



Placental growth hormone, pituitary growth hormone, insulin-like growth factor, and ghrelin in umbilical cord blood serum and amniotic fluid

Hormon wzrostu łożyskowy, przysadkowy hormon wzrostu, insulinopodobny czynnik wzrostu i grelina w surowicy krwi pępowinowej oraz w płynie owodniowym

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Abstract

Introduction: In the search for biomarkers that allow the prediction of neonatal growth and development, placental growth hormone (PGH), pituitary growth hormone (GH1), insulin-like growth factor 1 (IGF-1), and ghrelin concentrations were assessed in the amniotic fluid and in the umbilical cord blood of 92 neonates.

Material and methods: The proteins were assayed by the ELISA method. Their concentration values were compared in 57 full-term neonates and 35 prematurely born neonates, as well as in both large (> 4,000 g) and small neonates (< 2,500 g). Also, body mass and placenta mass were compared.

Results: Statistically significant differences both between prematurely born neonates and full-term neonates and between large and small neonates were obtained only in terms of the body mass of neonates and placenta mass. The concentration values of the hormones studied did not show statistically significant differences. A distinct tendency was noticed towards an increase in PGH concentration in both prematurely born and small neonates. In large neonates, statistically significantly higher IGF-1 concentrations were found compared to the prematurely born neonates.

Conclusions: Our studies indicate an important role for PGH in maintaining a proper IGF-1 pool and demonstrate the existence of a direct influence on the function of the placenta in prematurely born neonates through the activation of compensation mechanisms, which stimulate IGF-1 synthesis. (*Endokrynol Pol* 2013; 64 (4): 293–298)

Key words: PGH, GH1, IGF-1, ghrelin, neonate, umbilical cord blood serum, amniotic fluid

Streszczenie

Wstęp: Poszukując czynników, które pozwoliłyby na prognozowanie dalszego rozwoju noworodka, oznaczano we krwi pępowinowej oraz w płynie owodniowym 92. noworodków stężenie łożyskowego hormonu wzrostu (PGH), przysadkowego hormonu wzrostu (GH1), insulinopodobnego czynnika wzrostu 1 (IGF-1) oraz greliny.

Materiał i metody: Omawiane białka były oznaczane metodą ELISA. Porównano wartości ich stężenia u 57. noworodków i 35. wcześniaków oraz u noworodków dużych (> 4000 g) i małych (< 2500 g). Porównano również masę ciała noworodków i masę łożyska.

Wyniki: Statystycznie znamienne różnice pomiędzy wcześniakami i noworodkami donoszonymi oraz między dużymi i małymi dziećmi uzyskano jedynie w odniesieniu do masy ciała noworodków i masy łożyska. Natomiast wartości stężeń badanych hormonów nie wykazywały statystycznie znamienych różnic. Zaobserwowano wyraźną tendencję wzrastania stężenia PGH u wcześniaków i noworodków małych. Natomiast u noworodków dużych uzyskano statystycznie znamienne wyższe stężenie IGF-1 w porównaniu z wcześniakami.

Wnioski: Badania nasze wskazują na ważną rolę PGH w utrzymaniu odpowiedniej puli IGF-1 oraz bezpośredni wpływ na funkcję łożyska u wcześniaków poprzez uruchomienie mechanizmów kompensacyjnych stymulujących syntezę IGF-1. (*Endokrynol Pol* 2013; 64 (4): 293–298)

Słowa kluczowe: PGH, GH1, IGF-1, grelina, noworodek, surowica krwi pępowinowej, płyn owodniowy

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Introduction

The proteins listed in the title belong to the group of growth and cell metabolism modulators. They are regu-

lated by complexes of factors that act in systemic and local systems that differ in the pre- and postnatal periods. Growth processes are relatively well recognised in the postnatal phase, where the key endocrine factor



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controlling growth is the growth hormone 1/insulin-like growth factor 1 (GH1/IGF-1) axis [1]. Considerably less is known about the mechanisms controlling growth in foetal life in which IGF-1 does not work in conjunction with GH1 [2, 3].

The placental growth hormone is a product of the GH-V gene; it is a member of a family of five growth hormone genes located on the long arm of chromosome 17. This hormone has been known since 1985, when it was discovered by Hennen et al. [4, 5]. It shows a high level of homology to the pituitary growth hormone, being different in terms of 13 amino acids and N-glycosylation sites. This protein is synthesised and secreted by a syncytiotrophoblast and extravillous cytotrophoblast [2, 6, 7]. In their detailed review, Fuglsang and Ovesen [2] show that PGH seems to be only a maternal phenomenon. There are some publications describing the presence of this protein both in umbilical cord blood serum and amniotic fluid [8–10].

