and mineralocorticosteroids antagonist are candidates for drugs acting against liver fibrosis [155]. Glucocorticosteroids and mineralocorticosteroids and their receptors interact with signal transduction of TGF β 1 at the transcription and translation level [37]. We still do not know how to select the candidates for the anti-TGF β 1 strategy or anti-TGF β 1 treatment. At this stage, it is difficult to predict the side effects of preparations whose primary purpose is interfering with the activity of TGF β 1 and its receptors. There are, however, commonly applied drugs, and one of the additional (or recently discovered) measures is the antifibrogenic effect.

In various clinical conditions, attempts are being made to apply TGF β 1. There have been thus obtained acceleration of healing of skin ulcers, scar formation in post-operative wounds, scarring skin lesions in psoriasis, and healing in the process of merging the bone [6]. In turn, the demonstrated ability to inhibit tumour growth via TGF β RII, and the discovery of factors inducing the expression of this receptor in tumour cells, may be relevant to the treatment of malignant disease.

References

- Moses HL, Branum EL, Proper JA et al. Transforming growth factor production by chemically transformed cells. *Cancer Res* 1981; 41: 2842–2848.
- Zhou S, Kinzler KW, Vogelstein B. Going mad with Smads. *N Engl J Med* 1999; 341: 1144–1146.
- Krzemień S, Knapczyk P. Aktualne poglądy dotyczące znaczenia transformującego czynnika wzrostu beta (TGF- β) w patogenezie niektórych stanów chorobowych. *Wiad Lek* 2005; 58: 536–539.
- Ota K, Quint P, Weivoda MM et al. Transforming growth factor beta 1 induces CXCL16 and leukemia inhibitory factor expression in osteoclasts to modulate migration of osteoblast progenitors. *Bone* 2013; 57: 68–75.
- Babyatsky MW, Rossiter G, Podolsky DK. Expression of transforming growth factors alpha and beta in colonic mucosa in inflammatory bowel disease. *Gastroenterology* 1996; 110: 975–984.
- Flisiak R, Wiercińska-Drapała A, Tynecka E. Transformujący czynnik wzrostu b w patogenezie chorób wątroby. *Wiad Lek* 2000; 53: 530–537.
- Marek A, Brodzicki J, Liberek A et al. TGF- β (transforming growth factor- β) in chronic inflammatory conditions — a new diagnostic and prognostic marker? *Med Sci Monit* 2002; 8: 145–151.
- Kato Y, Inoue H, Yoshioka U et al. Effects of transforming growth factor β 1, interleukin-1b, tumor necrosis factor α and platelet-derived growth factor on the collagen synthesis and the proliferation of periacinar fibroblastoid cells isolated and cultured from rat pancreatic acini. *Pathophysiology* 1999; 3: 175–179.
- Gressner AM. Cytokines and cellular cross-talk involved in the activation of fat-storing cells. *J Hepatol* 1995; 22 (Suppl. 2): 28–36.
- De Bleser PJ, Niki T, Rogiers V et al. Transforming growth factor- β gene expression in normal and fibrotic rat liver. *J Hepatol* 1997; 26: 886–893.
- Blobe GC, Schiemann WP, Lodish HF. Role of transforming growth factor beta in human disease. *N Engl J Med* 2000; 342: 1350–1358.
- Gacka M, Adamiec R. Rola TGF- β w patogenezie miażdżycowego uszkodzenia naczyń. *Pol Arch Med Wewn* 2002; 108: 987–991.
- Yoshiji H, Noguchi R, Ikenaka Y et al. Renin-angiotensin system inhibitors as therapeutic alternatives in the treatment of chronic liver diseases. *Curr Med Chem* 2007; 14: 2749–2754.
- Moreno M, Ramalho LN, Sancho-Bru P et al. Atorvastatin attenuates angiotensin II-induced inflammatory actions in the liver. *Am J Physiol Gastrointest Liver Physiol* 2008; 0: 00462.2007v1.
- Turkay C, Yonem O, Arici S et al. Effect of angiotensin-converting enzyme inhibition on experimental hepatic fibrogenesis. *Dig Dis Sci* 2008; 53: 789–793.
- Yayama K, Miyagi R, Sugiyama K et al. Angiotensin II regulates liver regeneration via type I receptor following partial hepatectomy in mice. *Biol Pharm Bull* 2008; 31: 1356–1361.
- Massague J, Andres J, Attisano L et al. TGF- β receptors. *Mol Reprod Dev* 1992; 32: 99–104.
- Heldin CH, Miyazono K, Ten Dijke P. TGF- β signaling from cell membrane to nucleus through SMAD proteins. *Nature* 1997; 390: 465–471.
- Wrana JL, Attisano L, Wieser R et al. Mechanism of activation of the TGF-beta receptor. *Nature* 1994; 370: 341–347.
- Flisiak R, Pytel-Krolczuk P, Prokopowicz D. Circulating transforming growth factor β 1 as an indicator of hepatic function impairment of liver cirrhosis. *Cytokine* 2000; 12: 677–681.
- Derynck R, Feng XH. TGF-beta receptor signaling. *Biochim Biophys Acta* 1997; 1333: F105–150.
- Massague J, Wotton D. Transcriptional control by the TGF-beta/Smad signaling system. *EMBO J* 2000; 19: 1745–1754.
- Attisano L, Wrana JL. Signal transduction by the TGF- β superfamily. *Science* 2002; 296: 1646–1647.
- Ten Dijke P, Goumans MJ, Itoh F et al. Regulation of cell proliferation by Smad proteins. *J Cell Physiol* 2002; 191: 1–16.
- Roulot D, Sevcsik AM, Coste T et al. Role of transforming growth factor β type II receptor in hepatic fibrosis: studies of human chronic hepatitis C and experimental fibrosis in rats. *Hepatology* 1999; 29: 1730–1738.
- Wells RG. TGF- β signaling pathways. *Am J Physiol* 2000; 279: G845–G850.
- Bissell DM, Roulot D, George J. Transforming growth factor β and liver. *Hepatology* 2001; 34: 859–867.
- Wells RG, Yankelev H, Lin HY et al. Biosynthesis of the type I and Type II TGF- β receptors. Implications for complex formation. *J Biol Chem* 1997; 272: 11444–11451.
- Wrana JL. Transforming growth factor- β signaling and cirrhosis. *Hepatology* 1999; 29: 1909–1910.
- Lopez-Casillas F, Wrana JL, Massague J. Betaglycan presents ligand to the TGF beta signaling receptor. *Cell* 1993; 73: 1435–1444.
- Rodriguez C, Chen F, Weinber RA et al. Cooperative binding of transforming growth factor (TGF)-beta 2 to the types I and II TGF-beta receptors. *J Biol Chem* 1995; 270: 15919–15922.
- Sankar RM, Mahooti-Brooks N, Centrella M et al. Expression of transforming growth factor type III receptor in vascular endothelial cell increases their responsiveness to transforming growth factor β 2. *J Biol Chem* 1995; 270: 13567–13572.

33. Centrella M, Ji C, McCarthy TL. Control of TGF-beta receptor expression in bone. *Front Biosci* 1998; 3: d113-d124.
34. Esparza-Lopez J, Montiel JL, Vilchis-Landeros MM et al. Ligand binding and functional properties of betaglycan, a co-receptor of the transforming growth factor-beta superfamily. Specialized binding regions for transforming growth factor-beta and inhibitor A. *J Biol Chem* 2001; 276: 14588-14596.
35. Wickert L, Steinkruger S, Abiaka M et al. Quantitative monitoring of the mRNA expression pattern of the TGF-beta-isoforms (beta 1, beta 2, beta 3) during transdifferentiation of hepatic stellate cells using a newly developed real-time SYBR Green PCR. *Biochem Biophys Res Commun* 2002; 295: 330-335.
36. Eickelberg O, Centrella M, Reiss M et al. Betaglycan inhibits TGF-beta signaling by preventing type I-type II receptor complex formation. Glycosaminoglycan modifications alter betaglycan function. *J Biol Chem* 2002; 277: 823-829.
37. Wickert L, Abiaka M, Bolkenius U et al. Corticosteroids stimulate selectively transforming growth factor (TGF)- β receptor type III expression in transdifferentiating hepatic stellate cells. *J Hepatol* 2004; 40: 69-76.
38. Nakao A, Okumura K, Ogawa H. Smad7: a new key player in TGF-beta associated disease. *Trends Mol Med* 2002; 8: 361-363.
39. Fink SP, Mikkola D, Willson JKV et al. TGF- β -induced nuclear localization of Smad2 and Smad3 in Smad4 null cancer cell line. *Oncogene* 2003; 22: 1317-1323.