The aim of this study is an assessment of the concentrations of the studied proteins in umbilical cord blood serum and amniotic fluid. In addition, the authors have tried to determine if there is any relationship between the results obtained with neonate body mass and placenta mass, and the search for factors which would allow a child's development to be predicted.

Material and methods

Investigations were performed on umbilical cord blood and amniotic fluid samples. A total of 92 umbilical cord blood samples originating from one triplet pregnancy, 12 twin pregnancies and 79 singleton pregnancies were collected. The majority of the pregnancies were normal, eight pregnant women had type I diabetes (six type G1, one G2, and one type C). Among the 92 neonates, 52 boys and 40 girls were born. There were 35 infants born prematurely between weeks 22 and 37 of pregnancy, and 57 full-term neonates, born between weeks 37 and 42.

Among the neonates, 33 weighed more than 4,000 g and 38 less than 2,500 g.

All the neonates were examined by a neonatologist and weighed after birth; placenta mass was also measured. In four of the twin pregnancies, the placentas were common.

In neonate blood and amniotic fluid samples, the following hormones were assayed: pituitary growth hormone (GH1), placental growth hormone (PGH), insulin-like growth factor 1 (IGF-1) and acylated ghrelin.

Most of the amniotic fluids originated from amniocentesis: four fluids from weeks 14 and 15 of pregnancy, four fluids from week 17, two fluids from weeks 19 and 22, and two fluids collected during birth. A total of

12 amniotic fluids were collected from healthy pregnant women. All the investigated proteins were assayed using the ELISA method. GH1 and IGF-1 were assayed with R&D Systems ELISA Kits (R&D Systems, Minneapolis, MN, USA), PGH protein with a Uscn life Science ELISA Kit (Uscn life Science, Wuhan, China) and ghrelin was assayed with a BioVendor ELISA Kit (BioVendor, Modrice, Czech Republic), all according to the manufacturers' protocols. Briefly, the samples and standards were added as duplicates to micro-titration plates coated with antibodies against the studied substance. After incubation and washing, a secondary antibody was added, and the sample was incubated again. After subsequent washing, a colour reaction substrate, catalysed by a conjugated enzyme, was added. The reaction was stopped with 2M sulphuric acid. The plates were read in an MRX reader (Dy nex, Chantilly, VA, USA). For statistical analysis, SigmaStat 3.5 software was used. The normality of the distribution of the variables was analysed with the Shapiro-Wilk method. For a comparison of the non-parametrical distributed variables, the Mann-Whitney Rank Sum Test was used; $p < 0.05$ was considered statistically significant.

Results

As per the results presented in Table I, the median value for all the investigated factors was higher in full-term neonates than in prematurely born infants, except for PGH concentration, whose value was higher in prematurely born infants. Statistically significant differences between the two groups were shown only for neonate body mass and placenta mass. Concentrations of the investigated hormones did not show statistically significant differences. The highest mean values in full-term and prematurely born neonates were observed in IGF-1 concentrations (87.5 ng/mL, 57.8 ng/mL) and in pituitary growth hormone concentrations (3,176.2 pg/mL, 2,886.5 pg/mL).

During the analysis of particular cases, special attention was paid to the distinct tendency of PGH concentration to increase in small neonates compared to larger ones.

A similar situation was observed in the case of ghrelin concentrations. In some premature infants, ghrelin concentrations were higher than 100 pg/mL, while in full-term neonates they were around 50 pg/mL. However, in both cases, no statistically significant differences were observed. Besides, in five neonates (three prematurely born, two full-term infants), exceptionally low IGF-1 values, lower than 10 ng/mL, were observed, and this is within the error limits.

Additionally, large ($> 4,000$ g) and small ($< 2,500$ g) neonates were compared. The results are presented in Table II.

As per the results from Table II, statistically significant differences were shown for neonate body mass

Table I. Minimal values, maximal values, and median of the investigated hormones concentrations in umbilical cord blood of full-term and premature born neonates; neonate and placenta mass**Tabela I.** Wartości minimalne, maksymalne, mediana stężeń badanych hormonów we krwi pępowinowej noworodków donoszonych i wcześniaków, masa ciała noworodków i łożyska

Investigated material	Full-term neonate		Prematurely born neonate		p value
	Min-max concentration	Median	Min-max concentration	Median	
GH1 [pg/mL]	937.8–8,131.6	3,176.2	882.2–13,358.3	2,886.5	0.946
PGH [pg/mL]	0–2,015.6	346.9	106.4–1,728.7	433.9	0.478
IGF I [ng/mL]	10–132.2	87.5	10–110.6	57.8	0.184
Ghrelin [pg/mL]	14.2–175.5	70.5	14.3–184.5	53.5	0.617
Neonate [g]	1,790–5,430	4,230	600–2,650	1,770	< 0.001
Placenta [g]	270–1,300	700	150–900	420	< 0.001