40. Xie W, Rimm DL, Lin Y et al. Loss of Smad signaling in human colorectal cancer is associated with advanced disease and poor prognosis. *Cancer J* 2003; 9: 302-312.
41. Matsuzaki K, Murata M, Yoshida K et al. Chronic inflammation associated with hepatitis C virus infection perturbs hepatic transforming growth factor beta signaling, promoting cirrhosis and hepatocellular carcinoma. *Hepatology* 2007; 46: 48-57.
42. Gressner AM, Weiskirchen R, Breitkopf K et al. Roles of TGF- β in hepatic fibrosis. *Front Biosci* 2002; 7: d793-d807.
43. Dooley S, Hamzavi J, Breitkopf K et al. Smad7 prevents activation of hepatic stellate cells and liver fibrosis in rats. *Gastroenterology* 2003; 125: 178-191.
44. Monteleone G, Kumberova A, Croft NM et al. Blocking Smad7 restores TGF- $\beta 1$ signaling in chronic inflammatory bowel disease. *J Clin Invest* 2001; 108: 601-609.
45. Lan HY, Chung AC. TGF- β /Smad signaling in kidney disease. *Semin Nephrol* 2012; 32: 236-243.
46. Genc H, Karadurmus N, Kisa U et al. Transforming growth factor β (TGF- β) levels in otherwise healthy subjects with impaired glucose tolerance. *Endokrynol Pol* 2010; 61: 691-694.
47. Kajdaniuk D, Marek B, Borgiel-Marek H et al. Vascular endothelial growth factor (VEGF) — part 1: in physiology and pathophysiology. *Endokrynol Pol* 2011; 62: 444-455.
48. Kajdaniuk D, Marek B, Foltyn W et al. Vascular endothelial growth factor (VEGF) — part 2: in endocrinology and oncology. *Endokrynol Pol* 2011; 62: 456-464.
49. Aaronson SA. Growth factors and cancer. *Science* 1991; 254: 1146-1153.
50. Bursch W, Oberhammer F, Schulte-Hermann R. Cell death and its protective role in disease. *Trends Pharmacol Sci* 1992; 13: 245-251.
51. Moses HL, Yang EY, Pietenpol JA. TGF-beta stimulation and inhibition of cell proliferation: new mechanistic insights. *Cell* 1990; 63: 245-247.
52. Hirschhorn T, Barzilay L, Smorodinsky NI et al. Differential regulation of Smad3 and of the type II transforming growth factor- β receptor in mitosis: implications for signaling. *PLoS One* 2012; 7: e43459.
53. Ibelgaufts H. Cytokines and Cells Online Pathfinder Encyclopaedia (COPE), 2007. <http://www.copewithcytokines.de/cope.cgi>
54. Dignass AU, Podolsky DK. Cytokine modulation of intestinal epithelial cell restitution: central role of transforming growth factor beta. *Gastroenterology* 1993; 105: 1323-1332.
55. Knabbe C, Zugmaier G. Expression of transforming growth factor- β in breast cancer. *Endocr Relat Cancer* 1994; 1: 5-17.
56. Nakamura T, Tomita Y, Hirai R et al. Inhibitory effect of transforming growth factor- β on DNA synthesis of adult rat hepatocytes in primary culture. *Biochem Biophys Res Commun* 1985; 133: 1042-1050.
57. Beck PL, Rosenberg IM, Xavier RJ et al. Transforming growth factor-beta mediates intestinal healing and susceptibility to injury in vitro and in vivo through epithelial cells. *Am J Pathol* 2003; 162: 597-608.
58. Choy L, Derynck R. The type II transforming growth factor (TGF)-beta receptor-interacting protein TRIP-1 acts as modulator of the TGF-beta response. *J Biol Chem* 1998; 273: 31455-31462.
59. Sambueli A, Diez RA, Sugai E et al. Serum transforming growth factor-beta 1 levels increase in response to successful anti-inflammatory therapy in ulcerative colitis. *Aliment Pharmacol Ther* 2000; 14: 1443-1449.
60. Grasl-Kraupp B, Rossmannith W, Ruttkay-Nedecky B et al. Levels of transforming growth factor beta and transforming growth factor beta receptors in rat liver during growth, regression by apoptosis and neoplasia. *Hepatology* 1998; 28: 717-726.
61. Masuhara M, Yasunaga M, Tanigawa K et al. Expression of hepatocyte growth factor, transforming growth factor α , and transforming growth factor $\beta 1$ messenger RNA in various human liver diseases and correlation with hepatocyte proliferation. *Hepatology* 1996; 24: 323-329.
62. Kosone T, Takagi H, Horiguchi N et al. Transforming growth factor-alpha accelerates hepatocyte repopulation after hepatocyte transplantation. *J Gastroenterol Hepatol* 2008; 23: 260-266.
63. Ichikawa T, Zhang Y-Q, Kogure K et al. Transforming growth factor β and activin tonically inhibit DNA synthesis in the rat liver. *Hepatology* 2001; 34: 918-925.
64. Di Mola FF, Fries H, Scheuren A et al. Transforming growth factors-betas and their signaling receptors are coexpressed in Crohn's disease. *Ann Surg* 1999; 229: 67-75.
65. Border WA, Noble NA. Fibrosis linked to TGF-beta in yet another disease. *J Clin Invest* 1995; 96: 655-656.
66. Nowak M, Marek B, Glogowska-Szeląg J et al. Powikłania oczne w reumatoidalnym zapaleniu stawów. *Reumatologia* 2005; 43: 216-221.
67. Tsunawaki S, Sporn M, Ding A, Nathan C. Deactivation of macrophages by transforming growth factor-beta. *Nature* 1988; 334: 260-262.
68. Boratyńska M. Urine excretion of transforming growth factor- $\beta 1$ in chronic allograft nephropathy. *Ann Transplant* 1999; 4: 23-28.
69. Kehrl JH, Roberts AB, Wakefield LM et al. Transforming growth factor beta is an important immunomodulatory protein for human B lymphocytes. *J Immunol* 1986; 137: 3855-3860.
70. Jakóbsiak M. Powstawanie przeciwciał. In: Gołąb J, Jakóbsiak M, Lasek W. (eds.). *Immunologia*. Wydaw. PWN, Warszawa 2002: 34-45.
71. Woszczyk D, Gola J, Jurzak M et al. Expression of TGF $\beta 1$ genes and their receptor types I, II, and III in low- and high-grade malignancy non-Hodgkin's lymphomas. *Med Sci Monit* 2004; 10: CR33-CR37.
72. Myśliwiec J, Palyga I, Nikolajuk A et al. Serum interleukin-16 and RANTES during treatment of Graves' orbitopathy with corticosteroids and teleradiotherapy. *Endokrynol Pol* 2012; 63: 92-96.
73. Cottrez F, Groux H. Regulation of TGF- β response during T cell activation is modulated by IL-10. *J Immunol* 2001; 167: 773-778.
74. Cerwenka A, Kowar H, Majdic O et al. Fas- and activation-induced apoptosis are reduced in human T cells preactivated in the presence of TGF-beta 1. *J Immunol* 1996; 156: 459-464.
75. Jakóbsiak M, Gołąb J. Odporność nieswoista. In: I Gołąb J, Jakóbsiak M, Lasek W. (eds.). *Immunologia*. Wydaw. PWN, Warszawa 2002: 157-175.
76. Christ M, McCartney-Francis NL, Kulkarni AB et al. Immune dysregulation in TGF- $\beta 1$ -deficient mice. *J Immunol* 1994; 153: 1936-1946.
77. Kanamaru Y, Nakao A, Mamura M et al. Blockade of TGF β signaling in T cells prevents the development of experimental glomerulonephritis. *J Immunol* 2001; 166: 2818-2823.
78. Fine A, Goldstein RH. The effect of transforming growth factor- β on cell proliferation and collagen formation by lung fibroblasts. *J Biol Chem* 1987; 262: 3897-3902.
79. Ignatz RA, Massague J. Transforming growth factor β stimulates the expression of fibronectin and collagen and their incorporation into extracellular matrix. *J Biol Chem* 1986; 261: 4337-4345.
80. Kuwahara F, Kai H, Tokuda K et al. Transforming growth factor- β function blocking prevents myocardial fibrosis and diastolic dysfunction in pressure-overloaded rats. *Circulation* 2002; 106: 130-135.
81. Duncan MR, Frazier KS, Abramson S et al. Connective tissue growth factor mediates transforming growth factor beta-induced collagen synthesis: down-regulation by cAMP. *FASEB J* 1999; 13: 1774-1786.