Table II. Minimal values, maximal values, and median of the investigated hormones concentrations in umbilical cord blood of large (> 4,000 g) and small (< 2,500 g) neonates**Tabela II.** Wartości minimalne, maksymalne, mediana badanych hormonów we krwi pępowinowej noworodków dużych, powyżej 4000 g i małych poniżej 2500 g

Investigated material	Neonate > 4,000 g		Neonate < 2,500 g		p value
	Min-max concentration	Median	Min-max concentration	Median	
GH1 [pg/mL]	937.8–5,999.3	3,117.3	882.2–13,358.3	2,891.9	0.847
PGH [pg/mL]	0–2,015.6	344.3	106.4–1,768.6	396.6	0.386
IGF I [ng/mL]	45.4–132.2	98.9	10–110.6	50.0	< 0.001
ghrelin [pg/mL]	14.3–165.4	75.4	14.3–184.5	60.2	0.505
Neonate [g]	4,000–5,430	4,400	600–2,500	1,840	< 0.001
Placenta [g]	550–950	720	150–1,300	420	< 0.001

and placenta mass. Higher IGF-1 concentrations in the blood serum of large neonates compared to small neonates was statistically significant. Moreover, as in Table I, higher PGH concentrations were recorded in small neonates than in large neonates.

The growth hormone concentration ratio was also evaluated: GH1/PGH, GH1/IGF-1, GH1/ghrelin, PGH/IGF-1, PGH/ghrelin in the blood serum of full-term and prematurely born infants (Table III). In this table, the median of GH1 concentration in umbilical cord blood serum of full-term neonates was nine times higher than PGH, and six times higher than in prematurely born infants. The ratio of GH1 and IGF-1 concentrations in both investigated groups of neonates was similar and the IGF-1 concentration was about 25 times higher than GH1. The median IGF-1 concentration was 252 times higher than PGH in full-term neonates and 133 times higher in prematurely born infants. As for concentration ratio, there were no statistically significant differences between full-term and prematurely born neonates.

Similar to the results from Table 2, a significant correlation in both investigated groups occurred between

neonate body mass and placenta mass, and between GH1 and PGH in cord blood serum. Only full-term neonate body mass correlated with IGF-1 (Table IV).

Pituitary growth hormone concentration was the highest in second trimester amniotic fluid. PGH concentration was 50% lower, and the lowest concentrations were those of IGF-1 and ghrelin, and these were within error limits (Table V). In the amniotic fluid collected during the delivery, both growth hormone concentrations were several times lower than those for the second trimester of pregnancy.

Discussion

Our research showed that statistically significant differences in full-term and prematurely born neonates occurred only for neonate body mass and placenta mass. Median hormone concentration values for GH1, IGF-1, and ghrelin did not show statistically significant differences, and these were higher in full-term neonates and in the group of large infants (> 4,000 g). This situation was different for PGH concentrations. The median

Table III. Evaluation of the ratios of the investigated hormones from umbilical cord blood of full-term and prematurely born infants with median value. Hormone concentrations are presented in pg/mL**Tabela III.** Ocena stosunku stężeń badanych hormonów z krwi pępowinowej noworodków donoszonych i wcześniaków z wykorzystaniem wartości mediany. Stężenia hormonów podano w [pg/mL]

Analysed hormones [pg/mL]	Full-term neonates		Prematurely born neonates		p value
	Median	Concentration ratio	Median	Concentration ratio	
GH1/PGH	3,176.2/346.9	9:1	2,886.5/433.9	7:1	0.940
GH1/IGF-1	3,176.2/87,500	1:28	2,886.5/57,800	1:20	0.146
GH1/ghrelin	3,176.2/70.5	45:1	2,886.5/53.5	54:1	0.805
PGH/IGF-1	346.9/87,500	1:252	433.9/57,800	1:133	0.209
PGH/ghrelin	346.9/70.5	5:1	433.9/53.5	8:1	0.542

Table IV. Correlations between neonate birth weight and placenta mass, and the investigated hormones in cord blood serum**Tabela IV.** Korelacje pomiędzy masą ciała noworodków i masą łożyska oraz badanymi hormonami we krwi pępowinowej

Prematurely born infants	r	p	n
Neonate birth weight and placenta mass	0.574	0.001	32
Placenta mass and GH1	0.447	0.0152	29
GH1 and PGH	0.388	0.0377	29
PGH and ghrelin	0.365	0.0517	29
PGH and IGF-1	-0.548	0.0325	15
Ghrelin and IGF-1	-0.57	0.0252	15
Full-term neonates	r	p	n
Neonate birth weight and placenta mass	0.428	0.00255	48
Neonate birth weight and IGF-1	0.485	0.0141	25
GH1 and PGH	0.341	0.0179	48

Table V. Hormone concentration values in amniotic fluid**Tabela V.** Wartości stężeń badanych hormonów w płynie owodniowym