82. Paradis V, Dargere D, Bonvoust F et al. Effects and regulation of connective tissue growth factor on hepatic stellate cells. *Lab Invest* 2002; 82: 767-774.
83. Weng HL, Ciucan L, Liu Y et al. Profibrogenic transforming growth factor-beta/activin receptor-like kinase 5 signaling via connective tissue growth factor expression in hepatocytes. *Hepatology* 2007; 46: 1257-1270.
84. Wiercińska-Drapała A, Flisiak R, Prokopowicz D. Effect of ulcerative colitis activity on plasma concentration of transforming growth factor $\beta 1$. *Cytokine* 2001; 14: 343-346.
85. McKaig BC, McWilliams D, Watson SA et al. Expression and regulation of tissue inhibitor of metalloproteinase-1 and matrix metalloproteinases by intestinal myofibroblasts in inflammatory bowel disease. *Am J Pathol* 2003; 162: 1355-1360.
86. Lionetti P, Pazzaglia A, Moriondo M et al. Differing patterns of transforming growth factor β expression in normal intestinal mucosa and active celiac disease. *J Pediatr Gastroenterol Nutr* 1999; 29: 308-313.
87. Chung HL, Hwang JB, Park JJ et al. Expression of transforming growth factor $\beta 1$, transforming growth factor type I and II receptors, and TNF- α in the mucosa of the small intestine in infants with food protein-induced enterocolitis syndrome. *J Allergy Clin Immunol* 2002; 109: 150-154.
88. Stromberg E, Edebo A, Svennerholm AM et al. Decreased epithelial cytokine responses in the duodenal mucosa of the *Helicobacter pylori*-infected duodenal ulcer patients. *Clin Diagn Lab Immunol* 2003; 10: 116-124.
89. Perez-Aisa A, Sopena F, Arceiz E et al. Effect of exogenous administration of transforming growth factor-beta and famotidine on the healing of duodenal ulcer under the impact indomethacin. *Dig Liver Dis* 2003; 35: 397-403.
90. Zorena K, Malinowska E, Raczynska D et al. Serum concentrations of transforming growth factor-Beta 1 in predicting the occurrence of diabetic retinopathy in juvenile patients with type 1 diabetes mellitus. *J Diabetes Res* 2013; 2013: Article ID 614908.

91. Arkwright PD, Chase JM, Babbage S et al. Atopic dermatitis is associated with low-producer transforming growth factor β 1 cytokine genotype. *J Allergy Clin Immunol* 2001; 108: 281–284.
92. Jude EB, Blakytyn R, Bulmer J et al. Transforming growth factor-beta 1, 2, 3 and receptor type I and II in diabetic foot ulcers. *Diabet Med* 2002; 19: 440–447.
93. Kim BC, Kim HT, Park SH et al. Fibroblasts from chronic wounds show altered TGF-beta signaling and decreased TGF-beta type II receptor expression. *J Cell Physiol* 2003; 195: 331–336.
94. Chakir J, Shannon J, Molet S et al. Airway remodeling-associated mediators in moderate to severe asthma: effects of steroids on TGF-beta, IL-11, IL-17, and type I and type III collagen expression. *J Allergy Clin Immunol* 2003; 111: 1293–1298.
95. Duvernelle C, Freund V, Frossard N. Transforming growth factor-beta and its role in asthma. *Pulm Pharmacol Ther* 2003; 16: 181–196.
96. Okada S, Hasegawa S, Hasegawa H et al. Analysis of bronchoalveolar lavage fluid in a mouse model of bronchial asthma and H1N1 2009 infection. *Cytokine* 2013; 63: 194–200.
97. Huang L, Jia J, Liu R. Decreased serum levels of the angiogenic factors VEGF and TGF- β 1 in Alzheimer's disease and amnesic mild cognitive impairment. *Neurosci Lett* 2013; 550: 60–63.
98. Misiorowski M. Parathyroid hormone and its analogues — molecular mechanisms of action and efficacy in osteoporosis therapy. *Endokrynol Pol* 2011; 62: 73–78.
99. Sowińska-Przepiera E, Andrysiak-Mamos E, Jarząbek-Bielecka G et al. Effects of oestrogen deficiency on bone mineralization in girls during "adolescent crisis". *Endokrynol Pol* 2011; 62: 538–546.
100. Foltyn W, Strzelczyk J, Marek B et al. The usefulness of determining the serum concentrations of vascular endothelial growth factor (VEGF) and its soluble receptor type 2 (sVEGF-2) in the differential diagnosis of adrenal incidentalomas. *Endokrynol Pol* 2012; 63: 22–28.
101. Telega A, Kos-Kudła B, Foltyn W et al. Selected neuroendocrine tumour markers, growth factors and their receptors in typical and atypical bronchopulmonary carcinoids. *Endokrynol Pol* 2012; 63: 477–482.
102. Skóra J, Barć P, Pupka A et al. Transplantation of autologous bone marrow mononuclear cells with VEGF gene improves diabetic critical limb ischemia. *Endokrynol Pol* 2013; 64: 129–138.
103. Renner U, Lohrer P, Schaaf L et al. Transforming growth factor-beta stimulates vascular endothelial growth factor production by folliculostellate pituitary cells. *Endocrinology* 2002; 143: 3759–3765.
104. De A, Morgan TE, Speth RC et al. Pituitary lactotrope expresses transforming growth factor beta (TGFbeta) type II receptor mRNA and protein and contains 125I-TGFbeta1 binding sites. *J Endocrinol* 1996; 149: 19–27.
105. Elenkova A, Atanassova I, Kirilov G et al. Transforming growth factor β 1 is not a reliable biomarker for valvular fibrosis but could be a potential serum marker for invasiveness of prolactinomas (pilot study). *Eur J Endocrinol* 2013; 169: 299–306.
106. Corduk N, Abban G, Yildirim B et al. The effect of vitamin D on expression of TGF β 1 in ovary. *Exp Clin Endocrinol Diabetes* 2012; 120: 490–493.
107. Maurya VK, Jha RK, Kumar V et al. Transforming Growth Factor-Beta 1 (TGF- β) Liberation from Its Latent Complex During Embryo Implantation and Its Regulation by Estradiol in Mouse. *Biol Reprod* 2013; 89: 1–17.
108. Tal R, Seifer DB, Shohat-Tal A et al. Transforming growth factor- β 1 and its receptor soluble endoglin are altered in polycystic ovary syndrome during controlled ovarian stimulation. *Fertil Steril* 2013; 100: 538–543.
109. Hou YL, Chen H, Dong ZH et al. Clinical significance of serum transforming growth factor- β 1 in lung cancer. *Cancer Epidemiol* 2013; 37: 750–753.
110. Idilman R, De Maria N, Colationi A et al. Pathogenesis of hepatitis B and C-induced hepatocellular carcinoma. *J Viral Hepat* 1998; 5: 285–299.
111. Grady WM, Rajput A, Myeroff L et al. Mutation of the type II transforming growth factor-beta receptor is coincident with the transformation of human colon adenomas to malignant carcinomas. *Cancer Res* 1998; 58: 3101–3104.
112. Lazzereschi D, Ranieri A, Mincione G et al. Human malignant thyroid tumors displayed reduced levels of transforming growth factor beta receptor type II messenger RNA and protein. *Cancer Res* 1997; 57: 2071–2076.
113. Turco A, Coppa A, Aloe S et al. Overexpression of transforming growth factor beta-type II receptor reduces tumorigenicity and metastatic potential of K-ras-transformed thyroid cells. *Int J Cancer* 1999; 80: 85–91.
114. Matoba H, Sugano S, Yamaguchi N et al. Expression of transforming growth factor-beta1 and transforming growth factor-beta type-II receptor mRNA in papillary thyroid carcinoma. *Horm Metab Res* 1998; 30: 624–628.
115. Liu G, Takano T, Amino N. TGF-beta 1 inhibits the cell proliferation stimulated by IGF-I by blocking the tyrosine phosphorylation of 175 kDa substrate. *Endocr Res* 1996; 22: 277–287.
116. Zhang J, Wang Y, Li D et al. Notch and TGF- β /Smad3 pathways are involved in the interaction between cancer cells and cancer-associated fibroblasts in papillary thyroid carcinoma. *Tumour Biol* 2013.
117. Wikstrom P, Damber J, Bergh A. Role of transforming growth factor-beta1 in prostate cancer. *Microsc Res Tech* 2001; 52: 411–419.
118. Ramos-Flores C, Romero-Gutiérrez T, Delgado-Enciso I et al. Polymorphisms in the genes related to angiogenesis are associated with uterine cervical cancer. *Int J Gynecol Cancer* 2013; 23: 1198–1204.