Week of pregnancy	GH1 [pg/mL]	PGH [pg/mL]	IGF-1 [ng/mL]	Ghrelin [pg/mL]
14–15	8,000	4,610.3	28.2	8.3
17	8,000	4,814.7	27.4	8.4
19–22	8,000	1,699.9	26.2	11.8
Collected during delivery	1,446.2	600	26.6	6

value of this hormone concentration was higher in prematurely born and small neonates (< 2,500 g), but the differences were not statistically significant (Table I and II). In the comparison of large and small infants (Table II), statistically significant differences also occurred in IGF-1 concentration. The median value of this hormone concentration was higher in large neonates. The placental growth hormone is synthesised by syncytiotrophoblast and secreted into the intervillous

space. The presence of this hormone in the syncytiotrophoblast was confirmed with immunohistochemical methods [6]. Besides, with molecular methods GH-V gene expression was shown in the placenta, and no expression of the GH-N gene, responsible for GH1 production, was demonstrated [7, 11, 12]. Moreover, the presence of PGH receptors was detected in the placenta, and this suggests a direct influence of PGH on placenta function through auto- and paracrine mechanisms,

which control the secretory and proliferative activity of the placenta [13–15]. Higher PGH concentrations observed in prematurely born and small fetuses may indicate compensation mechanisms, in which elevated PGH secretion stimulates IGF-1 synthesis in placental tissue, which accelerates the proliferation and growth of placental tissue [6].

It seems that the foetal pituitary gland does not play an important role in those compensation processes. No GH-V gene expression was noted in the pituitary gland, and no GH-N gene transcript was detected in the placenta [2, 7, 11, 16, 17]. Furthermore, the median value of GH1 concentrations was similar in large and small neonates and also in full-term and prematurely born infants (Table I and II). By comparing concentrations of the particular proteins, we showed that IGF-1 concentration values in full-term neonates is 252 times higher compared to PGH, and in prematurely born infants only 133 times higher (Table III). The IGF-1 to GH1 ratio of hormone concentrations was about 25 times higher and did not show higher differences in full-term and prematurely born neonates. These results enable us to suggest that PGH is an important modulator that maintains proper IGF-1 concentrations, and its secretion is not pulse. Chellakooty et al. [18] performed several examinations during pregnancy and showed a positive correlation between PGH concentration and foetal growth, as assessed by ultrasonography (USG). Our studies based on multivariate analysis indicate a positive correlation between neonate body mass and placenta mass in both investigated groups and also a correlation between full-term neonate body mass and IGF-1 concentration. In prematurely born infants, the correlation between PGH and IGF-1 concentrations was negative (Table IV). The results presented here and our suggestions require further investigations on a larger study group.

In terms of ghrelin concentrations, we have not shown statistically significant differences between full-term and prematurely born neonates, though in individual cases ghrelin concentrations were considerably higher in small neonates. Some authors have observed significantly higher concentration values for ghrelin in small neonates, but most researchers have not found any statistically significant differences [19–22]. Our previous research on human fetuses demonstrated that stimulation of GH1 secretion by ghrelin is independent of the feedback control and these two hormones act as a metabolic balance signal [3, 23]. In adults, the hypothalamic pituitary axis regulates pulse secretion of GH1 from the pituitary gland, but in fetuses is not functionally mature before the third trimester of pregnancy, independent of earlier anatomical differentiation [3].

The very high concentration of PGH in amniotic fluid, about tenfold higher than in umbilical cord blood, is surprising. Perhaps PGH undergoes partial degradation in umbilical cord blood caused by the different time of delivery and there could be a problem with the collection of purified serum connected with this delivery. On the other hand, when comparing our results to the results of other researchers, the PGH concentrations in umbilical cord blood and amniotic fluid obtained by us are over three times higher. Sifakis et al. [10] observed high PGH concentrations in amniotic fluid in Down's syndrome pregnancies.

There is still no answer to the question as to how PGH, which is secreted by the syncytiotrophoblast, penetrates into the foetal circulation and amniotic fluid. A question arises as to whether the synthesis of this hormone also takes place in foetal cells. Finally, the recognition of the role of PGH in a foetal organism requires further thorough studies on a larger scale.

Conclusion

Our research indicates both an important role for PGH in maintaining the proper IGF-1 pool, and a direct influence on placenta functions through the activation of compensation mechanisms, which stimulate IGF-1 synthesis. The role of PGH in postnatal life, and its concentrations, particularly in the first months of life, is yet to be clarified. In the mother's organism this hormone undergoes a complete degradation after birth, and its fate in neonate blood is unknown. The trace concentration of IGF-1 in umbilical cord blood serum of five neonates, of whom three weighed less than 2,000 g, is also unclear. Molecular studies in this area will be conducted in the future.

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