119. Myeroff LL, Parsons R, Kim S-J et al. A transforming growth factor beta receptor type II gene mutation common in colon and gastric but rare in endometrial cancers with microsatellite instability. *Cancer Res* 1995; 55: 5545–5547.
120. Renault B, Calistri D, Buonsanti G et al. Microsatellite instability and mutations of p53 and TGF-beta RII genes in gastric cancer. *Hum Genet* 1996; 98: 601–607.
121. Thorgeirsson SS, Teramoto T, Factor VM. Dysregulation of apoptosis in hepatocellular carcinoma. *Semin Liver Dis* 1998; 18: 115–122.
122. Gressner AM, Lahme B, Mannherz HG et al. TGF β -mediated hepatocellular apoptosis by rat and human hepatoma cells and primary rat hepatocytes. *J Hepatol* 1997; 26: 1079–1092.
123. Furuta K, Misao S, Takahashi K et al. Gene mutation of transforming growth factor beta1 type II receptor in hepatocellular carcinoma. *Int J Cancer* 1999; 81: 851–853.
124. Dong ZZ, Yao DF, Yao M et al. Clinical impact of plasma TGF-beta1 and circulating TGF-beta1 mRNA in diagnosis of hepatocellular carcinoma. *Hepatobiliary Pancreat Dis Int* 2008; 7: 288–295.
125. Lu Y, Wu L-Q, Li C-S et al. Expression of transforming growth factors in hepatocellular carcinoma and its relations with clinicopathological parameters and prognosis. *Hepatobiliary Pancreat Dis Int* 2008; 7: 174–178.
126. Coulouarn C, Factor VM, Thorgeirsson SS. Transforming growth factor-beta gene expression signature in mouse hepatocytes predicts clinical outcome in human cancer. *Hepatology* 2008; 47: 2059–2067.
127. Ray S, Broor SL, Vaishnav Y et al. Transforming growth factor beta in hepatitis C virus infection: In vivo and in vitro findings. *J Gastroenterol Hepatol* 2003; 18: 393–403.
128. Kim HG, Chung YH, Song BC et al. Expression of TGF β -1 in chronic hepatitis and hepatocellular carcinoma associated with hepatitis C virus infection. *Korean J Intern Med* 2000; 15: 165–170.
129. Shirai Y, Kawata S, Tamura S et al. Plasma transforming growth factor- β 1 in patients with hepatocellular carcinoma. Comparison with chronic liver diseases. *Cancer* 1994; 73: 2275–2279.
130. Sobue S, Nomura T, Ishikawa T et al. Th1/Th2 cytokine profiles and their relationship to clinical features in patients with chronic hepatitis C virus infection. *J Gastroenterol* 2001; 36: 544–551.
131. Tsai JF, Jeng JE, Chuang LY et al. Urinary transforming growth factor- β 1 levels in hepatitis C virus-related chronic liver disease: correlation between high levels and severity of disease. *Hepatology* 1997; 25: 1141–1146.
132. Jakowlew SB, Moody TW, You L et al. Reduction in transforming growth factor-beta type II receptor in mouse lung carcinogenesis. *Mol Carcinog* 1998; 22: 46–56.
133. Yuan X, Wei Q, Komaki R et al. TGF β 1 polymorphisms predict distant metastasis-free survival in patients with inoperable Non-Small-Cell Lung Cancer after definitive radiotherapy. *PLoS One* 2013; 8: e65659.
134. Schedlich LJ, Yenson VM, Baxter RC. TGF- β -induced expression of IGFBP-3 regulates IGF1R signaling in human osteosarcoma cells. *Mol Cell Endocrinol* 2013; 377: 56–64.
135. Imai K, Minamiya Y, Goto A et al. Bronchioloalveolar invasion in non-small cell lung cancer is associated with expression of transforming growth factor- β 1. *World J Surg Oncol* 2013; 11: 113.
136. Krasagakis K, Tholke D, Farthmann B et al. Elevated plasma levels of transforming growth factor (TGF)-beta1 and TGF-beta2 in patients with disseminated malignant melanoma. *Br J Cancer* 1998; 77: 1492–1494.
137. Awad MR, EL-Gamel A, Hasleton P et al. Genotypic variation in the transforming growth factor- β 1 gene. Association with transforming growth factor- β 1 production, fibrotic lung disease, and graft fibrosis after lung transplantation. *Transplantation* 1998; 66: 1014–1020.
138. Samarakoon R, Dobberfuhl AD, Cooley C et al. Induction of renal fibrotic genes by TGF- β 1 requires EGFR activation, p53 and reactive oxygen species. *Cell Signal* 2013; 25: 2198–2209.
139. Tahashi Y, Matsuzaki K, Date M et al. Differential regulation of TGF-beta signal in hepatic stellate cells between acute and chronic rat liver injury. *Hepatology* 2002; 35: 49–61.
140. Shah R, Reyes-Gordillo K, Arellanes-Robledo J et al. TGF- β 1 up-regulates the expression of PDGF- β receptor mRNA and induces a delayed PI3K-, AKT-, and p70S6K-dependent proliferative response in activated hepatic stellate cells. *Alcohol Clin Exp Res* 2013; DOI: 10.1111/acer.12167.
141. Date M, Matsuzaki K, Matsushita M et al. Modulation of transforming growth factor β function in hepatocytes and hepatic stellate cells in rat liver injury. *Gut* 2000; 46: 719–724.
142. Benzoubir N, Lejamtel C, Battaglia S et al. HCV core-mediated activation of latent TGF- β via thrombospondin drives the cross-talk between hepatocytes and stromal environment. *J Hepatol* 2013; DOI: 10.1016/j.hep.2013.07.036.

143. Martin-Vílchez S, Sanz-Cameno P, Rodríguez-Muñoz Y et al. The hepatitis B virus X protein induces paracrine activation of human hepatic stellate cells. *Hepatology* 2008; 47: 1872–1883.
144. Ueki T, Kaneda Y, Tsutsui H et al. Hepatocyte growth factor gene therapy of liver cirrhosis in rats. *Nat Med* 1999; 5: 226–230.
145. Qi Z, Atsuchi N, Ooshima A et al. Blockade of type beta transforming growth factor signaling prevents liver fibrosis and dysfunction in the rat. *Proc Natl Acad Sci USA* 1999; 96: 2345–2349.
146. Ueno H, Sakamoto T, Nakamura T et al. A soluble transforming growth factor beta receptor expressed in muscle prevents liver fibrogenesis and dysfunction in rats. *Hum Gene Ther* 2000; 11: 33–42.
147. George J, Roulot D, Kotliansky VE et al. In vivo inhibition of rat stellate cell activation by soluble transforming growth factor β type II receptor: A potential new therapy for hepatic fibrosis. *Proc Natl Acad Sci USA* 1999; 96: 12719–12724.
148. Isaka Y, Akagi Y, Ando Y et al. Gene therapy by transforming growth factor beta-receptor-IgG Fc chimera suppressed extracellular matrix accumulation in experimental glomerulonephritis. *Kidney Int* 1999; 55: 465–475.
149. Ling H, Roux E, Hempel D et al. Transforming growth factor β neutralization ameliorates pre-existing hepatic fibrosis and reduces cholangiocarcinoma in thioacetamide-treated rats. *PLoS One* 2013; 8: e54499.
150. Nakamura T, Sakata R, Ueno T et al. Inhibition of transforming growth factor β prevents progression of liver fibrosis and enhances hepatocyte regeneration in dimethylnitrosamine-treated rats. *Hepatology* 2000; 32: 247–255.
151. Yata Y, Gotwals P, Kotliansky V et al. Dose-dependent inhibition of hepatic fibrosis in mice by a TGF β soluble receptor: implications for antifibrotic therapy. *Hepatology* 2002; 35: 1022–1030.
152. Kohjima M, Enjoji M, Higuchi N et al. NIM811, a nonimmunosuppressive cyclosporine analogue, suppresses collagen production and enhances collagenase activity in hepatic stellate cells. *Liver Int* 2007; 27: 1273–1281.
153. Dong ZX, Su L, Brymora J, Bird C et al. Resistin mediates the hepatic stellate cell phenotype. *World J Gastroenterol* 2013; 19: 4475–4485.
154. Kondou H, Mushiake S, Etani Y et al. A blocking peptide for transforming growth factor-beta1 activation prevents hepatic fibrosis in vivo. *J Hepatol* 2003; 39: 742–748.
155. Caligiuri A, De Franco RM, Romanelli RG et al. Antifibrogenic effects of canrenone, an anti-aldosteronic drug, on human hepatic stellate cells. *Gastroenterology* 2003; 124: 504–520.